# A Genome-Wide RNAi Screen for Modifiers of Polyglutamine-Induced Neurotoxicity in Drosophila 



## Doctoral Thesis

In partial fulfilment of the requirements for the degree "Doctor rerum naturalium (Dr. rer. nat.)" in the Molecular Medicine Study Programme at the Georg-August University Göttingen

submitted by<br>Hannes Voßfeldt

born in
Zerbst/Anhalt, Germany

Für meine Familie

Im Gedenken an Nadine
Du fehlst.

IT MATTERS NOT HOW STRAIT THE GATE, How Charged with punishments the scroll,<br>I AM THE MASTER OF MY FATE:<br>I AM THE CAPTAIN OF MY SOUL.

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## List of Publications

Parts of this work have already been published with authorisation of Prof. Jörg B. Schulz, Head of the Department of Neurology, University Medical Centre of the RWTH Aachen University, on behalf of the thesis committee.

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## Table of Contents

List of Figures .....
List of Tables ..... XI
List of Abbreviations ..... XII
1 Abstract ..... 1
1 Zusammenfassung ..... 2
2 Introduction ..... 3
2.1 Overview: proteopathies and polyglutamine diseases .....  3
2.2 Pathogenic mechanisms of polyglutamine diseases .....  6
2.2.1 Cytotoxicity of polyglutamine structures .....  6
2.2.2 Molecular pathways to polyglutamine disease. ..... 10
2.3 Examples of polyglutamine diseases ..... 13
2.3.1 Huntington's Disease (HD) ..... 13
2.3.2 Spinocerebellar ataxias. ..... 14
2.3.2.1 Spinocerebellar ataxia type 1 (SCA1) ..... 15
2.3.2.2 Spinocerebellar ataxia type 2 (SCA2) ..... 16
2.3.2.3 Spinocerebellar ataxia type 3 (SCA3)/Machado-Joseph disease (MJD) ..... 18
2.4 Drosophila melanogaster as an animal model in research ..... 21
2.4.1 The UAS/GAL4 expression system ..... 21
2.4.2 RNA interference (RNAi) ..... 22
2.4.3 Rough eye phenotype (REP) ..... 24
2.4.4 Drosophila models of polyglutamine disease ..... 24
2.4.5 Previously implemented modifier studies ..... 26
3 Aim of the Study ..... 27
4 Material and Methods ..... 28
4.1 Chemicals, reagents and equipment ..... 28
4.2 Fly experiments. ..... 32
4.2.1 Transgenic flies and housing conditions. ..... 32
4.2.2 Mating procedures ..... 33
4.2.3 Evaluation of rough eye phenotype modification ..... 34
4.2.4 Documentation of eye phenotypes. ..... 35
4.2.5 Dissection and staining of eye imaginal discs. ..... 35
4.2.6 Longevity analysis ..... 35
4.2.7 Protein collection from fly heads ..... 36
4.2.8 Immunoblotting ..... 36
4.2.9 Filter Retardation Assay ..... 37
4.2.10 Histological and immunohistochemical staining of paraffin sections ..... 38
4.2.11 Immunohistochemical staining of cryo sections ..... 39
4.2.12 Semi-thin tangential sectioning of fly heads and photoreceptor quantification ..... 39
4.3 Cell culture experiments. ..... 40
4.3.1 Cell culture conditions and media ..... 40
4.3.2 Generation of stable shRNA-expressing cells. ..... 40
4.3.3 Plasmid transfection ..... 41
4.3.4 Protein collection from cell culture and immunoblotting. ..... 41
4.3.5 Cytochemistry ..... 42
5 Results. ..... 43
5.1 Characterisation of a SCA3 fly model for the modifier screen ..... 43
5.1.1 Phenotypes of the disease model flies ..... 43
5.1.2 Assessment of SCA3tr-Q78 protein expression and effects in the eye ..... 45
5.1.3 Evaluation of photoreceptor integrity ..... 48
5.2 Modifier screen for polyQ-induced neurotoxicity ..... 49
5.2.1 Screen for unspecific RNAi effects in control flies ..... 50
5.2.2 Primary screen for polyglutamine modifiers ..... 50
5.2.3 Specificity of RNAi effects for SCA3tr-Q78-induced neurotoxicity ..... 55
5.2.4 Evaluation of gene silencing by RNAi lines ..... 56
5.3 Impact of modifiers on polyQ toxicity and aggregation ..... 57
5.3.1 Evaluation of tissue integrity of SCA3tr-Q78-shRNA-coexpressing flies ..... 57
5.3.2 Filter retardation analysis of RNAi influence on polyQ aggregates. ..... 58
5.3.3 RNAi effects on polyQ inclusions in situ ..... 60
5.4 Summary of RNAi screen results ..... 61
5.5 Analysis of the effect of TRMT2A silencing on polyQ toxicity in Drosophila. ..... 62
5.5.1 Impact of TMRT2A silencing on polyglutamine-induced REPs ..... 62
5.5.2 Evaluation of photoreceptor integrity of polyQ flies with TRMT2A knockdown ..... 64
5.5.3 Assessment of adult-onset polyQ fly longevity ..... 65
5.5.4 Influence of CG3808 downregulation on aggregate formation in Drosophila. ..... 66
5.6 Impact of TRMT2A knockdown on polyQ toxicity in a mammalian system ..... 68
5.6.1 Generation of stable TRMT2A knockdown HEK cells. ..... 69
5.6.2 Transfection of stable TRMT2A knockdown cells with polyQ constructs ..... 70
5.6.3 Investigation of aggregation in polyQ-transfected knockdown cells. ..... 72
5.7 Attempts on revelation of the molecular mechanism of TMRT2A knockdown on polyQ proteins ..... 73
6 Discussion ..... 75
6.1 Characterisation of the utilised polyQ Drosophila model ..... 75
6.2 Modifiers of Ataxin-3-induced REP in Drosophila ..... 77
6.2.1 Comparison to related polyQ modifier screens. ..... 78
6.2.2 Chaperones as polyQ misfolding and aggregation modifiers ..... 80
6.2.3 Components of the UPS in polyQ pathogenesis ..... 81
6.2.4 PolyQ-induced neurotoxicity modifiers involved in transcriptional regulation ..... 82
6.2.5 Nuclear transport proteins are modifiers of polyQ toxicity ..... 83
6.2.6 Further remarks on polyQ toxicity modifiers and the RNAi screen ..... 83
6.3 Aggregation in SCA3tr-Q78-shRNA-coexpressing flies ..... 84
6.4 The role of TRMT2A in polyQ pathogenesis ..... 85
7 Summary and Concluding Remarks ..... 88
8 Bibliography ..... 89
Curriculum Vitae. ..... 104
Private Danksagungen ..... 106
Appendix ..... 107
I Additonal eye phenotypes ..... 107
II RNAi lines modifying SCA3tr-Q78-induced REP ..... 108
III Fly lines used for verification of RNAi ..... 135
IV List of screened RNAi lines obtained from the VDRC as human orthologue sublibrary ..... 138

## List of Figures

Figure 1. Exemplary overview of proteopathies and the respective disease subcategories.
Figure 2. Model of conformational change, oligomerisation and aggregation as underlying pathogenic mechanism for polyQ diseases.8

Figure 3. Pathogenic processes during the development of polyQ diseases. 12
Figure 4. Model of the UAS/GAL4 expression system. 22

Figure 5. Mechanism of RNAi with shRNA. 23

Figure 6. Phenotypes induced by GMR-mediated expression of different transgenes.
Figure 7. Biochemical detection of SCA3tr-Q78 protein levels and aggregation together with verification of SCA3tr-Q78 expression, aggregation and induced cell death in larval imaginal discs.

Figure 8. Histological and immunohistochemical analysis of utilised fly models.
Figure 9. Photoreceptors in semi-thin sections of SCA3 disease models.
Figure 10. Flow chart of the implemented screen to identify modifiers of SCA3-induced toxicity including subsequent analysis of primary screen candidates.

Figure 11. Modification of the SCA3tr-Q78-induced phenotype by enhancing and suppressing candidate RNAi lines.

Figure 12. Summary of the SCA3tr-Q78 modifier screen and overview of modifier categories.
Figure 13. Influence of selected shRNAs on tissue integrity of SCA3tr-Q78 fly head sections.
Figure 14. Analysis of SDS-insoluble SCA3tr-Q78 aggregate load with shRNA modifiers. 59

Figure 15. Influence of RNAi on microscopically detectable Ataxin-3 inclusions in situ.
Figure 16. Rescue of polyQ-induced REP by shRNA against CG3808.
Figure 17. Evaluation of photoreceptor integrity in polyQ flies with CG3808 RNAi.
Figure 18. Adult-onset model of SCA3tr-Q78 in Drosophila and extension of polyQ fly life time by CG3808 RNAi.66
Figure 19. Overview of anti-aggregation effects of CG3808 RNAi in different polyQ models and settings. ..... 67
Figure 20. Stable shRNA-mediated silencing of TRMT2A expression after viral transduction of HEK293 cells. ..... 70

Figure 21. Aggregation properties of normal and expanded Huntingtin in control and TRMT2A knockdown HEK cells.

Figure 22. Impact of TRMT2A knockdown on different SDS-insoluble Huntingtin aggregates.
Figure 23. Overlap between screens for genetic modifiers of polyQ-induced neurotoxicity or aggregation.
Figure 24. Putative mechanistic explanation of polyQ toxicity amelioration by TRMT2A knockdown.

## List of Tables

Table 1. Overview of polyglutamine diseases. ..... 5
Table 2. Chemicals and reagents. ..... 28
Table 3. Equipment. ..... 30
Table 4. Software and online resources. ..... 31
Table 5. Utilised Drosophila melanogaster strains. ..... 32
Table 6. Stocks utilised for screening approaches. ..... 34
Table 7. Antibodies utilised for Drosophila head and cell lysate immunoblotting and for immunohistochemical stainings. ..... 37
Table 8. Lentiviral clones and non-target strain utilised for TRMT2A silencing experiments in HEK293 cells. ..... 40
Table 9. List of candidates with viable progeny modifying Ataxin-3-induced REP in Drosophila. ..... 52

## List of Abbreviations

| Abbreviation | Denotation |
| :--- | :--- |
| ADCA | Autosomal dominant cerebellar ataxia |
| Ago2 | Argonaute2 |
| ALS | Amyotrophic lateral sclerosis |
| AO | Acridine orange |
| ATXN | Ataxin |
| BDNF | Brain -derived neurotrophic factor |
| CACNA1 $A$ | Calcium channel, voltage-dependent, P/Q type, alpha 1A subunit |
| CAA | Cytosine-adenine-adenine (trinucleotide coding for glutamine) |
| CAG | Cytosine-adenine-guanine (trinucleotide coding for glutamine) |
| cAMP | Cyclic adenosine monophosphate |
| CAT | Cytosine-adenine-thymine (trinucleotide coding for valine) |
| CG | Protein-coding gene (in Drosophila melanogaster) |
| CNS | Central nervous system |
| CBP | CREB-binding protein |
| CREB | cAMP responsive element-binding protein |
| DRPLA | Dentatorubral-pallidoluysian atrophy |
| dsRNA | Double-stranded RNA |
| eGFP | Enhanced green fluorescent protein |
| EP | Histone deacetylase |
| FTD | Enhancer/promoter |
| FRA | Frontotemporal dementia |
| GABA | Hass multiple reporter retardation assay |
| GMR | Huntingtin-associated protein 1 |
| HA | Histone acetyltransferase |
| HAP1 | Huntington's disease |
| HAT |  |
| HDAC |  |


| HDJ1 | Human DnaJ protein 1 |
| :---: | :---: |
| HEAT | Huntingtin, EF3, PP2 $\underline{\mathbf{A}}$, TOR1 $^{\text {T }}$ |
| HEK | Human embryonic kidney cells |
| HIP1 | Huntingtin-interacting protein 1 |
| HRP | Horseradish peroxidase |
| HSP | Heat shock protein |
| HTS | High-throughput screen |
| HTT/Htt | Huntingtin |
| IR | Inverted repeats |
| kDa | Kilodalton |
| Lys | Lysine |
| MF | Morphogenetic furrow |
| MJD | Machado-Joseph disease |
| miRNA | MicroRNA |
| MOI | Multiplicity of infection |
| mRNA | Messenger RNA |
| MSN | Medium spiny neuron |
| NII | Neuronal intranuclear inclusion |
| ORF | Open reading frame |
| polyQ | Polyglutamine |
| RBM17 | RNA-binding motif protein 17 |
| REP | Rough eye phenotype |
| RISC | RNA-induced silencing complex |
| RLC | RISC loading complex |
| RNAi | RNA interference |
| SBMA | Spinal bulbar muscular atrophy |
| SCA | Spinocerebellar ataxia |
| SEM | Scanning electron microscopy |
| shRNA | Short hairpin RNA |
| siRNA | Small interfering RNA |
| Sp1 | Specificity protein 1 |
| TBP | TATA box-binding protein |
| TDP-43 | TAR DNA-binding protein 43 |


| TPR2 | Tetratricopeptide repeat protein 2 |
| :--- | :--- |
| TRMT2A | tRNA methyltransferase homologue 2A |
| tRNA | Transfer RNA |
| UAS | Upstream activation sequence |
| UIM | Ubiquitin-interacting motif |
| UPS | Ubiquitin-proteasomal system |
| VDRC | Vienna Drosophila RNAi Centre |
| WT | Wild type |


#### Abstract

1 Abstract

Spinocerebellar ataxia type 3 (SCA3) or Machado-Joseph disease (MJD) belongs to the group of polyglutamine (polyQ) neurodegenerative diseases and is the most prevalent autosomal dominant cerebellar ataxia worldwide. A highly variable polyglutamine tract is thought to confer toxicity upon the otherwise unrelated proteins causing polyQ diseases. Apart from the polyQ extension, the physiological function and cellular context of these proteins and their interaction partners appear to be crucial for the specific pathogenesis and course of the disorders. In order to elucidate the molecular disease mechanisms triggered by trinucleotide repeats, we intended to identify genetic interactors enhancing or suppressing polyQ toxicity.

Therefore, expression of a human Ataxin-3-derived polyQ transgene was targeted to the Drosophila compound eye. The resulting photoreceptor degeneration induced a rough eye phenotype (REP) in adult flies. Eye-specific silencing of distinct genes (fly genes with a human orthologue, ca. 7,500 genes) by RNAi was utilised to identify genetic interactors of the REP. Changes in the observed REP are likely to originate from the knockdown of the RNAi target. Thus, silenced candidate genes are capable of modifying polyQ-induced neurotoxicity.

The gene products that were discovered in this manner represent various biological pathways and molecular functions. Secondary investigations were conducted with a set of candidate genes to gain more insight into the mode and quality of the interactions and revealed novel modifier genes involved for example in tRNA methylation or sphingolipid metabolism. These results are likely to shed further light on the molecular pathogenesis of MJD and other polyQ disorders together with the role of Ataxin-3 and its modulator proteins in this process.


## 1 Zusammenfassung

Die Spinozerebelläre Ataxie Typ 3 (SCA3) oder Machado-Joseph-Krankheit (MJD) gehört zur Gruppe der neurodegenerativen Polyglutaminerkrankungen (PolyQErkrankungen) und ist die häufigste autosomal-dominante zerebelläre Ataxie weltweit. Ein in der Länge hochvariabler Polyglutaminabschnitt ist vermutlich die Ursache für die Toxizität der ansonsten nicht verwandten Proteine, welche die PolyQ-Erkrankungen verursachen. Abgesehen von dem verlängerten Polyglutaminbereich scheinen die physiologische Funktion und der zelluläre Kontext dieser Proteine und ihrer Interaktionspartner entscheidend für die spezifische Pathogenese und den Krankheitsverlauf zu sein. Diese Arbeit soll dazu beitragen, genetische Interaktoren zu identifizieren, welche die PolyQ-Toxizität verstärken oder vermindern, um somit die molekularen Krankheitsmechanismen zu entschlüsseln, die durch die TrinukleotidWiederholungen ausgelöst werden.

Dafür wurde ein humanes, von Ataxin-3 abgeleitetes Transgen in den Facettenaugen von Drosophila exprimiert. Die daraus resultierende Degeneration der Photorezeptoren induziert einen Raue-Augen-Phänotyp (Rough Eye Phenotype, REP) in adulten Fliegen. Um genetische Modifikatoren des REP zu identifizieren, wurde die Expression bestimmter Gene (Fliegengene mit einem humanen Ortholog, insgesamt ca. 7.500) augenspezifisch per RNAi vermindert. Mögliche Veränderungen im beobachteten REP sind dann höchstwahrscheinlich auf den RNAi-vermittelten Knockdown der Genexpression zurückzuführen. Damit wären die stummgeschalteten Kandidatengene zur Modifizierung der PolyQ-induzierten Neurotoxizität fähig.

Die auf diese Weise identifizierten Genprodukte sind in verschiedene biologische Prozesse involviert und stehen stellvertretend für unterschiedlichste molekulare Funktionen. Für eine Auswahl von Kandidatengenen wurden zusätzliche Untersuchungen angestellt, um die Art und das Ausmaß der Interaktionen zu bestimmen. Dabei wurden neue Modifikatorengene analysiert, welche z. B. in die Methylierung von tRNA oder den Sphingolipid-Metabolismus involviert sind. Diese Ergebnisse können neue Erkenntnisse bei der Aufklärung der Pathogenese der MJD und anderer PolyQ-Erkrankungen hervorbringen und gleichzeitig zum Verständnis der Rolle von Ataxin-3 und seinen Modulatorproteinen beitragen.

## 2 Introduction

Neurodegenerative diseases affecting and impairing the central nervous system are on the rise throughout the world. Senescence being the main risk factor for these diseases, the number of age-related disorders is rising dramatically especially in industrial countries where life expectancy advances. Concomitantly, more and more inherited diseases of the nervous system can be precisely diagnosed and investigated, revealing underlying mechanisms and connections, but also posing new questions. In contrast to the scientific progress in this field as well as to the increasing burden of neurodegenerative diseases both to society and individuals stands the lack of efficient therapeutical options, let alone of a cure for the vast majority of these devastating and mostly fatal disorders.

### 2.1 Overview: proteopathies and polyglutamine diseases

Proteopathies are disorders in which abnormal accumulation of specific proteins represents the pathological hallmark of the respective diseases. Therefore it can be suggested that the altered proteins might cause the corresponding medical condition [5]. Nowadays, more than 40 groups of proteopathies are known, occurring through the mutation and putative misfolding of various proteins like hemoglobin, rhodopsin, fibrinogen, tau or amyloid $\beta$ peptide [6]. In tissues affected by a proteopathy, aggregates of the respective mutated protein can be detected. It is generally believed that these accumulations play a role in the pathogenesis, although it is not clear whether they are the actual toxic species.

Trinucleotide repeat disorders, also called triplet repeat expansion disorders, make up an own heterogeneous group in the entity of proteopathies. They are characterized by the expansion of a tract of trinucleotide repeats within the particular disease gene. Healthy individuals bear a distinct repeat range in the normal allele and only upon elongation of this nucleotide stretch above a certain threshold the gene product is rendered toxic [7-9]. The trinucleotide disorders can be grouped into two categories: the polyglutamine diseases and the non-polyglutamine diseases, the latter being caused by genes exhibiting repeats different from the CAG (coding for glutamine) triplets characteristic for polyglutamine diseases (Figure 1).

The group of polyglutamine (polyQ) diseases comprises nine heritable neurodegenerative disorders, including Huntington's disease (HD), spinal bulbar muscular atrophy (SBMA) and six
spinocerebellar ataxias (SCA). All nine arise from a gain-of-function mutation in their respective disease genes, resulting from an autosomal dominant (except for the X-linked SMBA) expansion of polyglutamine repeats [9-11]. Therefore they are also entitled polyglutamine expansion disorders
(Table 1).


Figure 1. Exemplary overview of proteopathies and the respective disease subcategories.
The entity of trinucleotide disorders is a subgroup of proteopathies together with other neurodegenerative diseases and comprises the polyglutamine and non-polyglutamine diseases. The members of the polyQ disease family are depicted entirely (see also Table 1), the listing of the other disease groups is not intended to be exhaustive.
SCA, spinocerebellar ataxia; SBMA, spinal bulbar muscular atrophy; HD, Huntington's disease; DRPLA, dentatorubral-pallidoluysian atrophy; FRAXA, fragile $X$ syndrome, FRAXE, fragile XE syndrome; FRDA, Friedreich ataxia; DM, myotonic dystrophy

Although the principle genetic basis of polyglutamine diseases has been known for 20 years, the molecular pathogenesis remains elusive and therapeutic approaches are merely aimed at the symptoms rather than the cause of the disorders [2].

Polyglutamine diseases have a remarkable genotype-phenotype correlation with most of the diseases emanating from an expansion above a threshold of 40 CAG repeats (Table 1). This origin of the disorders is regardless of the predicted functions of the causative genes or the surrounding amino acids of the polyQ stretch. The age of onset is inversely correlated to the length of the polyglutamine tract, whereas the severity increases with the number of trinucleotide repeats [12, 13].

Apart from the polyQ tract, the gene products share no homology to each other, suggesting a common pathogenic mechanism leading to the development of disease. Furthermore, the specificity for affecting certain brain regions in the diverse polyQ diseases cannot be explained by differential expression patterns of the disease genes. With regard to similar toxicity of heterogeneous proteins in different cellular and spatial settings, there is overwhelming
need for insights into polyQ protein-interacting genes in order to decipher the molecular processes leading to neurotoxicity.

Table 1. Overview of polyglutamine diseases.

| Disease | Gene product | Inheritance | Normal repeat length | Expanded repeat length | Distinguishing clinical features ${ }^{1}$ |
| :---: | :---: | :---: | :---: | :---: | :---: |
| HD | Huntingtin | AD | 6-34 | 36-121 | Chorea, dystonia, cognitive deficits, psychiatric problems |
| $\begin{aligned} & \text { SCA1 } \\ & \text { (ADCA) } \end{aligned}$ | Ataxin-1 | AD | 6-44 | 39-82 | Pyramidal signs, peripheral neuropathy |
| $\begin{aligned} & \text { SCA2 } \\ & \text { (ADCA) } \end{aligned}$ | Ataxin-2 | AD | 15-24 | 32-200 | Slow saccadic eye movements, peripheral neuropathy, decreased deep tendon reflexes, dementia |
| $\begin{aligned} & \text { SCA3 } \\ & \text { (ADCA) } \end{aligned}$ | Ataxin-3 | AD | 13-36 | 61-84 | Pyramidal and extrapyramidal signs, lid retraction, nystagmus, decreased saccade velocity, amyotrophy, fasciculations, sensory loss |
| $\begin{aligned} & \text { SCA6 } \\ & \text { (ADCA) } \end{aligned}$ | CACNA1A | AD | 4-19 | 10-33 | Sometimes episodic ataxia, very slow progression |
| $\begin{aligned} & \text { SCA7 } \\ & \text { (ADCA) } \end{aligned}$ | Ataxin-7 | AD | 4-35 | 37-306 | Visual loss with retinopathy |
| $\begin{aligned} & \text { SCA17 } \\ & \text { (ADCA) } \end{aligned}$ | TBP | AD | 25-42 | 47-63 | Mental deterioration, occasional chorea, dystonia, myoclonus, epilepsy |
| $\begin{aligned} & \text { DRPLA } \\ & \text { (ADCA) } \end{aligned}$ | Atrophin | AD | 7-34 | 49-88 | Chorea, seizures, dementia, myoclonus |
| SBMA | Androgen receptor | XR | 9-36 | 38-62 | Motor weakness, swallowing difficulties, gynecomastia, decreased fertility |

${ }^{1}$ all ADCAs have gait ataxia.
ADCA, autosomal-dominant cerebellar ataxia; AD, autosomal-dominant; XR, X-linked recessive; HD, Huntington's disease; SCA, spinocerebellar ataxia; DRPLA, dentatorubral-pallidoluysian atrophy; SBMA, spinal bulbar muscular atrophy; CACNA1 ${ }_{A}$, Calcium channel, voltage-dependent, P/Q type, alpha 1A subunit; TBP, TATA box-binding protein [8,14]

### 2.2 Pathogenic mechanisms of polyglutamine diseases

Despite the revelation of the connection with polyQ tract expansion within the respective proteins, the molecular mechanisms resulting in polyQ diseases are still under debate. It has been widely believed that aggregation of polyglutamine proteins, namely inclusions bodies, is the causative agent [15], nevertheless, research over the years has diversified this opinion [16] and has put focus on different species and structures of polyQ proteins (Figure 2). Additionally, it is crucial for the understanding of the disorders and for the development of therapeutical approaches to identify the very molecular pathways and cellular context by which toxicity of the proteins eventually leads to neuronal death.

### 2.2.1 Cytotoxicity of polyglutamine structures

## Polyglutamine monomers

The conformational change of the molecular structure of native polyglutamine proteins into $\beta$-sheet-rich monomeric proteins is an essential step in the toxification of these gene products [17, 18]. Due to the obstacles that arise while trying to observe the structure of these $\beta$-strands in the actual disease protein, most of the studies with this focus have been conducted utilising artificial proteins [19-21]. Numerous investigations proposed cylindrical, hairpin and intramolecular $\beta$-sheet models, however it is not clear which of these might be the predominant form in affected cells. It has been shown that polyQ monomers are cytotoxic in cultured cells [22]. Despite these findings it remains elusive whether toxicity is conferred directly by the monomers themselves or if the transition into oligomers is responsible for the cytotoxicity. It is noteworthy that the monomer-oligomer transition propagates rapidly throughout the cell and can also take place in the reverse direction [23].

## Polyglutamine oligomers

Oligomers of disease proteins have been proposed as the toxic species leading to cell death in a variety of neurodegenerative disorders, including Alzheimer's disease [24]. There are several lines of evidence for oligomeric intermolecular structures of expanded polyglutamine proteins. For instance an anti-parallel $\beta$-sheet structure with intermolecular hydrogen bonds called "polar zipper" [25], a parallel $\beta$-sheet conformation [18] or a cylindrical assembly designated "nanotube" [20], all analysed in vitro, have been described. Nevertheless, the predominance of any one of these species in living cells could not be verified. Studies investigating the formation of expanded polyQ oligomers out of monomers revealed a bidirectional transition of these species and the predominant cytotoxic potential of the oligomer fraction towards neuronal cells [23, 26, 27]. Furthermore, polyQ oligomers are more toxic than inclusion bodies [26] and heat shock proteins 40 and 70 (HSP40/70) are capable of ameliorating the deleterious effects of expanded polyQ proteins without influencing the formation of inclusions [28-30]. In a mouse model of SBMA, the presence of oligomers exhibited a close correlation to disease symptoms [31]. From these findings and other studies concerning polyQ oligomers [32-35], a pivotal role of these structures in polyQ pathogenesis can be deduced (toxic oligomer hypothesis). Some reports even favour a common toxic structure hypothesis [33,36] based upon the cross-reactivity of antibodies against $\mathrm{A} \beta$ oligomers with other amyloidogenic proteins (like $\alpha$-synuclein and polyglutamine proteins). Accordingly, amyloidogenic proteins causing neurodegenerative diseases would share a common toxic structure regardless of their amino acid sequence. However, these findings have not yet been verified in tissue of polyQ disease patients.

## Polyglutamine inclusions

The formation of intranuclear inclusion bodies composed of expanded polyQ proteins has for a long time been considered to be the toxic event underlying the pathogenesis of the respective disorders [37-41]. Apart from the polyQ gene products themselves, a variety of other proteins like ubiquitin and heat shock proteins have been shown to be present in nuclear inclusions. Deprivation of these proteins from other cell compartments may result in dysfunction of neuronal cells [37, 42] concomitantly with disruption of axonal transport and nuclear function [43]. Despite these findings, results of more recent studies have established a rather cell-protective role of polyQ inclusion bodies. In addition to the lacking
correlation between inclusion body formation on the one hand and cellular imbalance and death on the other [44, 45], polyQ inclusion bodies proved to be beneficial in rat striatal neurons exposed to mutant Huntingtin (Htt) [46]. Furthermore, cells with inclusions survived significantly longer than those with soluble oligomers [23]. Although this hypothesis is not yet fully verified in vivo, formation of polyQ inclusions appears to mitigate detrimental effects of the mutated proteins rather than being the initial molecular step of polyQ disease emergence.


Figure 2. Model of conformational change, oligomerisation and aggregation as underlying pathogenic mechanism for polyQ diseases.
Poly $Q$ pathogenesis requires an expanded poly $Q$ tract in the disease protein and a cellular environment promoting the accumulation of conformationally altered polyQ monomers. Cytotoxic effects are exerted in the course of oligomerisation of aggregate precursors and the formation of different aggregation states and species with varying impact on cellular dysfunction. Subsequent cellular impairment renders the environment even more aggregationprone. Eventually, the toxic effects exceed the cell's coping capability and lead to death of the dysfunctional cell and to disease onset.
Adapted from [1, 2].

## Influence of residues adjacent to the polyglutamine tract

Although the expansion of the polyQ stretch in disease proteins is the molecular basis of cytotoxiciy and pathogenicity in polyQ diseases, it does not explain the selectivity for distinct neuronal populations and tissues in the respective disorders. The different disease proteins exhibit a widespread distribution throughout the central nervous system (CNS) and are not confined to the especially vulnerable cell types. For instance, Huntington's disease mainly affects striatal GABAergic medium spiny neurons (MSNs) [47] whereas Ataxin-1 in SCA1 is most detrimental in Purkinje cells of the cerebellum [48]. In contrast, toxicity of Ataxin-3 in SCA3 affects a wide range of cell types in pons, substantia nigra, thalamus and diverse brain stem nuclei [49, 50]. An explanation for this discrepancy may be found in the disease protein portions apart from the polyQ stretch. Mutation in the CAG tract may also alter the protein-protein interactions of the non-polyQ parts of the protein. The association of mutated Htt for instance is more tightly with Htt-associated protein 1 (HAP1) and less strong with Htt-interacting protein 1 (HIP1) compared to wild-type Htt [51]. The modified interaction properties lead to the disruption of axonal transport of brain-derived neurotrophic factor (BDNF) and disturbances of clathrin-mediated endocytosis respectively. The correlation of Ataxin-1 mutation and Purkinje cell demise probably arises from a complex the disease protein forms with the neurotoxic RNA-binding motif protein 17 (RBM17). RBM17 is highly expressed in Purkinje cells and opposes another interactor of Ataxin-1, the neuroprotective Capicua [52]. Mutation of Ataxin-1 shifts the interaction balance towards a stronger association with RBM17 and results in cerebellar cell loss [53, 54].

Posttranslational modifications of amino acid residues outside the polyQ stretch have a remarkable impact on the toxicity of the disease proteins by influencing proteinprotein interactions as well as by determining processing of the respective gene products. For example, phosphorylation of distinct amino acids of Htt, Ataxin-1 and the androgen receptor (AR) alters the affinity properties to ligands [55] and is capable of either reducing [56, 57] or increasing [58] the formation of inclusion bodies and toxicity.

Ubiquitination of polyQ-containing proteins subjects them to degradation by the ubiquitin-proteasomal system (UPS) and therefore represents a toxicity-ameliorating mechanism. On the contrary, the competing sumoylation renders the proteins more stable and promotes cell death via aberrant transcription and an increase in the amount of toxic oligomers [59, 60]. Selective expression of cofactors influencing posttranslational
modifications of polyQ proteins adds to the specificity of toxicity to certain cell populations [61].

According to the toxic fragment hypothesis, proteolytical processing of polyQ proteins is the initial step in rendering them toxic, leading to an increase in aggregation and to nuclear translocation [62]. Htt, Ataxin-3 and AR have all been described to be susceptible to cleavage by caspases at specific amino acid sites [31, 63-65]. Mutation or phosphorylation of these sites is sufficient to decrease inclusion body formation as a result of reduced proteolytical cleavage and hence toxicity [66, 67].

### 2.2.2 Molecular pathways to polyglutamine disease

## Transcriptional dysregulation

The nuclear translocation and accumulation of expanded and proteolytically processed polyQ proteins suggests hampering of regular transcription in neuronal cells via altered interactions with transcriptional factors and cofactors. Several nuclear transcriptional regulators like CREB-binding protein (CBP), TAFII130, Sp1 and p53 have been shown to interact with polyQ proteins and are recruited to nuclear inclusions [40, 68, 69]. Microarray-based experiments with HD and DRPL mouse models exhibited similar alterations in gene expression [70]. Due to the pivotal role of histone acetylation for gene transcription, aberrant interactions of mutant polyQ proteins with histone acetyltransferases (HAT) influence gene expression as shown for Htt and CBP [71]. HAT activators have also been proposed as a therapeutic strategy in neurodegenerative diseases [72], the same applies to inhibitors of histone deacetylases (HDAC) [73, 74]. For the latter, improvements of polyQ-induced phenotypes in mouse and Drosophila models could be shown [75, 76]. Remarkably, the SCA3 causative protein, Ataxin-3, is a transcriptional repressor in its native state, involved in chromatin binding and histone deacetylation via HDAC3. Mutated Ataxin-3 loses its repressor function, leading to increased histone acetylation in cultured cells and the pons of SCA3 patients [77].

## Impairment of the ubiquitin-proteasomal system (UPS)

The ubiquitin-proteasomal system is responsible for clearance and degradation of defective, aged and misfolded proteins in the cell. As polyQ inclusion bodies are ubiquitinpositive and components of the proteasome are recruited to these accumulations, studies suggest an impairment of proper UPS function in polyQ disease as a trigger for neuronal
cell death $[78,79]$. This hypothesis is supported by the fact that cells with inclusion bodies exhibit decreased UPS activity [80]. Accordingly, mice and patients with polyQ disease present with global dysfunction of the UPS [81]. There is evidence that eukaryotic proteasomes are not capable of properly degrading polyQ sequences of the respective proteins, subsequently leading to proteasomal blockage [82]. Moreover, aberrant forms of ubiquitin have been shown to enhance aggregation [83].

In contrast, no malfunction of the UPS has been described for mouse models of HD and SCA7 [84-86]. Additionally, the reasoning of proteasomal component sequestration leading to increased cell death contradicts the rather non-pathogenic role of polyQ inclusion bodies.

It is noteworthy that the causative protein for SCA3, Ataxin-3, is the first deubiquitinating enzyme known whose catalytic activity is modulated by ubiquitination itself, enhancing its activity in cleaving Lysin63 linkages in ubiquitin chains [87] and thereby also modulating protein quality control via the UPS per se.

## Impairment of mitochondrial function

Especially in Huntington's disease, evidence for an involvement of mitochondrial dysfunction during disease pathogenesis is established [88, 89]. Reports show signs of impairment of mitochondrial function such as decreased glucose metabolism and mitochondrial complex activity in HD patients [90] as well as lower membrane potentials in HD mice and patients compared to controls [91]. Transcriptional repression of PGC-1 $\alpha$ (a transcriptional coactivator of genes involved in energy metabolism) by mutant Huntingtin results in dysregulation of mitochondrial function and eventually in neuronal cell death [92]. These findings render mitochondrial impairment a side effect of transcriptional derangement in polyQ diseases. Huntingtin has also been implicated in the fission-fusion balance of mitochondria. Here, mutant Htt promotes mitochondrial fragmentation in vitro and in vivo, preceding the onset of Huntingtin aggregates and neurological deficits. In consequence, defects in anterograde and retrograde mitochondrial transport lead to neuronal cell death [93]. Antioxidants such as coenzyme Q10 and mitochondrial stability enhancers proved to be beneficial for the motor functions of HD mice [94-96], however, positive effects for other polyQ diseases cannot be deduced from these results.

## Impairment of axonal transport

Huntingtin seems to play a role in axonal transport as a lack of normal protein levels in Drosophila neurons disrupts this process crucial for mobility of mitochondria, mRNA and proteins and thus survival of the neurons [97, 98]. Furthermore, polyQ length correlates with inhibition of anterograde and retrograde axonal transport by mutant Htt and AR [9799]. In addition, expanded polyQ proteins and the resulting aggregates or inclusion bodies themselves are capable of blocking axonal transport in disease models, triggering neurotoxicity [100-102].


Figure 3. Pathogenic processes during the development of polyQ diseases.
Disease genes with an expanded CAG trinucleotide tract are transcribed and the mRNA is translated into a fulllength protein with an elongated polyQ stretch. The mutant full-length protein itself already adopts novel interactions with other proteins and is furthermore proteolytically cleaved to a truncated form. These processed polyQ protein may alter ion transport into the cell and are prone to aggregation, thereby forming cytoplasmic aggregates and intranuclear inclusions upon transport into the nucleus. Toxic truncated polyQ proteins are a target for proteasomal degradation (intranuclear inclusions are ubiquitinated) and retained in a native conformation by chaperones if possible (not depicted). The alterations or impairment of the processes above are all presumptively capable of resulting in cellular dysfunction and eventually cell death.
Impairment of mitochondrial function and axonal transport are not shown.
Adapted from [4].

### 2.3 Examples of polyglutamine diseases

### 2.3.1 Huntington's Disease (HD)

## Epidemiology and clinical features

Huntington's disease is the most common polyQ disease with a prevalence of 4-10 cases per 100,000 people in the Western world and many more at risk. The mean age of onset of HD is 40 years [47]. Clinically, extrapyramidal motor signs like chorea (usually the first motor symptom in adults), bradykinesia and dystonia together with features like progressive motor dysfunction, cognitive decline and psychiatric disturbance hint to the diagnosis Huntington's disease. Caudate and cortex are the brain regions most affected by atrophy diagnosed via neuroimaging. Additionally, the caudate and also the putamen present atrophy in neuropathology [103]. GABAergic medium-sized spiny striatal neurons are the cells most vulnerable to the detrimental effects of mutated Huntingtin [104, 105]. Secondary to the loss of striatopallidal projection fibres is atrophy of the globus pallidus, together with common cerebral cortical cell loss. Death occurs inevitably 10-20 years after emergence of the disease. Patients usually decease from bulbar dysfunction and complications like pneumonia or heart failure [47, 106].

## Molecular genetics and pathology

The molecularpathological hallmark of Huntington's disease is an expansion of a highly variable and unstable CAG repeat tract at the $N$-terminus (exon 1 ) of the disease gene huntingtin (HTT) [47, 107]. The gene itself is located on the short arm of chromosome 4 at position 16.3 [107]. The repetitive trinucleotide stretch within HTT has a length of 6-34 repeats in the normal population. After crossing a threshold repeat length of about 36 , the overlong polyQ tract of the translated gene product renders the protein toxic, with a reduced penetrance in counts of 36-39 [108]. The longest ever reported repeat length amounts to about 250 glutamines [109].

The age of onset of HD is inversely correlated with the polyQ tract length. A juvenile form of HD originates from a glutamine repeat count of 70 and more. The gene product Huntingtin is a very large protein with a molecular weight of 348 kDa and can be detected in several tissues, but especially in the brain, from early embryogenesis on [110-112]. Huntingtin has been proposed to act as a scaffolding protein due to its multiple HEAT
repeats [113] and the large number of interacting proteins revealed in a yeast two-hybrid screen [114]. The protein seems to be crucial during embryogenesis as mice lacking the functional gene are lethal [115]. Furthermore, Htt has an influence on the expression of brain-derived neurotrophic factor (BDNF) via unknown mechanisms [116]. Several studies suggest an association of Huntingtin with vesicles and microbtubules, indicating a role in cytoskeletal anchoring and transport of mitochondria [93, 117, 118].

### 2.3.2 Spinocerebellar ataxias

The spinocerebellar ataxias and the more complex dentatorubral-pallidoluysian atrophy belong to the group of autosomal-dominant cerebellar ataxias (ADCAs) which one to three among 100,000 Europeans suffer from. Among these disorders there are seven polyQ diseases (SCA1-3, SCA6-7, SCA17, DRPLA), the most frequent of which will be addressed here $[14,119]$. When using the term SCAs in the following text, it will refer only to these seven polyQ-related ones, leaving out the other 25 spinocerebellar ataxias and certain episodic ataxias unless pointed out otherwise. The group of spinocerebellar ataxias (SCAs) is a growing entity of disorders sharing many clinical and pathological features. Neurodegeneration in these disorders mainly affects the cerebellum and its afferent and efferent connections. Due to this classification, dentatorubral-pallidoluysian atrophy (DRPLA) can also be grouped into this disease category, although not being an actual SCA [4]. The disambiguation of the single spinocerebellar ataxias from each other is almost impossible if only the clinical manifestation and neuroimaging are being considered. Juvenile occurrence of SCAs has been observed, as well as late-onset forms; nevertheless the typical manifestation is in middle-aged patients. After disease onset, the SCAs progress to premature death after 10-20 years. Differential severity and age of onset can be explained by the highly variable number of expanded glutamine repeats, leading to a more severe disease course at high repeat numbers and being inversely correlated with the age at disease initiation. In this context and like in other polyQ diseases, the phenomenon known as anticipation plays an important role. This term describes the increase in CAG repeat number in successive generations, rendering the disorder more severe in the descendants of an affected, specifically male individual [120-122].

Statements about the epidemiology of SCAs are rather hard to make due to only few and mostly regionally restricted data on prevalence and incidence. The heterogeneous
presentation of the diseases also leads to significant variations in ethnic and continental populations which are even more enhanced by founder effects (reviewed in [4]).

### 2.3.2.1 Spinocerebellar ataxia type 1 (SCA1)

The disease gene for spinocerebellar ataxia type 1, namely Ataxin-1, was the first ataxia gene to be discovered with an unstable trinucleotide repeat stretch in the line of various other genes responsible for SCAs [123]. SCA1 is ranked third in prevalence among the polyQ ataxia subtypes. The disease makes up for 6-8 \% of the worldwide ADCA cases and is the most common SCA in South Africa and Italy [119, 124, 125].

## Clinical features

SCA1 usually presents when the individual affected is in his or her forties, although juvenile and late onset forms have been reported. Clinical signs for SCA1 are highly variable which makes the disease hard to distinguish from the other spinocerebellar ataxias [126]. Symptoms include ataxia of the gait and stance, spasticity together with dysarthria, oculomotor abnormalitites and pyramidal signs [127]. Differentiation of SCA1 from the other SCAs is possible by investigating central motor pathways with motor-evoked potentials in which the conduction time is remarkably longer than in SCA2, SCA3 and SCA6 [128].

## Molecular genetics and pathology

Spinocerebellar ataxia type 1 is caused by an abnormal CAG trinucleotide repeat expansion in the open reading frame (ORF) of the ATXN1 gene located on the short arm of chromosome 6. It is expressed in a variety of different tissues [129], however, the exact functions of the gene product Ataxin-1 at its nuclear localisation are not known. No phenotypes resembling those of SCA1 patients have been found in ATXN1 knockout mice, speaking against a loss-of-function of the protein as disease origin [130]. Normal alleles bear a repeat number of 6-44 CAGs. In the range of 36 to 44 CAG repeats are considered non-pathogenic if they are interrupted by one to three CAT trinucleotides [129, 131]. Alleles carrying 36-38 CAG repeats without CAT interruptions, called mutable normal or intermediate alleles, are unlikely to be symptomatic but have a high chance to elongate during inheritance to progeny. There are reports of seldom reduced penetrance of
expanded CAG repeat alleles with CAT interruption [131], nevertheless, full penetrance and pathogenicity of ATXN1 starts with uninterrupted 39 CAG repeats [132]. As in all polyQ diseases, anticipation and the rule of a longer, uninterrupted CAG repeat stretch causing a more severe course of the disease and an earlier age of onset apply [133, 134].

Like in the other polyQ diseases, the elongated polyQ stretch in Ataxin-1 is believed to confer abnormal folding properties onto the protein, rendering it prone to selfaggregation and accumulation in nuclear inclusion bodies (NIs). In these insoluble aggregates components of the protein degradation machinery such as chaperones or heat shock proteins (HSPs) and proteasomal constituents together with ubiquitin have been detected. These findings suggest the aggregates to be interfering with the cell's protein clearance mechanisms, consequently leading to SCA1 pathogenesis [135-138]. Studies furthermore proved the dependency of Ataxin-1 on phosphorylation and interaction with various proteins for aggregation and toxicity [58, 139].

Pathologically, SCA1 is characterised by atrophy of the brain stem and the cerebellum, where demise of especially Purkinje cells is observed [48].

### 2.3.2.2 Spinocerebellar ataxia type 2 (SCA2)

SCA2 is the second most prevalent autosomal dominant ataxia worldwide (15 \% of all ADCA families). There is a particularly higher number of cases in Italy [125], India [140] and especially Cuba (Holguín province) [141-143].

## Clinical features

The clinical manifestations of SCA2 differ from those of other SCAs insofar as that deep tendon reflexes present decreased and that there is saccadic slowing which is the most outstanding symptom in comparison to the resembling disorders SCA1 and 3 [144]. Patients show pyramidal findings and sometimes parkinsonism [145]. Other symptoms include cerebellar dysfunction in all SCA2 patients and peripheral neuropathy with varying frequency [146, 147]. SCA2 may also present as pure familial parkinsonism without cerebellar signs which is responsive to L-dopa treatment, but only affects a few patients with a smaller number of CAG repeats [148]. Disease onset is usually in the fourth life decade, afterwards progressing for approximately 10 to 15 years until premature death [149].

## Molecular genetics and pathology

The underlying cause for SCA2 is the instability of the CAG trinucleotide tract in the gene ATXN2 coding for Ataxin-2. ATXN2 alleles containing 31 or fewer CAG repeats are considered non-pathological. Repeat numbers exceeding this threshold are causative for SCA2, with 32 and 33 CAG repeats resulting in late onset SCA2 after the age of 50 years. The expanded CAG allele may be interrupted by a CAA trinucleotide, increasing the meiotic stability of the repeat [150], although not influencing the pathogenicity since it codes for glutamine as well [151-153].
The protein has a cytoplasmic localization in normal as well as in SCA2 brains where it associates with Golgi membranes [154]. There is no difference in the expression pattern of SCA2-affected and non-affected individuals, additionally, aggregates of Ataxin-2 exhibit neither ubiquitination nor nuclear translocation [155].

The interaction of Ataxin-2 with the RNA-recognition motif-containing Ataxin-2 binding protein 1 implies an involvement of Ataxin-2 in mRNA translation or transport [156]. Despite this, Ataxin-2-deficient mice do not show marked neurodegeneration, however, they present with decreased fertility, obesity and altered hippocampal plasticity [157, 158]. Recent studies presented evidence for an association of intermediate-length polyQ expansions (27-33Q) in Ataxin-2 with amyotrophic lateral sclerosis (ALS). This influence is thought to be mediated by the RNA-dependent interaction of Ataxin-2 with one of the putative ALS causative proteins, namely TDP-43 [159].

Neuropathologically, SCA2 post-mortem brains show a significant reduction of cerebellar Purkinje and granule cells, whereas other cerebellar nuclei are greatly spared. Furthermore, the inferior olive and the pontocerebellar nuclei in the brain stem together with the substantia nigra show neuronal loss. Spinal cords are demyelinated in the posterior columns and degenerated thalami and reticulotegmental nuclei of the pons have been reported, but not all of these findings were consistent in all patients [160-163]. One study also revealed involvement of the cerebral cortex, presenting with gyral atrophy especially in the frontotemporal lobes and atrophic as well as gliotic white matter [160].

### 2.3.2.3 Spinocerebellar ataxia type 3 (SCA3)/Machado-Joseph disease (MJD)

SCA3 is the most frequent among the SCA subtypes in most populations, comprising about $21 \%$ of the worldwide cases of autosomal-dominant cerebellar ataxias, however, there are considerable regional variations of prevalence [14, 119]. SCA3 is also known as Machado-Joseph disease (MJD) after a family of Azorean immigrants to the US in which the disease was first diagnosed [164]. A similar founder effect is believed to have resulted in the high prevalence of SCA3 cases for example in Brazil.

## Clinical features

SCA3 has one of the most heterogeneous clinical phenotypes of all cerebellar ataxias [165]. It includes progressive cerebellar ataxia and pyramidal signs associated to a variable degree with a dystonic-rigid extrapyramidal syndrome or peripheral neuropathy [166168]. These symptoms may or may not be accompanied by progressive external ophthalmoplegia, pseudoexophthalmus due to lid retraction [167], familial parkinsonism [169] and restless-legs syndrome [170, 171]. A rather specific sign of SCA3 is impaired temperature discrimination in limbs, trunk and face [172]. Based on the phenotypic variability arising from the combination of different clinical signs in family members, SCA3 has been classified into several subtypes, illustrating the extreme clinical heterogeneity [173, 174].
$\diamond$ Type I disease (13 \% of patients, dystonic-rigid form): early age of onset combined with spasticity, rigidity, bradykinesia and often little ataxia, presumably caused by a longer disease-associated repeat allele (mean 80).
$\diamond$ Type II disease (57 \%, ataxia with pyramidal signs): presents with ataxia and upper motor neurons signs, also spastic paraplegia is possible. This disease type correlates to a wide range of intermediate length disease-causing repeat alleles (mean 76).
$\diamond$ Type III disease ( $30 \%$, with peripheral amyotrophy): has the latest age of onset with ataxia and peripheral neuropathy, linked to shorter disease-causing repeat alleles (mean 73) [175].

Comparable to the heterogeneity of symptoms is the variability in age of onset of SCA3 which is commonly between the second and the fifth life decade with a mean of 37 years
[165]. Again, there is inverse correlation of age of symptom onset and length of the CAG repeat in the disease gene. Due to the multitude of debilitating clinical symptoms, SCA3 patients are increasingly dependent on external help as the disease progresses. After onset of brain stem signs like facial atrophy and dysphagia, eventually death occurs from pulmonary complications and cachexia from six to 29 years after onset (recent studies show a 21-year mean survival time) [176, 177].

## Molecular genetics and pathology

The disease gene responsible for Machado-Joseph disease when mutated, ATXN3 (also called MJD1), was mapped to the long arm of chromosome 14 [178]. The CAG trinucleotide repeat coding for polyglutamine is located in exon 10 of the gene [178, 179]. Fifty-six alternative splicing variants for ATXN3 have been described, of which at least 20 are translated into different protein isoforms [180]. Non-pathogenic alleles with variations of the CAG repeat in normal individuals can range from 12 to 43 repeats [175, 181-185]. A bimodal pattern of distribution of the normal allele frequency with peaks at 14 and 21-23 repeats has been shown for SCA3 patients [186, 187]. Furthermore, different nucleotides flanking the CAG sequence seem to correlate with specific repeat numbers and hence influence the stability of the polyQ stretch [187, 188]. CAG repeat numbers expanded above the normal length in pathogenic alleles are the cause of Machado-Joseph disease [145, 172, 178, 181-185, 189]. Trinucleotide repeat numbers ranging from 52 to 86 have been found in SCA3 patients. Alleles with seldom intermediate repeat numbers of 45 to 51 CAGs may exhibit reduced penetrance. As in other polyQ diseases, somatic and gametic instability is common in alleles with a prolonged CAG tract. This may result in spermatozoa having larger repeat counts than somatic cells and in cerebellar tissues with shorter repeat tracts than other brain regions [190, 191]. Anticipation has been described for SCA3, preferentially via paternal transmission [182, 192, 193].

The wild-type gene product Ataxin-3 encoded by ATXN3 is a highly conserved and ubiquitously expressed 42 kDa protein [194]. It is predominantly located in the cytoplasm but also capable of nuclear shuttling [194]. Ataxin-3 has been found to be a deubiquitinating protease [195-200] via a globular amino-terminal Josephin domain [201] and three ubiquitin-interacting motifs (UIMs) contained in the flexible carboxy-terminal tail [202]. The UIMs flank the polyQ tract, however, it is not known whether or how pathological expansion influences the enzymatic activity of the protein. As already
mentioned, ubiquitination of Ataxin-3 regulates its ubiquitin chain-editing function [87]. The Josephin domain has also been shown to interact with the Huntingtin-associated protein 1 (HAP1) [203], together with the polyQ domain it determines stability and aggregation of Ataxin-3 [204-208]. Overall findings propose a role of Ataxin-3 in cellular protein quality control, supported by suppression of polyQ-induced neurodegeneration in Drosophila [209], the regulation of aggresome formation [210], protein degradation and enzymatic activity [211].

Most data suggest a toxification mechanism for Ataxin-3 with an expanded polyQ tract, rendering it prone to misfolding and aggregation [212]. As for other polyQ proteins, this process has been experimentally proven by various studies in vitro as well as in vivo [78, 213-219]. There is no difference in the expression patterns of the normal and the mutated form of Ataxin-3 in brains and unaffected tissue of SCA3 patients [78].

In contrast to the predominantly cytoplasmic distribution of the native protein, mutated Ataxin-3 tends to accumulate in the nucleus of affected neurons, forming neuronal intranuclear inclusions (NIIs) in various brain regions [78]. NIIs can also be accompanied by axonal inclusions [220] and appear ubiquitinated and in association with heat shock proteins (HSP70 and 90, HDJ-2) and proteasomal subunits ( 20 S proteasome core, 11 S and 19 S regulatory caps of 26 S proteaseome) [79, 221, 222]. It is currently under heavy debate whether these inclusion bodies are the actual pathogenic species of mutated Ataxin-3 and other polyQ proteins or, on the contrary, merely are a safe storage for misfolded proteins to shield the cell from their toxicity $[212,223]$.

In accordance with the toxic fragment hypothesis, cleavage of Ataxin-3 to form shorter, polyQ-containing polypeptides seems to greatly enhance pathogenesis compared to the full-length protein, as has been shown in transgenic mice and flies. Moreover, these cleavage fragments appear to be the accumulating species in affected cells, which eventually undergo apoptosis [62, 65, 209, 219, 224, 225]. There are also alternative approaches to explain the aetiology of the disease apart from aggregation, including frameshifting during translation leading to deleterious polyalanine tracts [226], and RNA toxicity [227].

### 2.4 Drosophila melanogaster as an animal model in research

The fruit fly Drosophila melanogaster has been proven to constitute an excellent model organism for scientific research for more than a century now (reviewed in [228]). Since roughly $75 \%$ of the known disease-associated genes in humans also have orthologues in flies (annotated genome with roughly 16,000 protein-coding genes [229]), it might be reasonable to draw conclusions from investigations on molecular mechanisms in the fly to those in humans. Drosophila melanogaster was one of the first multicellular organisms whose genome has been sequenced completely and the corresponding genetic knowledge is well-established. Creation of transgenic animals allows for the modelling of human diseases by expressing toxic gene products. Besides these rationales, Drosophila also conjoins additional advantageous properties especially for high-throughput approaches. Due to the fast replication cycle and the high number of offspring, experiments can be conducted within short time periods with a reasonable number of individuals, allowing for drug and genetic modifier screening [230]. Although being an invertebrate organism, experimental findings are gained from an in vivo situation and conclusions about molecular mechanisms in higher animals can be drawn without raising ethical issues. Last but not least, several powerful genetic tools have been introduced in the past years in order to render research with the fruit fly even more feasible, precise and easy to handle. Some of these tools and a number of respective models and studies for polyQ disease are reviewed below.

### 2.4.1 The UAS/GAL4 expression system

The bipartite UAS/GAL4 ectopic expression system is frequently used in Drosophila as a means of overexpression of transgenes [231-233]. It makes use of the yeast transcriptional activator GAL4. Enhancer trap constructs (designed to facilitate GAL4 expression) were randomly inserted into the fly genome. If the insertion took place in the vicinity of an endogenous gene, GAL4 expression might mimic the expression pattern of this particular gene. To date there are plenty of so-called GAL4 driver lines available, mediating GAL4 expression in virtually every tissue at different time points throughout fly development. The gene of interest is introduced into a different fly line and put under the control of a GAL4 target, the upstream activation sequences (UAS). Upon crossbreeding of
these two fly lines, both moieties of the system are conjoined. GAL4 is produced under the control of the endogenous enhancer and able to bind to the UAS flanking the previously silent transgene of interest. Thus, expression of the gene of interest is enabled and directed in a spatiotemporal manner in the offspring (Figure 4). This renders the UAS/GAL4 system a valuable tool in fly genetics, although caution has to be taken since high GAL4 expression levels can have detrimental effects during development [234].


Figure 4. Model of the UAS/GAL4 expression system.
A tissue-specific endogenous enhancer binds to the promoter (grey) of the enhancer trap construct, thereby enabling gal4 expression (red) in the driver fly line. Association of GAL4 with the upstream activation sequence (UAS) of the fly line transgenic for the gene of interest activates expression (green). The bipartite nature of the system allows for tissue specificity and temporal restriction of activation of the gene of interest.

### 2.4.2 RNA interference (RNAi)

The gene silencing effect induced by double stranded RNA (dsRNA) termed RNA interference (RNAi) was fully established after experiments in Caenorhabditis elegans in 1998 [235]. It was the final step in a series of fundamental findings in plants [236, 237] as well as animals [238, 239]. Originally an endogenous mechanism involved in translational repression [240], development [241] and defence against parasitic genes [242], RNAi quickly evolved to be a powerful technique in scientific research, e.g. in mimicking knockout experiments without the extensive work effort of creating classical knockout animals. The effectors of RNAi are diverse small interfering RNAs (siRNAs) categorised according to their origin, biogenesis, mode of action and size [243, 244]. The source of siRNAs used in this work are transgenes coding for short hairpin RNAs (shRNAs). These transgenes consist of 100-400 base pairs present as an inverted repeat (IR) separated by miscellaneous nucleotides. Following expression of the IR, it will form a short hairpin RNA
(shRNA) which is exported from the nucleus to the cytoplasm. The shRNA is bound and cleaved by the ribonuclease protein Dicer-2, resulting in a double stranded structure without loop and RNA tails [245]. This small interfering RNA (siRNA) is then bound and translocated to the RNA-induced silencing complex (RISC) by the RISC loading complex (RLC) protein R2D2 which additionally discriminates between guide and passenger strands of the siRNA [246, 247]. The RLC recruits Argonaute2 (Ago2) and transfers the dsRNA to it, resulting in decay of the passenger strand by this endonuclease [248]. Subsequent to the release of the passenger strand and disassembly of R2D2, the active RISC is formed. The complex is capable of recognising and binding the messenger RNA (mRNA) target by base pairing with the guide strand. Eventually, this leads to cleavage of the mRNA and effectively to silencing of gene expression (Figure 5) [249].

Based upon this mechanism, Dietzl et al. were the first to establish a Drosophila RNAi library covering $\sim 90 \%$ of the entire fly genome. It utilises the conditional UAS/GAL4 expression system for induction of shRNAs under UAS control, leading to RNAi for the respective gene upon crossbreeding with a GAL4 driver line [250].


Figure 5. Mechanism of RNAi with shRNA.
The inverted repeats of the shRNA transgene are transcribed and the RNA is assembled into an shRNA. Following export from the nucleus, the shRNA is processed into double-stranded siRNA by Dcr-2.R2D2 forms the RLC together with the siRNA and discriminates the guide and the passenger strand, the latter is degraded upon binding of the RLC to Ago2. The guide strand and Ago2 form the RISC, eventually binding to and cleaving the target mRNA. Partially adapted from Dan Cojocari, Dept. of Medical Biophysics, University of Toronto, 2010.

### 2.4.3 Rough eye phenotype (REP)

The Drosophila compound eye is a highly ordered structure made up of about 800 single unit eyes termed ommatidia. Each ommatidium consists of eight photoreceptor cells arranged in an asymmetric trapezoid pattern accompanied by cone and pigment cells [251]. Cellular dysfunction and cell death as well as perturbation of crucial developmental pathways during compound eye formation lead to disturbances in this exact lattice and a so-called rough eye phenotype (REP). Consequently, a REP can be induced by the overexpression of toxic gene products. Expression of a disease gene can be targeted to postmitotic cells, including photoreceptor neurons, of the compound eye with the driver line glass multiple reporter (GMR)-GAL4 in combination with the UAS [252]. Glass expression starts at day one of larval stage L3 in all cells posterior of the morphogenetic furrow of the eye disc [253] as well as in a minor population of cells in the brain. The severity of the REP is directly correlated to the loss of underlying photoreceptor neurons reflected by vacant, interstitial or fused ommatidia and disordered sensory bristles. Since the fly compound eye is a neuronal structure easily accessible by light microscopy, the REP is an easy readout to assess changes in the decline of photoreceptor neurons caused by eye-specific expression of neurotoxic proteins. Thus, changes in REP have been successfully used in genetic screens set to identify modifiers of neurodegenerative disorders [28, 136, 254-256]. However, neurodegeneration in the fly eye cannot completely mimic the complex processes leading to disease in the human brain.

### 2.4.4 Drosophila models of polyglutamine disease

Disease models for polyQ disorders in Drosophila mostly involve overexpression of the common pathogenic feature of the causative proteins, concentrating on the expanded polyQ tract itself. Several fly lines have been introduced containing proteins entirely composed of normal or mutated polyQ stretches of different length (20Q, 22Q, 108Q, 127Q; [28, 257]). The expanded polyQ peptides in these models were sufficient to cause neurotoxicity despite the absence of their disease gene context. Additionally, studies have shown that a pure polyQ domain is much more toxic than a polyQ domain flanked by even relatively small protein sequences [258]. The detrimental intrinsic cytotoxic effects could be modified by genetic factors or modulations of the polyQ tract alone. These pure polyQ
approaches naturally neglect disease gene-specific characteristics and do not explain cell type specificity of distinct polyQ diseases. However, the previous work utilising such polyQ peptides revealed valuable novel insights into disease mechanisms.

The utilisation of truncated disease gene models is a feasible approach to study polyQ toxicity since it is assumed that causative proteins are also cleaved and thus truncated in vivo prior to oligomerisation and noxious effects. Several fly models for polyQ diseases are described. These models rely on expression of polyQ repeats either embedded in the C-terminal region of the human Ataxin-3/MJD protein (SCA3tr-Q27, SCA3tr-Q78; [219]), in an N-terminal truncated fragment of human Htt (Q2, Q75, Q120, Q128; [259, 260]) or exon 1 of Htt only (Q93; [75]). The distinction between the pathologies of polyQ diseases is more apparent at the level of truncated polyQ proteins. For instance, expression of the viral antiapoptotic protein p35 mitigated the REP in the SCA3tr-Q78 model, but failed to do so in Htt-trQ120 [219, 259]. Most of these truncated disease gene models exhibit progressive protein aggregation, forming nuclear inclusions in neurons, and late-stage neurodegeneration. On the contrary, flies with a normal number of repeats show a diffuse and cytoplasmic protein distribution and no overt neurotoxicity.

Expression of human versions of the full length polyQ proteins in Drosophila is an approach to investigate the pathogenic potential of elongated polyQ tracts in their native protein context. Studies show that high levels of wild-type full length Ataxin-1 (Q30) exert disturbances in eye morphology and expansion of the polyQ tract (Q82) leads to detrimental effects and a rough eye phenotype that can be modified by genetic interactors [136]. Investigations on Huntington's disease involve generation of a fly line with a fulllength Huntingtin containing 128Q [261], presenting a neurodegenerative eye phenotype due to demise of photoreceptor neurons. Similar approaches have been described for the androgen receptor in SBMA [262] and for full-length Ataxin-3 with polyQ expansion of 84 repeats in SCA3 [209]. In the latter model, flies showed severe and adult-onset neural degeneration when expression of the toxic disease protein was restricted to the compound eye or the nervous system, which was not observed for the wild-type protein. The fulllength mutated protein is more selectively toxic to the nervous system compared to the truncated isoform and accumulated in ubiquitinated inclusions. Coexpression of wild-type full-length Ataxin-3 on the contrary is able to ameliorate the detrimental effects of the toxic variant even in models of SCA1 and HD.

### 2.4.5 Previously implemented modifier studies

Several studies were conducted in order to reveal genes that modify the cytotoxicity and deleterious consequences of polyQ expansion in the diverse causative disease proteins. As already mentioned above, the baculovirus antiapoptotic gene p35 suppressed truncated mutant Ataxin-3-dependent degeneration in the eye, as does the human heat shock protein HSP70 [30, 219]. Kazemi-Esfarjani et al. published results of one of the first large scale modifier screens in flies expressing prolonged polyQ peptides only [28]. They utilised a set of 7,000 P-element insertions for crossbreeding with the disease flies and assessed the offspring for suppression or enhancement of the polyQ-induced rough eye phenotype. Out of a number of potential candidates they presented two chaperone-related gene products, dHDJ1 (equivalent to human HSP40/HDJ1) and dTPR2 (equivalent to human TPR2) as potent genetic suppressors of polyQ toxicity. Fernandez-Funez et al. utilised a fly model based on full-length Ataxin-1 Q82 expression for two screens with 1,500 lethal P-elements and 3,000 EP insertions respectively, also evaluating the change of REP in the F1 generation [136]. They identified 18 genes that, if altered in expression, enhanced or suppressed Ataxin-1 toxicity. Among these genes were some coding for ubiquitin-related proteins, chaperones, RNA-binding molecules and transcription factors. Bilen et al. described the crucial involvement of microRNAs (miRNAs) pathways in the modulation of polyQ toxicity induced by Ataxin-3 after screens in Drosophila and human cells [263]. The same group conducted a genome-wide EP element-based screen for modifiers of Ataxin-3induced neurodegeneration in Drosophila [256]. They identified 25 modifiers representing 18 genes that are mainly involved in biological processes affecting protein misfolding and ubiquitin-related pathways. Among others, this experiment was designed as a genetic highthroughput screen based on the misexpression of endogenous genes [264]. Despite their general feasibility, these screening strategies create artificial expressions states potentially masking the native influence of the respective gene product. Furthermore, they are mostly confined to a small portion of the genome.

These disadvantages can be overcome by mimicking classical knockout experiments with the help of RNA interference-mediated gene silencing. This powerful technique has been successfully implemented into high-throughput screening for modifiers of Huntingtin aggregation in yeast [265] and Drosophila cells [266]. Genome-wide studies utilising RNAi revealed regulators of polyQ and Huntingtin aggregation in C. elegans [267] and Drosophila [268] respectively.

## 3 Aim of the Study

The comprehensive understanding of molecular mechanisms and cellular pathways resulting in polyQ neurotoxicity and pathogenesis are a prerequisite for the development of effective treatments for the corresponding disorders. In an attempt to contribute to this process we intended to conduct a genome-wide high-throughput modifier screen in order to identify genetic modifiers of polyQ neurotoxicity in Drosophila melanogaster. This should be accomplished first of all by characterisation of a feasible disease model exhibiting a readily accessible readout for the large scale experiments. Expression of a human variant of truncated Ataxin-3, harbouring a stretch of 78 glutamines, in the compound eye results in an REP. This REP combines pathological involvement of neuronal photoreceptor cells with an easy exterior observability of neurodegeneration. By means of RNA interference we planned to knockdown a genome-wide set of potential modifier genes, thereby evaluating the impact of the gene silencing on the REP. For this purpose, we obtained a set of fly RNAi strains from the VDRC representing all Drosophila genes known to have an orthologue in humans. A genetic modifier screen with these $\sim 7,500$ genes would represent the most comprehensive endeavour in this field and setting so far. The gene knockdown approach, the in vivo situation and the easy assessment of neurotoxicity modification mark the advantages of this work in comparison to previously conducted modifier screens. By subsequent thorough analysis and processing of our results and the obtained modifiers we hope to aid in conceiving polyQ diseases better and opening avenues for therapeutic approaches.

## 4 Material and Methods

### 4.1 Chemicals, reagents and equipment

Composition of reagents and buffers is specified in Table 2 and referred to as such with the respective name in the text.

Table 2. Chemicals and reagents.

| Name | Specification/Composition | Source/Manufacturer |
| :---: | :---: | :---: |
| Acridine Orange | 3,6-bis[Dimethylamino]acridin (A-6014) | Sigma, USA |
| Acrylamide | Acrylamide 2K ( 30 \%) | AppliChem, Germany |
| APS | Ammonium persulphate $\geq 98 \%$ (9592.2) | Roth, Germany |
| Bromophenol blue | (A512.1) | Roth, Germany |
| $\mathrm{CaCl}_{2} \times 2 \mathrm{H}_{2} \mathrm{O}$ | Calciumchloride dihydrate $\geq 99$ \% p.a. (5239.2) | Fluka, Germany |
| Chloroform | Trichlormethane $\geq 99$ \% (3313.1) | Roth, Germany |
| Citrate buffer | 1.8 mM citric acid <br> 8.2 mM sodium citrate, pH 6.0 |  |
| Citric acid | (1.00244.1000) | Merck, Germany |
| DEPC | Diethylpyrocarbonate (K028.1) | Roth, Germany |
| Drosophila's Ringer | $\begin{aligned} & 182 \mathrm{mM} \mathrm{KCl} \\ & 46 \mathrm{mM} \mathrm{NaCl}^{2} \\ & 3 \mathrm{mM} \mathrm{CaCl}_{2} \times 2 \mathrm{H}_{2} \mathrm{O} \\ & 10 \mathrm{mM} \mathrm{Tris}, \mathrm{pH}_{7.2} \end{aligned}$ |  |
| DTT | 1,4-Dithiothreitol (6908.1) | Roth, Germany |
| Entellan | (1.07961.0500) | Merck, Germany |
| Ethanol | $\geq 99.8$ \% p.a. (9065.4) | Roth, Germany |
| Fluoromount | Fluoromount-G (0100-01) | Southern Biotech, USA |
| Glycerol | 99 \% (G5516-1L) | Sigma, USA |
| Glycine | $\geq 99$ \% p.a. (3908.2) | Roth, Germany |
| HEPES | $\geq 99.5$ \% p.a. (9105.4) | Roth, Germany |
| ECL solution | Immun-Star ${ }^{\text {TM }}$ WesternC $^{\text {TM }}$ Kit (170-5070) | Bio-Rad, USA |
| Isopropanol | 2-Propanol (T910.1) | Roth, Germany |
| KCl | Potassium chloride, $\geq 95.5$ \% p.a. (6781.1) | Roth, Germany |
| $\mathrm{KH}_{2} \mathrm{PO}_{4}$ | Potassium dihydrogen phosphate, $\geq 99$ \% p.a. | Roth, Germany |


| Laemmli buffer (5x) | 250 mM Tris, pH 6.8 <br> 10 \% (v/v) SDS <br> 1.25 \% (w/v) bromophenol blue <br> 10mM EDTA <br> 0.03 \% (v/v) $\beta$-mercaptoethanol <br> 50 \% (v/v) glycerol |  |
| :---: | :---: | :---: |
| Methanol | (717.1) | Roth, Germany |
| Methyl benzoate | (822330.1000) | Merck, Germany |
| $\mathrm{MgCl}_{2}$ | Magnesium chloride $\geq$ 98,5 \% p.a. (KK36.1) | Roth, Germany |
| $\mathrm{NaH}_{2} \mathrm{PO}_{4} \times \mathrm{H}_{2} \mathrm{O}$ | Monosodium phosphate monohydrate, $\geq 99 \%$ p.a. (1.06342.1000) | Merck, Germany |
| NaCl | Sodium chloride, $\geq 95.5$ \% p.a. | Roth, Germany |
| $\mathrm{Na}_{2} \mathrm{HPO}_{4} \times \mathrm{H}_{2} \mathrm{O}$ | Disodium phosphate monohydrate, $\geq 99.5$ \% p.a. (1.06580.1000) | Merck, Germany |
| $\mathrm{NaHCO}_{3}$ | Sodium hydrogen carbonate, $\geq 99$ \% pure (8551.1) | Roth, Germany |
| NGS | Normal goat serum (S-2007) | Sigma, USA |
| Nitrocellulose membrane | Protran ${ }^{\circledR}$ BA 83 (10 402 396) | Whatman, Germany |
| Paraffin | 32.3 \% Paraffin 42-44 (107150) <br> 32.3 \% Paraffin 51-53 (107157) <br> 32.3 \% Paraffin 57-60 (107158) <br> 3.2 \% bees wax | Merck, Germany (Paraffin) Roth, Germany (beeswax) |
| PBS (0.01 M) | Phosphate-buffered saline $8 \mathrm{mM} \mathrm{Na}_{2} \mathrm{HPO}_{4} \times \mathrm{H}_{2} \mathrm{O}$ <br> $1.8 \mathrm{mM} \mathrm{NaH}_{2} \mathrm{PO}_{4} \times \mathrm{H}_{2} \mathrm{O}$ <br> 0.1 M NaCl |  |
| PBS-T | 0.14 M NaCl <br> $6 \mathrm{mM} \mathrm{Na}_{2} \mathrm{HPO}_{4}$ <br> 2.7 mM KCl <br> $1.5 \mathrm{mM} \mathrm{KH}_{2} \mathrm{PO}_{4}$ <br> 0.4 \% Triton-X100 |  |
| PFA | 4 \% paraformaldehyde (0335.2) in PBS/PBS-T | Roth, Germany |
| Phalloidin | Alexa Fluor ${ }^{\circledR} 568$ Phalloidin | Invitrogen, Germany |
| Resolving gel buffer (SDS-PAGE) | $0.4 \text { \% SDS }$ <br> 1.5 M Tris, pH 8.8 |  |
| RIPA lysis buffer | 50 mM Tris, pH 8.0 <br> 0.15 M NaCl <br> 0.1 \% (v/v) SDS <br> 1 \% NP-40 <br> 0.5 \% Sodium deoxycholate <br> Protease inhibitor (Roche) |  |
| Running buffer (SDSPAGE) | 0.1 M Tris <br> 1 M glycine 0.5 \% SDS |  |
| SDS | Sodium dodecyl sulphate, AccuGene 10 \% (51213) | Cambrex, USA |


| Skim milk | Milk powder (T145.2) | Roth, Germany |
| :---: | :---: | :---: |
| Sodium citrate | (1.06448.0500) | Merck, Germany |
| Stacking gel buffer (SDS-PAGE) | $\begin{aligned} & 4 \text { \% SDS } \\ & 0.25 \text { M Tris, pH } 6.8 \end{aligned}$ |  |
| Stripping buffer (WB) | 0.2 M glycine 0.5 M NaCl pH 2.8 |  |
| Sucrose | $\mathrm{D}(+)$-saccharose $\geq 99.5$ \% p.a. (4621.1) | Roth, Germany |
| TBS | 25 mM Tris <br> 140 mM NaCl <br> pH 7.5 |  |
| TBS-T | $\begin{aligned} & 25 \mathrm{mM} \text { Tris } \\ & 140 \mathrm{mM} \mathrm{NaCl} \\ & \mathrm{pH} 7.5 \\ & 0.05 \% \text { Tween-20 } \end{aligned}$ |  |
| TEMED | N,N, ${ }^{\prime}$, $\mathrm{N}^{\prime}$-Tetramethylethylendiamide | AppliChem, Germany |
| Transfer buffer (WB) | 25 mM Tris <br> 192 mM glycine <br> 20 \% (v/v) methanol |  |
| Tris | $\geq 99.3$ \% (AE15.2) | Roth, Germany |
| Triton-X100 |  | Sigma, USA |
| Tween ${ }^{\text {® }} 20$ | (9127.1) | Roth, Germany |
| Vectashield | Vectashield mounting medium $\mathrm{w} /$ and $\mathrm{w} / \mathrm{o}$ DAPI | Vector, USA |
| Xylene |  | Otto Fischar, Germany |

Table 3. Equipment.

## Name/Specification Manufacturer

Microtome HM 360
Cryotome CM3050 S
Zoom stereo microscope SZ51 with KL200 LED light source
Biological microscope CX31
Zoom stereo microscope SZX10 with S80-55 RL ring light and SC30
digital camera
Routine microcope CKX41 (inverted) with U-RFLT50 light source and SC30 camera

Thermo Scientific, Germany
Leica, Germany
Olympus, Germany
Olympus, Germany
Olympus, Germany

Olympus, Germany
$\qquad$

Research microscope BX51 with X-Cite ${ }^{\circledR}$ 120Q light source and DP72 digital camera

Tissue homogenizer SpeedMill P12
Chemiluminescence documentation system Alliance
LD4.77.WL.Auto
Scanning electron microscope ESEM XL 30 FEG

Multimode microplate reader Infinite ${ }^{\circledR}$ M200

Olympus, Germany

Analytik Jena, Germany
Biometra, Germany

FEI, Eindhoven, Netherlands

Tecan, Germany

Table 4. Software and online resources.

| Name | Application |
| :--- | :--- |
| Adobe $^{\circledR}$ Illustrator ${ }^{\circledR}$ CS4 | Creation of illustrations |
| Adobe ${ }^{\circledR}$ Photoshop ${ }^{\circledR}$ CS4 | Image processing |
| ImageJ 1.42 | Counting of photoreceptor neurons |
| GNU Image Manipulation Program (GIMP) 2.6 | Image processing, compilation of figures |
| GraphPad ${ }^{\circledR}$ Prism ${ }^{\text {TM } 5}$ | Statistical analysis |
| Olympus Cell A | Documentation of eye phenotypes <br> Alym cytochemical micrographs |
| Alliance UVItec 15.11 | Documentation of Western blots and filter <br> retardation assays |
| hioDoc Analyse 2.1 | Quantification of Western blots and filter retardation <br> assays |
| http://www.ncbi.nlm.nih.gov/homologene | Alignments for determination of gene orthologues <br> between Drosophila melanogaster and Homo sapiens |
| http://www.ihop-net.org/UniPub/iHOP/ | Identification of gene orthologues between <br> Drosophila melanogaster and Homo sapiens |
| Identification of gene orthologues between |  |
| Drosophila melanogaster and Homo sapiens |  |

### 4.2 Fly experiments

In this work, genes are generally set in italics, Drosophila genes are written in small letters in contrast to human orthologue genes which are written in capital letters. Gene names are phrased as the official short form or designation according to NCBI Gene database [269]. If not stated otherwise, candidate genes are referred to as the fly variant. Transgenes, fly genotypes and fly stocks are written in italics and as proposed by FlyBase nomenclature [270].

### 4.2.1 Transgenic flies and housing conditions

Fly stocks were maintained on standard cornmeal-agar-yeast-molasses-based food at $18{ }^{\circ} \mathrm{C}$. Standard crossbreeding and other experiments with larvae and adult Drosophila were conducted on $25^{\circ} \mathrm{C}$. For adult-onset expression, GAL80-expressing flies were shifted from permissive temperature at $18{ }^{\circ} \mathrm{C}$ to restrictive temperature of $29^{\circ} \mathrm{C}$.

Table 5. Utilised Drosophila melanogaster strains.

| Transgenic line | Genotype ${ }^{1}$ | Source |
| :---: | :---: | :---: |
| Actin5C-GAL4 | $y[1] w[*] ; P\{w[+m C]=A c t 5 C-G A L 4\} 25 F 01 / C y O, y[+]$ | Bloomington \#4414 |
| CxD/TM3 | $y[1], w[*] ; ; C x D / T M 3, S b[1], S e r[1] ~$ | Bloomington \#6309 |
| elav ${ }^{\text {C155-GAL4 }}$ | $P\{w[+m W . h s]=$ GawB $\}$ elav[C155] | Bloomington \#458 |
| elav-tub-GAL80 | $\begin{aligned} & P\{w[+m W \cdot h s]=G a w B\} e l a v[C 155] ; P\{w[+m C]=t u b P- \\ & \text { GAL8O[ts] } 20 \end{aligned}$ | created by Malte Butzlaff <br> (original GAL80 stock: <br> Bloomington \#7019) |
| FM7 | FM7a | Bloomington \#785 |
| GMR-GAL4 | $w\left[^{*}\right] ; P\{w[+m C]=$ GAL4-ninaE.GMR\}12 | Bloomington \#1104 |
| Oregon-R-C | wild type | Bloomington \#5 |
| Rhodopsin-GAL4 | $P\{r y[+t 7.2]=r h 1-G A L 4\} 3, r y[506]$ | Bloomington \#8691 |
| sal/Cyo | salm[1],cn[1],bw[1]sp[1]/CyO | Bloomington \#3274 |
| UAS-ATXN1 Q82 | $y[1] w[118] P\{[+]=U A S-S C A 1.82 Q\}[F 7]$ | gift of Juan Botas [136] |


| UAS-eGFP | $w[*] ; P\{w[+]=U A S-e G F P\} \# 4.2 / C y O$ | created by Aaron Voigt |
| :---: | :---: | :---: |
| UAS-Htt Exon1 Q93 | $w\left[{ }^{*}\right] ; P\{w[+]=U A S-Q 93 e x 1\} K 6,9,15 R$ | gift of Lawrence Marsh [75] |
| UAS-lacZ | $P\{w[+m C]=U A S$-lacZ.Exel $\} 2$ | Bloomington \#8529 |
| UAS-SCA3-Q84 | $w[*] ; P\{w[+]=U A S-M J D-Q 84\} 7.2 / T M 3, S b$ | gift of Nancy Bonini [209] |
| UAS-SCA3tr-Q27 |  | Bloomington \#8149 |
| UAS-SCA3tr-Q78 |  | Bloomington \#8150 |
| UAS-Tau[R406W] | $w[*] ; P P\{w[+m C]=U A S-h T a u[R 406 W]\}$ | gift of Mel Feany [271] |

${ }^{1}$ genotype as suggested by Flybase [270].

RNAi fly strains comprising the human orthologue sublibrary were purchased from the Vienna Drosophila RNAi Centre (VDRC) [250] where they have been generated by random integration of shRNA-transcribing inverted repeats under UAS-GAL4 control into the Drosophila genome (UAS-shRNA).

The 7,488 RNAi lines for the human orthologue sublibrary used in this study were selected by the VDRC considering known or predicted human orthologues to the fly genes (see Appendix Table 3 for complete list).

### 4.2.2 Mating procedures

Mating procedures for the subsequent screens were essentially the same. Screening for eye changes by shRNA expression itself was conducted using the GMR-GAL4 line as a control. GMR-GAL4 and UAS-SCA3tr-Q78 strains were recombined in order to generate the screening stock for the polyQ modifier screen (GMR_SCA3tr-Q78). For the screening of polyQ specificity of RNAi candidates, flies overexpressing the mutant tau transgene UASTau[R406W] in the eye were utilised (GMR_Tau[R406W]). For screening purposes, stocks were crossbred with $U A S$-shRNA lines from the VDRC.

The random integrations of shRNA were located on chromosomes one, two and three. RNAi lines with integration on the first chromosome (X) have been generated utilising an artificial double $\mathrm{X}(\mathrm{dX}$ ) gonosome (male: XY; female: dXY ) with combination of three X chromosomes ( dXX ) being lethal. Integrations are found on the single X
chromosome, leading to the restriction of the shRNA transgene to male carriers. Thus, male flies of the RNAi strains were generally crossbred to virgin female flies of the screening stocks and effects of the gene knockdown were assessed in female progeny.

Table 6. Stocks utilised for screening approaches.

| Screening stock | Genotype |
| :--- | :--- |
| GMR_GAL4 | $w\left[^{*}\right] ; P\{w[+m C]=G A L 4-$ ninaE.GMR $\} 12$ |
| GMR_SCA3tr-Q78 | $y\left[^{*}\right] w\left[^{*}\right] ; P\{w[+m C]=G A L 4-$ ninaE.GMR $\} 12, P\{w[+]=U A S-M J D-t r Q 78\} S t r o n g / C y O$ |
| GMR_Tau[R406W] | $w\left[^{*}\right] ; P\{w[+m C]=G A L 4-n i n a E . G M R\} 12 / C y O ; P\{w[+]=U A S-T a u[R 406 W]\} / T M 3, S b$ |

### 4.2.3 Evaluation of rough eye phenotype modification

For assessment of REP modulation, at least five female flies were analysed for changes in the severity of eye degeneration. Modifications by the induction of RNAi in polyQ and tau models were categorized as follows: "wild type-like phenotype (1)", obvious REP suppression (2)", "subtle REP suppression (3)", "no change of REP (4)", "subtle enhancement of REP (5)", "obvious enhancement of REP (6)" and "lethal (7)". For the GMRGAL4 screening only the "no change" and enhancement terms apply. RNAi lines exhibiting such effects in the GMR-GAL4 control flies were excluded from subsequent experiments due to impact unconnected to expression of elongated polyQ. Designation of an RNAi line as polyQ modifier candidate required no change in control flies and an at least obvious enhancement/suppression of the REP in three independent experiments. Candidate lines were tested for polyQ specificity by rescreening with Tau[R406W] screening stock. RNAi lines exhibiting similar effects in both models were excluded from the polyQ candidate set and remainder strains were subjected to more detailed analysis.

Data on screen results were managed and stored making use of a databank generated with MySQL by Dr. Malte Butzlaff (Dept. of Neurology, Aachen).

### 4.2.4 Documentation of eye phenotypes

Drosophila compound eyes were pictured using an Olympus zoom stereo microscope (Table 3) at 6.3 x magnification and Cell A software (Table 4). Eye documentation with scanning electron microscopy (SEM) was conducted on unfixated and uncoated flies utilising the ESEM scanning electron microscope (Table 3) in low vacuum mode (0.8-1.5 Torr) and an accelerating voltage of 10 kV . GIMP 2.6 and Adobe ${ }^{\circledR}$ Photoshop ${ }^{\circledR}$ CS4 software was used for rotating and cropping of images. All whole compound eye images compiled in this work feature dorsal-up and anterior-left orientation.

### 4.2.5 Dissection and staining of eye imaginal discs

Dissection of L3 Drosophila larvae was performed as described previously [272]. For staining with vital dye, preparation was conducted in Drosophila's Ringer solution (Table 2), otherwise PBS-T was used. For subsequent immunohistochemistry, isolated eye discs were subjected to fixation in $4 \%$ paraformaldehyde (PFA) in PBS-T for 30 min , followed by several washing steps in PBS-T. Subsequent to blocking with $4 \%$ normal goat serum (NGS), tissue was incubated with an antibody directed against the HA-tag of the truncated Ataxin3 protein (mouse anti-HA, Covance) over night at $4^{\circ} \mathrm{C}$. Primary antibody was detected with an Alexa Fluor ${ }^{\circledR}$ 488-coupled secondary anti-mouse antibody. Tissue was mounted in DAPI-containing Vectashield mounting medium on glass slides and imaged afterwards using an Olympus BX51 fluorescence microscope equipped with 20x and 40x objectives.

For acridine orange staining, vital dye was dissolved in Drosophila's ringer solution to achieve a concentration of $1.6 \times 10^{-6} \mathrm{M}$ for dissection. Isolated eye discs were placed on a glass slide, fastened with a cover slide and immediately visualised using the same microscope properties as described for immunohistochemistry.

### 4.2.6 Longevity analysis

For evaluation of life span, male flies with the respective genotype were raised and selected at $18{ }^{\circ} \mathrm{C}$ (for adult-onset system with temperature-sensitive GAL80) or $25^{\circ} \mathrm{C}$ and grouped into vials containing 20 flies maximum. Longevity experiments were conducted at
$25^{\circ} \mathrm{C}$ for normal UAS/GAL4 strains; lines containing GAL80 were shifted to the restrictive temperature of $29^{\circ} \mathrm{C}$ for onset of transgene expression. Flies were transferred to fresh food twice a week and death events were counted at least every second day. A minimum of 20 flies per genotype were scored for every longevity assay. GraphPad Prism 5 was used for Kaplan-Meyer plotting and statistical analysis, featuring Log-Rank test for comparison of two longevity curves.

### 4.2.7 Protein collection from fly heads

The procedure was utilised for eye-specific and pan-neural transgene expressing strains alike. Flies were placed in reaction tubes and flash frozen in liquid nitrogen. Frozen flies were vortexed for separation of heads from bodies. Heads were transferred to vials containing radioimmunoprecipitation assay (RIPA) buffer (10 $\mu \mathrm{L}$ per head) and a small quantity of ceramic beads. The samples were homogenised with the SpeedMill P12 homogeniser ( $2 \times 2 \mathrm{~min}$, predefined programme for insect tissues). Subsequent to homogenisation, samples were centrifuged in order to pellet the beads and crude debris (13,300 rpm, $5 \mathrm{~min}, 4^{\circ} \mathrm{C}$ ), followed by collection of the supernatant and an additional centrifugation step ( $13,300 \mathrm{rpm}, 5 \mathrm{~min}, 4^{\circ} \mathrm{C}$ ) for clearing. Protein concentration was measured using the Bio-Rad $D_{C}$ Protein Assay Kit and the Tecan ${ }^{\circledR}$ multimodal microplate reader.

### 4.2.8 Immunoblotting

For Western blot analysis, protein samples were diluted in loading buffer (5x Laemmli) and boiled ( $95^{\circ} \mathrm{C}$, 5 min ) before loading onto a polyacrylamide gel ( $10 \%, 12 \%$ or 4-12 \% gradient gels) and subsequent SDS-PAGE (run at 100 V for 90 min with running buffer). Resolved proteins were transferred onto nitrocellulose membrane ( 225 mA per gel for 90 min with transfer buffer). The membranes were then blocked with skim milk (5 \% in TBS-T for 60 min ) followed by incubation with the primary antibody at $4{ }^{\circ} \mathrm{C}$ overnight (in 0.5 \% skim milk in TBS-T, for antibody concentrations see Table 7).

Table 7. Antibodies utilised for Drosophila head and cell lysate immunoblotting and for immunohistochemical stainings.

| Antibody | Species | Dilution | Manufacturer |
| :--- | :--- | :--- | :--- |
| primary antibodies | mouse monoclonal | $1: 500$ | Covance, USA |
| anti-haemagglutinin | mouse monoclonal | $1: 1,000$ | Millipore, USA |
| anti-polyglutamine | mouse monoclonal | $1: 1,000$ | Millipore, USA |
| anti-myc tag | mouse monoclonal | $1: 1,000$ | Roche, Germany |
| anti-GFP | mouse monoclonal | $1: 500$ | DSHB, USA |
| anti-TRMT2A | rabbit polyclonal | $1: 1,000$ | Abcam, UK |
| anti- $\beta$-tubulin | mouse monoclonal | $1: 2,500$ | DSHB, USA |
| anti-VDAC/porin |  |  | Sigma-Aldrich, USA |
| anti-syntaxin | sheep | $1: 10,000$ | GE Healthcare, Germany |
| secondary antibodies |  |  |  |
| ECL anti-mouse IgG, | donkey | $1: 10,000$ | GE Healthcare, Germany |
| HRP-coupled |  |  |  |
| ECL anti-rabbit IgG, | goat |  |  |
| HRP-coupled | Alexa Fluor® 488 | anti-mouse IgG (H+L) |  |

After three washing steps of 5 min in TBS-T, membranes were probed with appropriate secondary antibodies for 60 min at room temperature. Following three additional washings in TBS-T, chemiluminescence signal was induced using Immun-Star ${ }^{\text {TM }}$ ECL solution (Bio-Rad, Germany) as a HRP substrate and captured with Alliance LD4 documentation system (Biometra, Germany) and Alliance UVItec software. Band intensity was quantified utilising BioDoc Analyse software (Biometra, Germany).

### 4.2.9 Filter Retardation Assay

Filter retardation assays were mainly conducted as described previously [273-275]. Equal protein amounts ( $15 \mu \mathrm{~g}$ ) of RIPA fly head lysates were adjusted to $2 \%$ SDS and 50 mM dithiothreitol followed by heating to $100{ }^{\circ} \mathrm{C}$ for 5 min . Using a dot blot filtration unit, the resulting solutions were filtered through a $0.2 \mu \mathrm{~m}$ nitrocellulose membrane
(Whatman, UK)) previously equilibrated with 0.1 \% SDS in TBS and afterwards washed in TBS-T. Membranes were further processed for immunodetection as described in section 4.2.8 by blocking and probing with primary (anti-HA, anti-GFP, anti-myc) and secondary antibodies followed by documentation.

For assessment of the aggregate load of the individual polyQ-shRNA-coexpressing fly lines, results of band intensity measurements were normalised to that of the polyQ control set as a hypothetical mean of 1.0 and calculated with One-sample t-test. If applicable, lysates from three independent crossbreeds were utilised.

### 4.2.10 Histological and immunohistochemical staining of paraffin sections

For paraffin sections, heads of female flies were fixated in $4 \%$ paraformaldehyde/PBS-T for 60 min and subsequently subjected to dehydration in an ascending alcohol series ( 30 min in each: $70 \%, 80 \%, 90 \%, 2 \times 100 \%$ ethanol). Fly heads were incubated in methyl benzoate for 60 min and three times in paraffin at $62{ }^{\circ} \mathrm{C}$ for 30 min each before embedding and hardening at room temperature overnight. Embedded fly heads were cut into $6 \mu$ m thick frontal sections with Feather C35 single-use blades using a Thermo Scientific microtome. For staining, sections were incubated in xylene for 20 min twice and rehydrated in an descending alcohol series (5 min in each: $2 \times 100 \%, 90 \%, 80 \%$, 70 \% ethanol) followed by rinsing in PBS.

For toluidine blue histological staining, sections were incubated for 5 min in $0.1 \%$ toluidine blue solution with $2.5 \% \mathrm{NaHCO}_{3}$, rinsed with deionised water and dehydrated in an ascending alcohol series (as described above), followed by dual incubation in xylene for 10 min each. Finally, sections were mounted with entellan and documented using an Olympus BX51 microscope and Cell F software.

For immunohistochemical staining, sections were subjected to heating in citrate buffer (Table 2) at 1000 W for 10 min in a microwave oven in order to demask protein epitopes. Afterwards, sections were washed in PBS for 5 min , blocked for 30 min with $4 \%$ normal goat serum and probed with primary antibody in $4 \%$ NGS/PBS for 3 h at $37{ }^{\circ} \mathrm{C}$. Following washing in PBS for 5 min , sections were incubated with Alexa Fluor ${ }^{\circledR}$ 488coupled anti-mouse secondary antibody (Invitrogen, see Table 7) for 60 min at room temperature. Rinsing in PBS was conducted prior to mounting of the sections in Vectashield ${ }^{\circledR}$ (Vector Laboratories, UK) mounting medium (with or without DAPI). For
documentation of the slides Cell F software and an Olympus BX51 microscope equipped with a fluorescence light source were utilised.

### 4.2.11 Immunohistochemical staining of cryo sections

For preparation of cryo sections, heads of female flies were fixated in $4 \%$ paraformaldehyde/PBS-T overnight, followed by cryoprotection with $10 \%$ sucrose for 12 h and $30 \%$ sucrose overnight. Specimens were cut into $16 \mu \mathrm{~m}$ thick frontal sections using a Leica cryostat and stored at $-20^{\circ} \mathrm{C}$.

For immunostaining, sections were rinsed for 5 min in PBS and immediately blocked with $4 \%$ NGS/PBS, followed by primary antibody incubation (Table 7) in $4 \%$ NGS/PBS for 3 h at $37^{\circ} \mathrm{C}$. Afterwards, sections were washed briefly in PBS and probed with Alexa Fluor ${ }^{\circledR}$ 488-coupled anti-mouse secondary antibody for 60 min at room temperature. Eventually, sections were washed in PBS and mounted in Vectashield ${ }^{\circledR}$ mounting medium. Visualisation was done with an Olympus BX51 microscope equipped with a fluorescent light source and Cell F software.

### 4.2.12 Semi-thin tangential sectioning of fly heads and photoreceptor quantification

For preparation of semi-thin sections for quantification of photoreceptor cells, fly heads of respective genotypes were fixated in 4 \% PFA/PBS-T overnight. Subsequently, heads were dehydrated in an ascending alcohol series ( 30 min in each: $70 \%, 80 \%, 90 \%$, $2 \times 100 \%$ ) and equilibrated in LR White embedding medium with LR White catalyst (Fluka, USA) for 3 x 1 h . Following this, LR White accelerator was added and resin left for hardening at room temperature for 24 h . After setting, specimens were roughly cut to size with razorblades and fine trimmed utilising glass blades. For $1 \mu \mathrm{~m}$ tangential sectioning of fly heads diamond blades and a Thermo Scientific microtome were employed. Tissue was stained with toluidine blue for 5 min and allowed to dry prior to mounting with Entellan ${ }^{\circledR}$. Visualisation was done using an Olympus BX51 microscope and Cell F software.

For quantification, rhabdomeres as an indicator of photoreceptor (PR) neurons were counted in 50 ommatidia of $n=3$ individuals per genotype and presented as mean $P R$ count $\pm$ standard error of the mean (SEM) per ommatidium. Two-tailed Mann-Whitney test was used to calculate the p-value when comparing the photoreceptor counts between two
groups. In experiments with three groups, Kruskal-Wallis test was performed followed by Dunn post-tests.

### 4.3 Cell culture experiments

### 4.3.1 Cell culture conditions and media

Human HEK293 cells were maintained in Dulbecco's modified Eagle's medium (DMEM, PAN Biotech, Germany) supplemented with 10 \% fetal calf serum (FCS, Biochrom AG, Germany), 100 units/mL penicillin and $100 \mathrm{mg} / \mathrm{mL}$ streptomycin (both Biochrom AG, Germany), in a humidified atmosphere of $5 \% \mathrm{CO}_{2} / 95 \%$ air at $3{ }^{\circ} \mathrm{C}$ (Thermo Scientific HERACell 150i incubator). For selection of stably transduced cells with shRNA, $0.5 \mu \mathrm{~g} / \mathrm{mL}$ puromycin (Invitrogen, Germany) was added.

### 4.3.2 Generation of stable shRNA-expressing cells

Silencing of TRMT2A expression in HEK293 cells was accomplished by infection of cells with lentiviral transduction particles carrying an inverted repeat sequence in a lentiviral plasmid vector ( $p L K 0.1$-puro). The sequence is integrated into the host cell DNA and shRNA directed against the TRMT2A mRNA is expressed under the control of a human phosphoglycerate kinase eukaryotic promoter. Commercially available transduction particles were all purchased from Sigma-Aldrich (as listed in Table 8). shRNA transduction particles having no known human target sequence were used as a negative control of expression knockdown, activating the RNAi machinery of the cell without affecting the expression of any gene.

Table 8. Lentiviral clones and non-target strain utilised for TRMT2A silencing experiments in HEK293 cells.

## Lentiviral clone ID

## Sequence

MISSION ${ }^{\circledR}$ shRNA Lentiviral Transduction Particles

| NM_182984.2-856s1c1 | CCGGGCAGACTGAGTATCGTAATAACTC- |
| :--- | :--- |
|  | GAGTTATTACGATACTCAGTCTGCTTTTTTG |
| NM_182984.2-1574s1c1 | CCGGCAGGACAATGAGTTGAGTAATCTC- |
|  | GAGATTACTCAACTCATTGTCCTGTTTTTTG |


| NM_182984.2-736s1c1 | CCGGGCAGAAACTTGCCAAGGAAATCTC- |
| :--- | :--- |
|  | GAGATTTCCTTGGCAAGTTTCTGCTTTTTTG |
| NM_182984.2-1985s1c1 | CCGGCCAGATAACACCCTACAAGAACTC- |
|  | GAGTTCTTGTAGGGTGTTATCTGGTTTTTTG |
| NM_182984.2-1505s1c1 | CCGGCGGAAGGTAAAGAGGGTCATTCTC- |
|  | GAGAATGACCCTCTTTACCTTCCGTTTTTTG |

MISSION ${ }^{\circledR}$ Non-Target shRNA Control Transduction Particles

Transfection of HEK293 cells with lentiviral transduction particles was completed following manufacturer's instructions by Daniela Otten (Department of Biochemistry, University Medical Centre Aachen). Multiplicities of infection (MOI) of 2, 5, 10 and 15 were utilised. Puromycin-resistant colonies were selected and cultured further, decrease of TRMT2A protein levels due to gene silencing was assessed by Western blot analysis.

### 4.3.3 Plasmid transfection

Transfection of plasmids into HEK293 cells was accomplished using Metafectene ${ }^{\circledR}$ (Biontex, Germany) following manufacturer's instructions. Cells were seeded on poly-Llysine (PLL)-coated plates ( 35,000 cells/ $\mathrm{cm}^{2}$ ) and then transfected 48 h before experimentation. Cells and expression of GFP-tagged constructs were visualised with an Olympus inverse cell culture microscope equipped with a fluorescence light source.

### 4.3.4 Protein collection from cell culture and immunoblotting

Plated cells were washed in ice cold PBS and then lysed in RIPA buffer under agitation for 30 min at $4^{\circ} \mathrm{C}$. Samples were centrifuged for 15 min at $13,300 \mathrm{rpm}$ at $4^{\circ} \mathrm{C}$ and supernatants were collected. Protein concentrations were measured using the Bio-Rad $\mathrm{D}_{\mathrm{C}}$ Protein Assay and Tecan ${ }^{\circledR}$ multimode microplate reader. Western blotting was then performed as outlined in chapter 4.2.8.

### 4.3.5 Cytochemistry

Cells were plated on laminin (mouse, Sigma, Germany)-coated glass slips. 48 hours after transfection with GFP-coupled huntingtin transgene plasmids, cells were washed once in PBS and fixed in $4 \%$ PFA for 30 min at RT. After three washes in PBS, cells were incubated with Alexa Fluor ${ }^{\circledR}$ 568-coupled phalloidin (Invitrogen, Germany; 1:500 in PBS-T) for 20 min at room temperature. Subsequently, cells were washed three times in PBS and nuclei were counterstained by briefly rinsing the cells in DAPI solution ( $2 \mu \mathrm{~g} / \mathrm{mL}$ ). Cells were again washed thrice in PBS before being mounted on glass slides using Fluoromount$\mathrm{G}^{\mathrm{m}}$ (Southern Biotech, USA). GFP-positive cells and thereof inclusion-positive cells were counted and statistics were calculated with unpaired t-test using GraphPad Prism 5.

## 5 Results

### 5.1 Characterisation of a SCA3 fly model for the modifier screen

We intended to conduct an RNAi screen to identify genetic modifiers of polyQinduced neurotoxicity in Drosophila. In order to achieve this goal, a screening stock was generated by recombination of a truncated version of the human SCA3 causative gene ATXN3 ( $P\{w[+m C]=U A S-H s a p \ M J D . t r-Q 78\}$ ) [219] (from now on termed SCA3tr-Q78) with the GMR-GAL4 ( $w[*] ; P\{w[+m C]=G A L 4-n i n a E . G M R\} 12$ ) driver. The resulting fly line with $w\left[^{*}\right] ; P\{w[+m C]=G A L 4-$ ninaE.GMR\}12, $P\{w[+m C]=U A S-H s a p \ M J D . t r-Q 78\} / C y O$ (referred to as GMR_SCA3tr-Q78 in the text) was used for screening. This disease transgene codes for a carboxy-terminal fragment of the Ataxin-3 protein comprised of 12 Ataxin-3 amino acids together with a haemagglutinin (HA) tag upstream of the polyQ tract with 78 repeats. The polyQ tract is flanked downstream by the residual 43 amino acids of the protein isoform 1a. For comparison with different polyQ settings, a similar transgenic fly line expressing truncated Ataxin-3 with a normal polyQ tract of 27 repeats $(P\{w[+m C]=U A S-H s a p \backslash M J D . t r-$ Q27\}) and a full length ATXN3 fly line with 84 glutamines ( $P\{w[+m C]=U A S-S C A 3 . f l-$ Q84.myc\}) [209] were utilised.

Additionally, a fly line expressing a mutated form of human microtubule-associated protein tau gene, Tau[R406W] ( $P\{w[+m C]=U A S-h T a u[R 406 W]\}$ ) [271] was employed. By means of this unrelated disease model for frontotemporal dementia (FTD) the specificity of the obtained modifiers for polyQ-induced neurotoxicity was tested.

### 5.1.1 Phenotypes of the disease model flies

In order to assess polyQ-induced neurotoxicity and the modulation thereof, the rough eye phenotype of GMR_SCA3tr-Q78 was used as readout. The REP reflects the demise of photoreceptor neurons [219] induced by expression of the toxic gene product in postmitotic cells of the Drosophila compound eye [252]. The driver line itself served as a reference phenotype throughout the screen (Figure 6B) instead of a wild type fly (Figure 6A) with which RNAi effects alone could not be analysed. Overexpression of a non-toxic lacZ transgene (coding for $\beta$ galactosidase, $P\{w[+m C]=U A S$-lacZ.Exel $\} 2$ ) did not show vast
changes compared to the control phenotype (Figure 6C). Both featured a subtly roughened eye phenotype at $25^{\circ} \mathrm{C}$, temperature increase to $29^{\circ} \mathrm{C}$ intensified the severity of the GMRGAL4 phenotype (not shown). Neither expression of the normal length truncated Ataxin-3, SCA3tr-Q27 (Figure 6D) nor of the full-length SCA-Q84 (Figure 6F) resulted in an overt REP although SCA3-Q84 eyes were slightly roughened, comparable to lacZ phenotype. GMRmediated targeting of SCA3tr-Q78 expression to the eye however led to a severe degenerative eye phenotype as described previously [219] (Figure 6H). Pigmentation of the compound eye is greatly reduced with only minor retention of colour at the rims. Additionally, the surface texture is disturbed with disarranged sensory bristles and occasionally appearing necrotic spots and dints, hinting to degeneration of underlying eye structures. Nevertheless, eye size itself appears unaffected by the otherwise detrimental consequences of polyQ expression. Finally, overexpression of the Tau[R406W] transgene resulted in a heavy REP presenting with deranged surface and overall shrinkage of the eye, yet eye colour is retained (see Appendix Figure 1). The eye morphology of SCA3 flies visible with light microscopy is also reflected in scanning electron micrographs. Whereas the surfaces of SCAtr-Q27 and full length SCA3-Q84 fly eyes appear regular and exhibit no predominant deterioration, the eyes of SCA3tr-Q78 flies are severely compromised in structure and shape. The eye is collapsed and features disordered sensory bristles in conjunction with a disorganised ommatidial pattern. Overall collapse of the compound eye may be caused by loss of underlying retinal tissue due to SCA3-induced degeneration, a process obviously not taking place in the other two SCA3 models, at least not to the same extent.

For investigations of neurodegenerative diseases like SCA3 in flies, it would be reasonable to evaluate the effect of SCA3tr-Q78 overexpression in photoreceptor neurons, but also in other neuron populations or the entire nervous system. However, pan-neural expression of the SCA3tr-Q78 transgene results in pupal lethality and yielded no viable progeny, neither did ubiquitous Ataxin-3 production, confirming previous reports [219].


Figure 6. Phenotypes induced by GMR-mediated expression of different transgenes.
Compound eye phenotypes of wild type ( $\mathbf{A}$ ) and control flies ( $\mathbf{B}, \mathbf{C}$ ) in comparison to flies bearing neurodegenerative disease-associated transgenes (D-I). In contrast to wild type eyes (A), control GMR-GAL4 flies exhibit a subtly roughened eye surface (B) slightly worsened by expression of a lacZ control transgene (C). Expression of normal length truncated SCA3 (D) and full-length elongated SCA3 (F) transgenes induces mildly roughened eye phenotypes whereas induction of a truncated disease gene results in a heavy REP with deteriorated surface and texture of the compound eye featuring dints and necrotic spots (H). Scanning electron micrographs underpin the light microscopy findings, displaying subtle surface changes for SCA3tr-Q27 (E) and SCA3-Q84 (G) and showing seriously compromised eye morphology in SCAtr-Q78 flies (I).
Orientation of images is dorsal-up and anterior-left, all scale bars apply to $200 \mu \mathrm{~m}$ respectively.

### 5.1.2 Assessment of SCA3tr-Q78 protein expression and effects in the eye

Expression of neurotoxic truncated Ataxin-3 protein is reflected in the demise of postmitotic cells of the eye and the resulting rough eye phenotype. Detection of monomeric polyQ protein in adult flies with biochemical methods like Western blot proved to be rather complicated (compare [276], Figure 7A) since detectable protein amounts were very low (molecular mass $\sim 32 \mathrm{kDa}$ ). SDS-insoluble aggregates resided in the stacking gel of polyacrylamide gels. Higher molecular strong bands were visible between 50 and 75 kDa,
presumptively representing dimers of truncated Ataxin-3 (Figure 7A). Protein levels reached a peak shortly after hatching, decreasing drastically at seven days post eclosion (dpe) and being hardly detectable at 12 dpe. Since there is no ATXN3 fly orthologue, levels of transgene expression levels could not be compared to endogenous protein levels. The presence of SDS-insoluble truncated Ataxin-3 aggregates could also be detected in freshly hatched flies by filter retardation assay. The aggregate load in head lysates of SCA3 flies was constant on a high level throughout a period of 12 days post eclosion (Figure 7B).


Figure 7. Biochemical detection of SCA3tr-Q78 protein levels and aggregation together with verification of SCA3tr-Q78 expression, aggregation and induced cell death in larval imaginal discs.
(A) Immunoblot for SCA3tr-Q78 protein levels in flies of different age, showing weak protein signal at $\sim 32 \mathrm{kDa}$ (arrow) and higher molecular strong signals above 50 kDa just after and one day post eclosion. Aggregated protein traces are retained in the stacking gel (bar). Protein levels at 7 dpe and 12 dpe are hardly detectable. (B) Filter retardation assay of SCA3 fly head lysates exhibits strong constant signal for SDS-insoluble Ataxin-3 aggregates already present at eclosion time. (C) Detection of HA-tagged truncated Ataxin-3 in the L3 larval imaginal disc posterior (right) of the morphogenetic furrow. (D) Truncated Ataxin-3 forms aggregates in the eye discs of SCA3trQ78 L3 larvae. (E) Cell death in SCA3tr-Q78 larval eye discs detected by acridine orange staining is correlated to the expression of truncated Ataxin-3 (arrows mark morphogenetic furrow and border of cell death).
Genotype of SCA3tr-Q78 flies used: w; GMR-GAL4/UAS-SCA3tr-Q78
Orientation is anterior-left. Blue in (C), nuclei stained with DAPI; green in (C) and (D), SCA3tr-Q78 stained with mouse anti-HA antibody; green in (E), acridine orange. Scale bars in (C) and (E) apply to $100 \mu \mathrm{~m}$ respectively.

Expression of the glass multiple reporter (GMR) is initiated with the progression of the morphogenetic furrow (MF) in a presumptive eye structure, the larval imaginal eye disc, after 12 h of larval third instar (L3) [252]. The MF demarcates the border between yet uncommitted cells anterior and postmitotic neuronal progenitor cells posterior to the MF [277, 278]. Concomitantly with onset of GMR expression and thereby GAL4 activation, induction of SCA3tr-Q78 via its UAS sequence takes place. This leads to first nuclear inclusions detectable at mid-third instar and morphological defects three days later in early pupal stages [219]. Consequently, expression of expanded polyQ protein can be discerned with antibodies directed against the HA tag in eye discs of SCA3tr-Q78 L3 larvae (Figure 7C). SCA3 fly eye discs also exhibit protein aggregation and inclusions in targeted cells as already shown previously [219] (Figure 7D).

Ataxin-3 with an expanded polyQ stretch has been described to induce apoptotic cell death [213, 279, 280]. In order to detect dying cells during eye morphogenesis, the fluorescent vital dye acridine orange (AO) was utilised. The compound crosses the cellular plasma membrane and intercalates into the DNA, emitting a bright green to orange signal of the nucleus with condensed chromatin in apoptotic cells. By these means cell death in larval eye discs of polyQ flies could be detected (Figure 7E) [281] and is co-localised with the expression of truncated Ataxin-3. Thus, the basis of the observed rough eye phenotype of the adult polyQ flies already at hatching can be traced back to the expression of toxic truncated Ataxin-3 and its aggregation in nuclear inclusions already at larval stages.

Production of the truncated ATXN3 gene product continues at pupal stages and in adult flies, resulting in a REP. Frontal fly head sections reveal greatly disturbed eye morphology with degenerated retinal structures and heavy cell loss (Figure 8E) compared to highly ordered, intact retinal structure in GMR control (Figure 8A). LacZ-expressing flies (Figure 8C) reveal slightly thinner retinae, but predominantly retained morphology.


Figure 8. Histological and immunohistochemical analysis of utilised fly models.
(A) Frontal sections of GMR control fly heads stained with toluidine blue exhibit ordered dense retinal structure and (B) prove negative for expression of HA-tagged truncated Ataxin-3. (C) Retina of lacZ-expressing flies is reduced in thickness and less dense than that of control flies, however, it shows regular patterning of profound eye structures (detachment of the retina from underlying tissue is a sectioning artefact). (D) LacZ fly head sections are negative for Ataxin-3 staining. (E) Frontal sections of GMR_SCA3tr-Q78 fly heads reveal severely degenerated retina and deeper eye structures with merely no retained structured tissue. (F) Remaining cells feature heavy HA-positive Ataxin-3 inclusions.
All pictures represent central parts of the fly retina. Red in (B, D, F), F-actin stained with Alexa Fluor 568-coupled phalloidin; green in (F), SCA3tr-Q78 stained with mouse anti-HA antibody. All scale bars apply to $50 \mu \mathrm{~m}$ respectively.

Immunohistochemistry approaches exhibit strong staining for HA-tagged polyQ protein and marked aggregates in remainder of GMR-polyQ fly retinae (Figure 8F) which are neither found in GMR-GAL4 controls (Figure 8B) nor in flies expressing non-toxic lacZ (Figure 8D).

### 5.1.3 Evaluation of photoreceptor integrity

In an attempt to quantify polyQ-induced neurodegeneration of the rough eye we intended to count photoreceptor (PR) neurons, a direct target of toxic protein expression by GMR-GAL4, and assess the changes in their stereotypic distribution pattern. In control eyes, there is a trapezoid pattern of seven out of eight photoreceptors visible for each ommatidium. The seventh and eighth PRs are located on top of each other and thus cannot be visualised separately. Neurodegeneration as a consequence of polyQ toxicity is destined to diminish photoreceptor count. Semi-thin sagittal sectioning of the eye was used for evaluation of photoreceptor integrity after histological staining.


Figure 9. Photoreceptors in semi-thin sections of SCA3 disease models.
(A) Photoreceptors of control flies appear in an ordered fashion with a regular rhabdomere count of seven. (B) Truncated SCA3 gene expression results in severely degenerated eye tissue with hardly any PR neurons left and vast cell-free areas. (C) In flies with full-length expanded SCA3 expression, ommatididal structure and PR count is predominantly retained as in the control with slight loosening up of overall tissue structure. GMR-GAL4 driver (A) was used to activate expression ( $\mathbf{B}, \mathbf{C}$ ) and driver-only served as control. Insets show detailed a view of semi-thin sections of compound eyes. All scale bars apply to $50 \mu \mathrm{~m}$.

Counting of photoreceptors in the GMR-GAL4 control flies resulted in an if atl slightly reduced number of visible neurons, mainly probably due to mild GAL4 toxicity. However, photoreceptors with toxic elongated polyQ expression exhibit severe neuronal loss and eyes present with overt lack of ommatidia themselves. In contrast to the truncated protein, full-length Ataxin-3 with 84 glutamines exhibited no decrease in rhabdomere number compared to the GMR control. Only the strict trapezoid pattern of the photoreceptor neurons was distorted to some degree. Concluding from this one can say that the exterior rough eye phenotype of disease flies indeed has its origin in the degeneration of photoreceptor neurons in the single ommatidia of the compound eye.

### 5.2 Modifier screen for polyQ-induced neurotoxicity

In order to conduct an RNAi-based screen for modifiers of polyQ-induced neurotoxicity, a subset of RNAi fly lines from the Vienna Drosophila RNAi Centre (VDRC) was obtained, comprised of 7,488 lines corresponding to 6,930 different genes. To our knowledge this represents the largest number of genes investigated in a modifier screen in Drosophila so far. The RNAi lines were chosen by the VDRC as silencing fly orthologues to human genes (termed human orthologue RNAi sublibrary). These RNAi library strains were subjected to consecutive steps of screening to reveal genetic interactors of truncated

Ataxin-3 protein in the course of SCA3 pathogenesis modelled in Drosophila eyes (summarised in Figure 10).


Figure 10. Flow chart of the implemented screen to identify modifiers of SCA3-induced toxicity including subsequent analysis of primary screen candidates.

The screening process is depicted, including the results of each consecutive screening step explained in chapter 5.2.

### 5.2.1 Screen for unspecific RNAi effects in control flies

To exclude RNAi lines from subsequent screening that per se induce a change of external eye structures, the complete RNAi sublibrary was crossbred with the GMR-GAL4 driver line. RNAi lines subtly or obviously worsening the eye appearance of the GMR-GAL4 line in the F1 generation were not considered for screening as gene silencing in this case apparently has deleterious effects apart from expression of a toxic protein. According to this paradigm, 844 RNAi lines were excluded from further investigation due to their modification in control (Figures 9, 11).

### 5.2.2 Primary screen for polyglutamine modifiers

Subsequent to exclusion of effectors in control flies, the primary screen was conducted by crossbreeding the remaining 6,644 RNAi lines with the GMR_SCA3tr-Q78 flies. As a result, F1 flies co-expressed truncated Ataxin-3 with 78 polyglutamine repeats and the respective shRNA. Modification of REP was assessed in the F1 generation with respect to
change of severity of degeneration, pigmentation and overall eye morphology. Potential candidates exhibiting a modulation of REP after the primary screening were subjected to dual rescreening for verification. Out of all lines investigated, 6,115 did not show any change in polyQ-induced rough eye phenotype and were therefore not considered as candidates. 529 RNAi lines exhibited an overt change of the polyQ-induced REP, as obvious suppression or enhancement was observed. In case SCA3tr-Q78 expression in combination with gene silencing yielded no viable offspring, this was considered as a lethal enhancement (Figure 11). It is reasonable to assume that alterations of the REP in either direction reflect amelioration or increment of Ataxin-3-induced toxicity.


Figure 11. Modification of the SCA3tr-Q78-induced phenotype by enhancing and suppressing candidate RNAi lines.

Only obvious alteration of the SCA3 REP in either direction was considered. It was assumed that modification of the screen REP by knockdown of a candidate gene reflects amelioration or increase in polyQ toxicity in affected cells respectively.

Expression of 36 shRNAs in SCA3 flies led to a suppression of the phenotype in F1 generation (obvious amelioration of REP or WT-like eye), but the overwhelming majority of modifiers were enhancers with 493 RNAi lines leading to an obviously worsened phenotype or no offspring at all. Knockdown of 457 genes out of these 483 enhancing candidates lead to a lethal outcome after crossbreeding with SCA3tr-Q78 flies (complete list of modifier RNAi lines in Appendix Table 1). Naturally it was not possible to analyse these candidates morphologically in further detail and focus was put on the other candidates, especially on the suppressors (modifier RNAi lines with viable progeny listed in Table 9).

In one suppressor case, silencing of the same gene (Hsc70-4) by two different RNAi lines (transformant IDs 26465 and 50222) yielded suppression of REP for both strains.

RNAi lines modifying SCA3tr-Q78-induced REP and having viable progeny (Table 9) were categorised according to their GO term (process) as proposed by the Gene Ontology Annotation Database [282]. Categories and respective number of modifier lines are as follows: protein folding and stress response (7), transcription/chromatin modification (7), nucleic acid metabolism (9), transport and secretion (4), signalling (6), lipid metabolism (4), ubiquitin- and proteasome-related pathways (4), development, differentiation and cell death (9), miscellaneous (15) and unknown function (7) (Table 9, Figure 12). Lethal candidate lines were not categorised, yet selected candidates were utilised for computational analysis (data not shown) and are discussed in chapter 6.

Apart from the primary candidates utilised for further analysis, a total of 1,002 RNAi lines resulted in subtle modification of the REP, 217 of them subtle suppressors and 785 subtle enhancers. This group was not investigated beyond this point due to unclear origin of the modification and possible interindividual differences in the eye phenotypes. However, subtle candidates might prove helpful for computational analysis.

Table 9. List of candidates with viable progeny modifying Ataxin-3-induced REP in Drosophila.

| Transformant ID ${ }^{1}$ | Drosophila gene $^{2}$ | Human orthologue ${ }^{3}$ | Process ${ }^{4}$ | $\Delta$ SCA3 REP ${ }^{5}$ |
| :---: | :---: | :---: | :---: | :---: |
| Protein folding and stress response |  |  |  |  |
| 26465 | Hsc-70-4 | HSPA1L | Stress and unfolded protein response | S |
| 50222 | Hsc70-4 | HSPA1L | Stress and unfolded protein response | S |
| 23637 | Droj-2 | DNAJA4 | Protein folding | S |
| 45596 | Hsc70-1 | HSPA8 | Stress response | S |
| 41696 | Hop | STIP1 | Stress response | S |
| 33581 | CG2887 | DNAJB1P1 | Protein folding (D) | E |
| 48692 | Hsf | LOC644383 | Response to heat (D), | E |
| Transcription/Chromatin modification |  |  |  |  |
| 11219 | RpII15 | POLR2I | Transcription | S |
| 3780 | dve-s | GBX1 | Transcription regulaton | S |
| 41530 | Brd8 | BRD8 | Transcription regulation | S |


| 6282 | EloA | TCEB3 | Transcription regulation | S |
| :--- | :--- | :--- | :--- | :--- |
| 43802 | MRG15 | MORF4L1 | Transcription regulation | S |
| 28386 | salr | SALL1 | Transcription regulation | E |
| 5684 | $c h m$ | KAT7 | Chromatin modification | E |

## Nucleic acid metabolism

| 34713 | CG3808 | TRMT2A |
| :--- | :--- | :--- |
| 26475 | CG4266 | SCAF8 |
| 43870 | DNApol- $\mathbf{\alpha 5 0}$ | PRIM1 |
| 36025 | tsu | RBM8A |
| 31777 | CG13298 | SF3B14 |
| 23659 | Smg5 | SMG5 |
| 24725 | CG3225 | DHX35 |
| 24070 | CG9601 | PNKP |
| 10942 | Gnf1 | RFC1 |

## Transport and secretion

| 33262 | CG5687 | SLC5A8 |
| :--- | :--- | :--- |
| 20536 | CCS | CCS |
| 20183 | Cha | CHAT |
| 8620 | CG4288 | SLC17A5 |


| Ion transport | S |
| :--- | :--- |
| Metal ion transport | E |
| Neurotransmitter secretion | E |
| Transmembrane transport | E |

Signalling

| 8780 | CG17048 | RASGRP1 | Ras protein signal transduction | S |
| :---: | :---: | :---: | :---: | :---: |
| 25030 | 5PtaseI | INPP5A | Cell communication | S |
| 31257 | Gbeta13F | GNB1 | G protein coupled ACh receptor signalling pathway | S |
| 1326 | AR-2 | KISS1R | G protein-coupled receptor signalling pathway | E |
| 36153 | CG34372 | DEF6 | G-protein coupled receptor signalling pathway (D) | E |
| 32370 | stai | ODZ3 | Signal transduction | E |
| Lipid metabolism |  |  |  |  |
| 8070 | bwa | ACER2 | Lipid metabolism | S |
| 30186 | CG15534 | SMPD1 | Sphingomyelin catabolic process | E |
| 42798 | Dnz1 | ZDHHC3 | Protein palmitoylation | E |
| 10020 | Spt-I | SPTLC1 | Sphingolipid biosynthesis | E |

Ubiquitin- and proteasome-related pathways

| 37221 | CG9153 | HERC4 |
| :--- | :--- | :--- |
| 24030 | Trbd | ZRANB1 |
| 43606 | CG6758 | FBXO42 |
| 37930 | CG14619 | - |

## Development, differentiation and cell death

| 23121 | LanB1 | LAMB2 |
| :--- | :--- | :--- |
| 16040 | Hrb27C | DAZAP1 |
| 35147 | l(3)neo38 | ZNF541 |
| 47569 | CG12935 | TMEM223 |
| 21293 | CG31048 | DOCK3 |
| 19450 | CG15399 | CHODL |
| 41960 | Exn | NGEF |
| 33837 | Pkcdelta | PRKCD |
| 13005 | Dab | - |


| Cell morphogenesis | S |
| :--- | :--- |
| Differentiation | S |
| Cell differentiation | E |
| Nervous system development | E |
| (D) | E |
| Axonal outgrowth | E |
| Muscle organ development | E |
| Apoptosis | E |
| Apoptosis | S |
| Differentiation/Neurogenesis |  |
| (D) |  |

## Miscellaneous

| 7903 | ppk14 | TREM2 | Axonal guidance | S |
| :---: | :---: | :---: | :---: | :---: |
| 17196 | DCX-EMAP | CYP2E1 | Steroid metabolic process | S |
| 16182 | aux | GAK | Cell cycle | S |
| 8408 | Cad88C | CDH7 | Cell-cell adhesion | S |
| 44362 | slmo | SLMO2 | Spermatogenesis (D) | S |
| 19066 | Doa | CLK2 | Protein phosphorylation | S |
| 22590 | timeout | TIMELESS | Mitosis | E |
| 24885 | DAAM | DAAM2 | Actin cytoskeleton organization | E |
| 40478 | Marf | MFN2 | Mitochondrial fusion | E |
| 44114 | CG11722 | NDUFAF4 | Mitochondrial respiratory chain complex I assembly | E |
| 48062 | CG1695 | SGSM1 | Regulation of Rab GTPase activity | E |
| 22454 | CG6873 | ADAM12 | Cell adhesion | E |
| 30717 | CG33128 | REN | Proteolysis | E |
| 36572 | Sbp2 | SECISBP2 | Translation | E |
| 15789 | Mal-A1 | - | Carbohydrate metabolic process (D) | S |


| Unknown function |  |  |  |  |
| :--- | :--- | :--- | :--- | :--- |
| 46473 | CG17919 | LOC647307 | - | S |
| 23843 | roq | RC3H2 | - | S |
| 40006 | CG15618 | THADA | - | S |
| 40044 | CG16890 | FRA10AC1 | - | S |
| 43612 | $C G 14966$ | C15orf40 | - | E |
| 29711 | $C G 6115$ | LOC493754 | - | E |
| 49792 | $C G 3678$ | $T T C 35$ | - | E |

${ }^{1}$ As indicated by VDRC.
${ }^{2}$ Drosophila gene as listed in Gene Database of NCBI [3].
${ }^{3}$ Human orthologue according to HomoloGene Database [283] or obtained from BLAST analysis [284]. Symbol as listed in Gene Database of NCBI [3].
${ }^{4}$ Referred to as biological process of the human orthologue gene according to GO term listed in Gene Ontology Annotation Database [282], otherwise marked with D for predicted process of Drosophila gene.
${ }^{5}$ Modification of SCA3tr-Q78-induced REP after GMR-GAL4-mediated expression of shRNA.
S, Suppression; E, Enhancement (both in case of RNAi of respective gene)

### 5.2.3 Specificity of RNAi effects for SCA3tr-Q78-induced neurotoxicity

In order to narrow down the candidates found in the primary screen to those modulating specifically SCA3tr-Q78 neurotoxicity without affecting other disease proteins, RNAi silencing of modifier genes was induced in flies expressing Tau[R406W]. By comparing the polyQ screen data with results from this verification experiment with an unrelated pathogenesis model, target genes that exclusively act on polyglutamine-induced effects should be identified. Only 4 \% (21 lines) of the RNAi candidates displayed a similar modification of the Tau[R406W]-induced REP (two suppressors, 19 enhancers). These candidates were not considered specific for polyglutamine, nevertheless they are included in the set of Ataxin-3 action modifiers. Two RNAi lines (with shRNA against Drosophila genes aux and CG3808) were found to modulate Tau[R406W]- and SCA3tr-Q78-induced REPs in opposite directions, suppressing polyQ- and enhancing Tau-induced phenotypes. Due to different modulation in the two disease models, aux and CG3808 were still termed modifier candidates specific for SCA3tr-Q78 neurotoxicity.

In conclusion, we obtained a total of 508 RNAi lines (34 suppressors, 474 enhancers) corresponding to 502 Drosophila genes. Silencing of these genes is assumed to modify Ataxin-3-induced neurodegeneration in the fly compound eye (see Figures 10, 12).


Figure 12. Summary of the SCA3tr-Q78 modifier screen and overview of modifier categories.
(A) Of the 7,488 utilised RNAi lines, 844 showed disturbed external eye structures after being crossed to GMR-GAL4 and were excluded from further screening. Screening identified 34 suppressors and 474 enhancers. The remaining 6,136 lines had no impact on REP or showed similar effects in the Tau screen, hence were not considered specific for polyQ. (B) Depiction of biological processes of silenced candidate genes.

### 5.2.4 Evaluation of gene silencing by RNAi lines

Given the large number of candidate genes, it was not possible to quantify silencing of RNAi target genes on mRNA levels. Nevertheless, in an attempt to evaluate the RNAi effect of the screened fly lines in a sampled fashion, the results of ubiquitous expression of the UAS-shRNA were assessed. For this experiment the human orthologue sublibrary was searched for genes that have been reported in the literature to be crucial for survival. These amounted to 59 lines representing 54 genes subsequently crossbred with the actin5C-GAL4 driver line ( $P\{w[+m C]=A c t 5 C-G A L 4\} 25 F 01$ ). It was assumed that effective silencing of target genes would lead to a reduced number or complete lack in offspring due to vital importance of the downregulated genes.

Ubiquitous silencing of this set of essential genes indeed eventuated in a lethal outcome in F1 generation for 45 of the 59 tested RNAi lines (76.2 \%), the remainder showing a reduced number of offspring (summarised in Appendix Table 2). Additionally, select suppressor candidates from the RNAi screen were able to rescue lethality in offspring pan-neurally expressing SCA3tr-Q78 (data not shown). Concluding from these results it was assumed that the majority of the RNAi lines yield effective silencing of their target genes.

### 5.3 Impact of modifiers on polyQ toxicity and aggregation

The modulation of the exterior polyQ-induced rough eye phenotype served as a readout of neurotoxicity and its modulation by modifier candidates. Additionally, integrity of eye structure and the connection between aggregation and neurodegeneration can be analysed in greater detail by histological and immunohistochemical approaches on the one hand and biochemical filtration methods on the other hand. We utilised both for certain candidate genes in order to gain insight into the modes of action of the discovered modifiers.

### 5.3.1 Evaluation of tissue integrity of SCA3tr-Q78-shRNA-coexpressing flies

A suitable approach to study morphology and thereby integrity of the compound eye is frontal sectioning of fly heads and the subsequent histological staining of the tissue. The morphological changes of the eye surface of polyQ flies originate from the degeneration of the underlying tissue of the compound eye. Namely, photoreceptor and adjacent cell architecture are severely deteriorated, leading to the overt rough eye phenotype and eventually collapse of the eye.

Selected suppressor candidates were used in order to assess preservation of deeper eye tissue in contrast to retinal damage in polyQ flies. Flies co-expressing polyQ protein and enhancer RNAi could not be analysed due to severe degeneration of eye structure. As expected, silencing of genes leading to improvement of REP was also able to alleviate the detrimental effects of polyQ expression in the retina, albeit to different degrees (Figure 13). For example, the two RNAi lines for knockdown of Hsc70-4 (Transformant IDs $50222 / 26465$ ) both ameliorated cell demise in the retina of SCA3 flies in line with their suppression effect on REP in the RNAi screen. Nevertheless, the effect of tissue preservation was significantly different for these shRNAs, with line 50222 showing almost wild-type retinal extend and structure whereas line 26465 exhibited good external phenotype mitigation yet retinal organisation presented diffuse and width was decreased.


Figure 13. Influence of selected shRNAs on tissue integrity of SCA3tr-Q78 fly head sections.
Control sections feature intact retinal tissue in GMR-GAL4 and serious degeneration of eye tissue in SCA3tr-Q78 fly sections. Following introduction of shRNA lines suppressing Ataxin-3-induced REP, retinal thickness is improved to different degrees and retinal tissue architecture is restored towards GMR control situation.
All scale bars apply to $50 \mu \mathrm{~m}$ respectively.

### 5.3.2 Filter retardation analysis of RNAi influence on polyQ aggregates

Due to the proposed toxicity of certain polyQ aggregate species and the possible influence of modifier gene knockdown thereon, it was intended to biochemically study whether there is an impact of silencing of screened modifier genes on the levels of SDSinsoluble polyQ aggregates as previously described [276].
In order to address this question, expanded polyQ protein was co-expressed together with the candidate shRNA in the eye. Where possible, offspring were collected and SDS-treated head lysates were subjected to filter retardation assay (FRA) analysis. Filtration of the protein lysates through a nitrocellulose membrane would lead to trapping of aggregates exceeding a certain size ( $0.2 \mu \mathrm{~m}$ ), allowing for assessment of polyQ aggregate load. Of course, lethal enhancers could not be analysed due to the absence of viable progeny. The hypothesis was that REP-suppressing candidates would decrease toxic aggregate levels, whereas enhancer shRNA expression would result in higher aggregate load. Nevertheless only a minor number of suppressor candidates was observed to effectively ameliorating aggregate number (6/34, 17.6 \%) with shRNA against CG3808 being the most potent aggregation suppressor. On the contrary, a considerable large group did not change
aggregate levels significantly or even enhanced (2/34, $5.8 \%$ ) the cellular polyQ aggregate burden after normalisation to polyQ control. Additionally, the majority of enhancer RNAi candidates exhibited a trend towards decreasing aggregate levels with only a few gene knockdowns resulting in higher aggregate load. However, absolute number of significant changes is smaller in enhancers compared to suppressors. Concluding from these results, aggregate levels in this experimental setting do not appear to correlate to exterior REP and vice versa (Figure 14).


Figure 14. Analysis of SDS-insoluble SCA3tr-Q78 aggregate load with shRNA modifiers.
(A) Exemplary filter retardation analysis for visualisation of aggregate load. GMR-GAL4 control is negative, SCA3trQ78 lysates exhibit heavy aggregation which is mitigated by suppressor shRNA. (B) Densitometric measurement of filter retardation analysis compared to SCA3tr-Q78 for suppressors and enhancers of polyQ-induced toxicity. $\mathrm{n} \geq 3$ if not indicated otherwise. Significant changes are: *p<0.05; ** $p<0.01$ : *** $p<0.001$.

### 5.3.3 RNAi effects on polyQ inclusions in situ

Expression of expanded polyQ protein leads to formation of protein aggregates in the compound eye as shown before and verified biochemically by the filter retardation experiments. Aggregation of toxic gene products is a hallmark of polyQ disease and considered to be at least in part causative for neurotoxicity and degeneration. On a microscopic level, inclusion bodies in retinal cells are detectable, presumably consisting of diverse polyQ aggregate species and various other proteins recruited to the agglomerate. In the eyes of the offspring of polyQ flies and flies concomitantly expressing modifier shRNA we intended to address the question whether improvement or worsening of the REP corresponds to the aggregate load in situ.

In the frontal head sections representative for select candidates we were however not able to show a robust connection between decrease of inclusions in the eye tissue and change of the REP. Three of the analysed RNAi lines featured a reduction of SDS-insoluble aggregates in filter retardation assays (Brd8, CG17919, CG33128, Figure 15A-C) with the latter being an enhancer of the REP. The two suppressor lines still featured inclusions, however to a seemingly decreased amount. The CG33128 shRNA (Figure 15C) led to an enhanced number of inclusions and concomitantly had the worst tissue integrity. The suppressor lines at least presented with improved retinal morphology compared to polyQ alone. Two lines with increased SDS-insoluble aggregate load in filter retardation analysis, CG17048 and Hsc70-4 (26465) (Figure 15D, E), had clearly delimited small inclusions in moderate numbers in parallel with an overall well-preserved tissue integrity. Thus, there was no clear trend such that modified eye structure and therefore altered neurotoxicity have their origin in a differential number of immunohistochemically detectable aggregates.


Figure 15. Influence of RNAi on microscopically detectable Ataxin-3 inclusions in situ.
RNAi effects differently affect inclusions of SCA3tr-Q78 protein in fly eyes. Whereas two out of the group of suppressors of SDS-insoluble aggregates had diffusely demarcated and seemingly less inclusions (A, B), the third one appeared to have high inclusion numbers and disturbed retinal morphology (C). Aggregate enhancers in FRA exhibited distinct small inclusions in moderate numbers, tissue integrity is well preserved (D, E).
Blue, cell nuclei stained with DAPI; green, SCA3tr-Q78 stained with mouse anti-HA antibody. All scale bars apply to 50 $\mu \mathrm{m}$ respectively.

### 5.4 Summary of RNAi screen results

We conducted a large-scale RNAi screen in Drosophila in order to identify genes that if silenced are capable of modifying polyQ toxicity. After excluding vitally crucial genes from the analysis, our efforts resulted in a set of 502 candidate genes. Knockdown of the vast majority of interactors together with elongated polyQ expression led to lethality in the progeny and pre-empted further investigations. Nevertheless, we were able to analyse several of 68 non-lethal candidates in more detail.

Histological and immunohistological evaluation revealed that suppressor candidates are to a certain degree capable of ameliorating the degeneration of eye tissue and photoreceptors responsible for the REP. However, the amount of exterior improvement of the REP is not consistently reflected in the preservation of the underlying eye structures. Tissue of enhancing candidates is rendered impossible to investigate since degeneration of the eye tissue is already too severe upon polyQ expression alone. Additionally, examination of aggregate formation in the eye of elongated polyQ-expressing flies showed heavy inclusion load in SCA3tr-Q78 flies. Improvement of REP by suppressors did not correlate to
their capability to prevent inclusion body formation. Contrary to our assumption, the improvement or aggravation of the REP by the modifiers could not be conclusively explained by their impact on SDS-insoluble aggregates in filter retardation analysis.

As a result of the consistently beneficial outcome of its silencing for polyQ toxicity in histological as well as biochemical testing, we chose the Drosophila gene CG3808, orthologous to the human tRNA methyltransferase homologue $2 A$ (TRMT2A), for subsequent analysis.

### 5.5 Analysis of the effect of TRMT2A silencing on polyQ toxicity in Drosophila

Silencing of CG3808, the Drosophila orthologue of TRMT2A, showed promising results in ameliorating the detrimental effects of elongated SCA3 protein. Thus, we addressed the question whether CG3808 knockdown would also prove beneficial during more detailed analysis and in other polyQ models apart from SCA3tr-Q78. Firstly, we assessed the capability of the RNAi to overcome lethality induced by pan-neural expression of SCA3tr-Q78. Indeed, co-expression of shRNA and SCA3tr-Q78 in all neural cells resulted in viable progeny with no overt abnormalities. Additionally, ubiquitous silencing of CG3808 by the means of RNAi did not render the offspring fatal and had no negative influence on overall life time of the respective flies, demonstrating that the protein is not of vital importance in Drosophila (data not shown).

For evaluation of CG3808 RNAi effects we again utilised histological and biochemical methods together with assessment of longevity. Finally, we transferred experiments to a cellular model in an attempt to verify the progress accomplished in Drosophila in a mammalian model.

### 5.5.1 Impact of TMRT2A silencing on polyglutamine-induced REPs

Expression of SCA3tr-Q78 in the compound eye led to a rough eye phenotype visible with light (Figure 16A) as well as scanning electron microscopy (SEM, Figure 16B). The SEM findings are in line with results obtained from REP pictures, namely the eyes presenting with heavily disarranged surface and even collapsed eye morphology probably due to underlying tissue degeneration. Silencing of CG3808 by RNAi led to amelioration of SCA3tr-Q78-induced REP with restored patterning and morphology of the exterior eye
surface. For another polyQ fly model, inducing a REP with exon 1 of the huntingtin gene under GMR control, similar observations were made. This htt transgene contains 97 glutamine repeats ( $w[*] ; P\{w[+]=U A S-Q 97 e x 1\} K 6,9,15 R$ ) and downregulation of CG3808 expression was sufficient to rescue the Htt-induced REP (Figure 16C, D), reversing the phenotype to almost wild type (Figure 16H). This was underpinned by SEM analysis, exhibiting a predominantly ordered surface without signs of degeneration (Figure 16I). Introduction of shRNA against CG3808 into a model for SCA1 with full-length ATXN1 Q82 expression ( $y[1] w[118] P\{[+]=U A S-S C A 1.82 Q\}[F 7]$, Figure 16E) yielded improvement of the degenerative eye phenotype to great extend as well (Figure 16J).

Concluding, knockdown of CG3808 expression is obviously capable of exerting neuroprotective effects in the course of eye degeneration caused by several different polyQ proteins.


Figure 16. Rescue of polyQ-induced REP by shRNA against CG3808.
Elongated polyQ proteins responsible for SCA3 (A, B), HD (C, D) and SCA1 (E) induced an REP in flies visible by light and scanning electron microscopy. Induction of shRNA directed against CG3808 transcripts mitigates this REP to almost wild type situation ( $\mathrm{F}-\mathrm{J}$ ).
All scale bars apply to $200 \mu \mathrm{~m}$.

### 5.5.2 Evaluation of photoreceptor integrity of polyQ flies with TRMT2A knockdown

Semi-thin sagittal sections of fly eyes expressing variants of Ataxin-1 and Ataxin-3 under control of GMR-GAL4 (GMR_SCA1 Q82 and GMR_SCA3tr-Q78 respectively) display severe degeneration of photoreceptor neurons as a consequence of polyQ neurotoxicity. Co-expression of shRNA against CG3808 on the contrary was capable of ameliorating the detrimental effects in the eye (Figure 17A). Quantification of photoreceptor neurons per ommatidium showed a severely decreased PR count in GMR_SCA3tr-Q78 (1.16 $\pm 0.06)$ flies and a moderately decreased one ( $4.24 \pm 0.27$ ) in the GMR_SCA1Q82 model. Silencing of CG3808 rescued PR degeneration almost to the level of GMR control conditions (6.85 $\pm$ $0.03)$ in GMR_SCA3tr-Q78 (6.31 $\pm 0.09)$ and GMR_SCA1 $Q 82(6.64 \pm 0.05)$ models (Figure 17B). Additionally, stereotypic patterning in the ommatidia was visible again in GMR_SCA3tr-Q78 flies in combination with CG3808 silencing. These results could be recapitulated also in the flies expressing the elongated full-length form of Ataxin-1 (GMR>SCA1 Q82, Figure 17A) and at least qualitatively with a transgene of exon 1 of HTT with 97 glutamine repeats (GMR_HTT Exon1 Q97, not shown). Therefore, silencing of CG3808 seems to have a strong neuroprotective effect opposing polyQ toxicity in the Drosophila eye.


Figure 17. Evaluation of photoreceptor integrity in polyQ flies with CG3808 RNAi.
(A) Depiction of PR neuron degeneration in polyQ models for SCA3 and SCA1 (upper panel) and rescue of number and patterning of PRs by silencing of CG3808 via RNAi (lower panel). (B) Quantification of PR number in SCA3 and SCA1 fly models compared to GMR control. Significant PR loss was rescued to a great extend by expression of shRNA against CG3808.
All scale bars in (A) apply to $50 \mu \mathrm{~m}$ respectively. Kruskal-Wallis test with Dunn's Multiple Comparison test was used for statistics in (B), significant changes are: ${ }^{* * *}$ p < 0.001; n.s., not significant.

### 5.5.3 Assessment of adult-onset polyQ fly longevity

In order to more closely mimic the disease situation in humans with late onset and progressive degeneration, further experiments in a pan-neural adult-onset model for polyQ diseases were performed. Therefore an elav-GAL4 fly strain with additional ubiquitous expression of the temperature-sensitive yeast transcriptional repressor GAL80 ${ }^{\text {ts }}$ ( $P\{w[+m W . h s]=G a w B\} e l a v[C 155] ; ~ P\{w[+m C]=t u b P-G A L 80[t s]\} 20)$ [285] (referred to as elavGAL80 in the text) was used. GAL80 is a competitor of GAL4 in binding to the UAS without activating properties, thereby preventing subsequent induction of gene expression at permissive temperature ( $\leq 20^{\circ} \mathrm{C}$ ). Upon shifting to restrictive temperature ( $\geq 25{ }^{\circ} \mathrm{C}$ ), GAL80 is unfolded, which prevents blockage of GAL4, consequently allowing GAL4-UAS binding and gene expression. Protein aggregation analysis and longevity experiments were performed making use of this system, facilitating pan-neural expression of SCA3tr-Q78 only following temperature shift from $18{ }^{\circ} \mathrm{C}$ to $29^{\circ} \mathrm{C}$.

Induction of polyQ expression could be shown in Western blot experiments climaxing four days post temperature shift and declining afterwards probably due to cell demise (Figure 18A). Aggregation of truncated Ataxin-3 was shown to be absent before induction of SCA3tr-Q78 expression on permissive temperature and to increase rapidly within a timeframe of 7 days after temperature shift (Figure 18B, C).

Overall lifetime of polyQ-expressing flies is a feasible tool for evaluation of toxicity and neurodegeneration. SCA3tr-Q78 flies showed no abnormalities at restrictive temperature due to absent toxic protein expression. However, locomotive abilities of the flies deteriorated fast after induction of expression concomitantly with a rapid decline in survival time resulting in a median survival time (timepoint when $50 \%$ of flies of overall flies are still alive) of only 10 days. Flies with adult-onset expression of a non-toxic control transgene ( $P\{w[+m C]=U A S$-eGFP $\}$ ) had an almost three times longer mean survival (27 days). Eventually, co-expression of SCA3tr-Q78 and shRNA against CG3808 increased median survival significantly to about 18 days. Therefore, silencing of this methyl transferase proved to be beneficial in alleviating detrimental polyQ effects on longevity, despite limitations compared to non-polyQ transgene expression.


B

c


D


Figure 18. Adult-onset model of SCA3tr-Q78 in Drosophila and extension of polyQ fly life time by CG3808 RNAi.
(A) Protein levels of truncated Ataxin-3 in adult-onset fly model are detectable one day post induction (dpi) by temperature shift and increase until 4 dpi . At 7 dpi , levels have already declined. (B, C) Aggregate load of SCA3tr-Q78 in adult-onset fly heads increases steadily after induction over a course of 7 days. (D) Expression of shRNA against CG3808 is sufficient to significantly prolong median survival and overall lifetime of pan-neural adult-onset SCA3trQ78 flies, although not to control levels (eGFP).
Log-rank test was used for statistics in (D), significant changes are: ${ }^{* * * *} \mathrm{p}<0.0001$.

### 5.5.4 Influence of CG3808 downregulation on aggregate formation in Drosophila

As already demonstrated, targeting of SCA3tr-Q78 expression to the eye leads to aggregate formation and gives rise to inclusion bodies of elongated polyQ protein and eye degeneration. By co-expression of shRNA against CG3808 the anti-aggregation properties of this gene knockdown could be shown in situ and in filter retardation analysis.

Paraffin frontal fly head sections were probed with an antibody directed against the HA-tag of the polyQ protein. It could be observed that induction of CG3808 RNAi is capable
of preventing assembly of inclusion bodies in the compound eye retina, concomitantly preserving the structure and architecture of the tissue to great extend. Additionally, aggregation of elongated full-length Ataxin-1 in the retina and impact of CG3808 induction thereon was estimated. Ataxin-1 did not show pronounced formation of inclusion, yet rather was localised to the nucleus. CG3808 RNAi did not feature an obvious change of Ataxin-1 distribution or amount, however, retinal structure appeared improved (Figure 19A).


Figure 19. Overview of anti-aggregation effects of CG3808 RNAi in different polyQ models and settings.
(A) Induction of CG3808 RNAi leads to a prominent decrease of inclusion number in the retina of SCA3 model flies (upper row). Ataxin-1 protein does not seem to form inclusion in SCA1 flies and CG3808 RNAi does not influence distribution or protein amount in the retina. (B) Adult-onset co-expression of CG3808 shRNA with SCA3tr-Q78 ameliorates aggregate load in fly head lysates also compared to an RNAi control (white shRNA). (C) Quantification of aggregate load in adult-onset SCA3 flies after introduction of control and CG3808 shRNA.
Scale bar in (A) applies to $50 \mu \mathrm{~m}$. Red in (A), F-actin stained with Alexa Fluor ${ }^{\circledR} 568$-linked phalloidin; green in (A) upper row, SCA3tr-Q78 stained with mouse anti-HA antibody; lower row, SCA1 Q82 stained with mouse anti-polyQ antibody. t-test was used for statistics in (C), significant changes are: ${ }^{*} \mathrm{p}<0.05 ;{ }^{* *} \mathrm{p}<0.01$; n.s., not significant.

The potent aggregate-reducing capacity of CG3808 downregulation has already been shown for eye-expressed polyQ protein (see chapter 5.3.2, Figure 14 B). Nevertheless, expression of SCA3tr-Q78 and of CG3808 shRNA under GMR control does not reflect the pathogenic situation in humans with respect to late disease onset. Therefore, elav-GAL80 fly strains for pan-neural adult-onset expression were utilised. Induction of polyQ expression alone by temperature shift produced a significant increase in SDS-insoluble aggregates five days post induction as detected by filter retardation assay (Figure 19B, C). Introduction of a control shRNA against white gene expression also showed a significant rise in aggregate load, whereas the moderate increase in flies expressing both polyQ and CG3808 RNAi was not statistically significant (Figure 19C). From these findings one can conclude that silencing of CG3808 expression is capable of decelerating the formation and/or accumulation of potentially toxic polyQ aggregates.

### 5.6 Impact of TRMT2A knockdown on polyQ toxicity in a mammalian system

All experiments so far have been conducted in fly polyQ models with shRNA targeting the expression of the fly orthologue CG3808 of the human TRMT2A gene. Unfortunately, there are no classical loss-of-function alleles of CG3808 available. In addition, the lack of independent RNAi lines to silence CG3808 prevented us from confirming our findings. Although Drosophila proved to be a feasible and beneficial tool for analysis of polyQ modifier genes, it is of crucial importance to confer the insights gained regarding amelioration of aggregation and toxicity to a mammalian system. Moreover, reconfirmation of the beneficial effects of TRMT2A silencing in the context of polyQinduced toxicity in a vertebrate system would be desirable. Demonstrating the favourable activity of TRMT2A silencing in polyQ diseases would eventually deduce a universal mechanism conserved between flies and vertebrates, highlighting the experimental rational of our screen.

### 5.6.1 Generation of stable TRMT2A knockdown HEK cells

For the cell culture experiments human embryonic kidney cells (HEK293) were utilised. The high transfection efficiency and general robustness regarding both growth and protein production rendered HEK cells a feasible model system for the polyQ investigations.

For stable silencing of TRMT2A expression, five different shRNA lentiviral transduction particles, targeting individual human TRMT2A mRNA sequences, were purchased for treatment of HEK293 cells. Additionally, one non-target shRNA control viral strain was used, coding for an shRNA without any known cellular targets. Subsequent to viral transduction (at Department of Biochemistry, University Medical Centre Aachen) with different multiplicities of infection (MOI), cell colonies having the shRNA stably integrated in their genome were selected. Western blot analysis was utilised for evaluation of successful TRMT2A downregulation. Viral strains \#856 and \#1574 exhibited almost complete silencing of TRMT2A expression, regardless of deployed MOI. Strains \#736 and \#1502 induced slight downregulation of expression, whereas transduction with strain \#1485 did not result in overt changes of expression levels. Scrambled shRNA viral transduction had no impact on TRMT2A protein levels and proved to be adequate as control. All expressional levels were compared to the amount of $\beta$-tubulin as control (Figure 20A).

Consequently, cells transduced with strains \#856 and \#1574 featured feasible prerequisites for further experiments regarding polyQ toxicity in a mammalian model system. Eventually cells with stable TRMT2A knockdown (derived from infection with strain \#1574) were used (Figure 20B, C). Additionally, TRMT2A silencing was confirmed by mass-spectrometric analysis (see chapter 5.7).

A


B


C


Figure 20. Stable shRNA-mediated silencing of TRMT2A expression after viral transduction of HEK293 cells.
(A) Subsequent to transduction of shRNA against TRMT2A by lentiviral particles, protein levels of viral strains \#856 and \#1574 were reduced most efficiently of all tested lines. Non-target shRNA control showed no marked change in TRMT2A protein levels. (B) Exemplary Western blot of decreased TRMT2A protein levels in ultimately utilised line \#1574 HEK cells compared to scrambled shRNA control. (C) Quantification of TRMT2A protein levels in line \#1574 HEK cells compared to control.
t-test was used for statistics in (C), significant changes are ${ }^{* *} \mathrm{p}<0.01$.

### 5.6.2 Transfection of stable TRMT2A knockdown cells with polyQ constructs

For replication of Drosophila results in mammalian cells, the impact of TRMT2A knockdown on polyQ aggregation was investigated. Therefore, stably transduced HEK293 cells were transfected with different huntingtin constructs harbouring either a 25 repeats polyQ tract (GFP-HttQ25) or a pathological tract of 103 glutamines (GFP-HttQ103, both kind gift by Jan Senderek, ETH Zürich). PolyQ protein expression and aggregation could be visualised and monitored via a carboxy-terminal GFP-tag and fluorescence microscopy. Whereas normal Huntingtin was equally distributed throughout the cytoplasm (Figure 21A, upper row), the expanded polyQ tracts rendered the protein prone to
aggregation, resulting in peri- or intranuclear inclusions (Figure 21A, lower row). For expression of mutant polyQ constructs, protein aggregation or cell toxicity no obvious discrepancies could be discerned between control and knockdown cells. Nevertheless, quantification of GFP-positive cells with inclusions showed a slight, however significant increment for inclusion bodies (Figure 21B) from control cells (Figure 21A, left column) to the most potent knockdown cell line, \#1574 (Figure 21A, right column).


Figure 21. Aggregation properties of normal and expanded Huntingtin in control and TRMT2A knockdown HEK cells.
(A) Transfection of HEK cells with normal GFP-tagged Huntingtin (Q25, upper row) led to cytoplasmic distribution of polyQ protein in control and knockdown cells. Expanded Huntingtin (Q103, lower row) forms prominent inclusion bodies in control and knockdown cells alike (detailed view in inset right lower row). (B) Significant increase in the fraction of transfected TRMT2A knockdown cells bearing inclusion bodies compared to control.
All scale bars in (A) apply to $50 \mu \mathrm{~m}$. t-test was used for statistics in (B), significant changes are * $\mathrm{p}<0.05$.

### 5.6.3 Investigation of aggregation in polyQ-transfected knockdown cells

Apart from microscopic evaluation, polyQ aggregation under the influence of TRMT2A knockdown was also investigated biochemically. Utilising the filter retardation assay, the formation of SDS-insoluble aggregates of two different huntingtin constructs was assessed in control and knockdown cells. One construct was the afore utilised exon 1 Huntingtin with a GFP-tag (GFP-HttQ103), the second one expresses Huntingtin with the 590 N-terminal amino acids and a myc-tag (myc-Htt590). As expected, non-expanded polyQ protein (in both constructs Q25) did not show increased susceptibility to aggregation and was not retained on the filter membrane neither in control nor in knockdown cells (Figure 22A). Upon GFP-HttQ103 and myc-Htt590 Q97 expression, control cells faced heavy polyQ protein aggregation. In line with the Drosophila findings, TRMT2A knockdown resulted in a significant amelioration of SDS-insoluble aggregate load in HEK293 cells trapped on the membrane (Figure 22A). Therefore, TRMT2A knockdown seems to be sufficient to significantly alleviate SDS-insoluble polyQ aggregate load in mammalian cells.


Figure 22. Impact of TRMT2A knockdown on different SDS-insoluble Huntingtin aggregates.
(A) Q25-Huntingtin shows no SDS-insoluble protein aggregates, Q103 and Q97 proteins result in heavy aggregate load in control cells which is mitigated in TRMT2A knockdown cells. (B) Quantification of decrease in Huntingtin aggregate load in TRMT2A knockdown cells compared to control.
For protein detection in (A), mouse anti-GFP and mouse anti-myc antibodies were used. t-test was used for statistics in (B), significant changes are * $p<0.05$

In conclusion, silencing of TRMT2A expression in mammalian cells partially was capable of recapitulating alleviating effects on SDS-insoluble polyQ aggregates. A dissolving effect like for RNAi of CG3808 affecting in situ polyQ inclusion could not be verified. Yet it remains to be solved how the consequences of TRMT2A silencing are brought about mechanistically on a molecular level.

### 5.7 Attempts on revelation of the molecular mechanism of TMRT2A knockdown on polyQ proteins

The methylation of tRNA bases by their respective methyltransferases such as TRMT2A is an important factor during translation for processes like binding of aminoacyltRNA synthetases, aminoacylation itself and binding of the tRNA to the ribosome. Concluding from that it might be possible that silencing of TRMT2A hampers one of these processes and leads to differential translation of polyQ proteins. Exchange of a single glutamine within the polyQ stretch for a different amino acid alters the conformational properties, therefore possibly mitigating toxicity. In order to account for this possible mechanism, it was intended to analyse the peptide sequence of polyQ proteins by matrixassisted laser desorption/ionisation-time of flight mass spectrometry (MALDI-TOF MS) after silencing of TRMT2A expression. Isolation of SCA3tr-Q78 protein from CG3808silenced flies with immunoprecipitation approaches proved to be difficult due to the previously mentioned low levels of available monomeric protein. Therefore, lysates of TRMT2A knockdown HEK cells overexpressing myc-tagged Huntingtin Q25 were prepared for analysis by Fabian Hosp (Department Cellular Signalling and Mass Spectrometry, Max Delbrück Center for Molecular Medicine, Berlin, Head: Prof. Matthias Selbach). Owed to the required tryptic digestion of the proteins prior to MS analysis and the absence of appropriate motifs in the polyQ stretch, a non-elongated form of Huntingtin was chosen to allow for ionisation of the peptide. Nevertheless, it was not possible to ionise the polyQ peptide in order to analyse the mass of the polyQ stretch in MALDI-TOF experiments, which prevented indication of a putative amino acid change in the polyQ sequence. Other peptides of the huntingtin transgene product could be identified, proving occurrence of the protein in general. Assuming a generally reduced specificity in translation upon TRMT2A silencing, a replacement of glutamine in other proteins and apart from the polyQ stretch might have been an indirect endorsement of the amino acid exchange hypothesis. An investigation of glutamine modifications in the global protein content of the cell lysates yielded no particularly increased accumulation of alterations in the amino acid sequence in the TRMT2A knockdown cells.

Another possible path of verification of the assumption regarding the TRMT2A mode of action on polyQ proteins is the introduction of novel sites for proteolytical digestion as a side effect of the introduction of an erratic amino acid into the polyQ stretch.

Due to the close relationship of glutamine and glutamate and the fact that the evolvement of different tRNAs for glutamine and glutamate is relatively new in evolution, it stands to reason that the exchanged amino acid for glutamine is likely glutamate. Introduction of glutamate into the polyQ stretch would generate a target site for a glutamyl endopeptidase. In this case, treatment of polyQ stretches derived from TRMT2A knockdown cells would most probably result in fragmentation of the polyQ tract. This in turn could be visualised in Western blot analysis due to mass shift of the specific protein bands. Initial experiments with a plasmid expressing a HA-tagged expanded polyQ tract (gift from Junying Yuan, Department of Cell Biology, Harvard Medical School, Boston) in TRMT2A-silenced HEK cells have already started, but have not yielded conclusive results yet.

## 6 Discussion

### 6.1 Characterisation of the utilised polyQ Drosophila model

In an attempt to identify novel genetic modifiers of Ataxin-3-induced neurotoxicity we utilised an established Drosophila model [256]. Expression of the SCA3tr-Q78 transgene results in a truncated Ataxin-3 protein containing a polyQ stretch of 78 repeats and residual amino acids N - and C-terminally of the tract together with an N-terminal hemagglutinin tag. However, the Josephin domain and the ubiquitin-interacting motifs of the protein are lacking, thereby compromising the enzymatic activity of the truncated protein as a deubiquitinating enzyme and transcriptional repressor. Expression of the transgene in all postmitotic cells of the Drosophila compound eye exerted severe neurodegeneration resulting in alteration of the exterior eye structure, a so-called rough eye phenotype (REP). The REP is characterised by depigmentation, disturbance of texture and ommatidial pattern, dints and necrotic spot formation. These effects obviously have their origin in polyQ toxicity. The deleterious visible changes are a continuation of the heavily destructed internal eye structures featuring decreased retinal thickness and compromised tissue integrity due to a loss of cell mass. As a result of polyQ protein toxicity, the stereotypic number and pattern of photoreceptor neurons is decreased and disrupted respectively, demonstrating the feasibility and transferability of this modelling approach for neurological disorders. At the same time, the easy accessibility for evaluation is an important advantage of the eye-specific polyQ protein expression. Induction of the SCA3 transgene in all neural cells already at embryonic stages results in pupal lethality as shown before [219] and therefore cannot be utilised in assessment of polyQ effects and modifier screening.

Changes in polyQ-induced REPs have been previously used to identify modifiers of toxicity [28, 136, 256]. For example, co-expression of the viral antiapoptotic caspase inhibitor p35 is capable of at least partially mitigating the rough eye phenotype of the SCA3tr-Q78 fly model, hinting to the presence of apoptotic processes in the course of polyQinduced cell degeneration [219]. This is supported by the discovery of cell death in polyQexpressing cells in the larval eye imaginal discs of SCA3tr-Q78 flies eventually manifesting in an impaired adult structure. However, the introduction of $p 35$ was not consistently as beneficial as previously described [219] and could not fully cope with the massive
neurotoxic effects of the SCA3tr-Q78 protein. Additionally, the protective p35 action obviously cannot be generalised for polyQ diseases since no mitigating effect has been observed for overexpression of toxic huntingtin transgenes [259].

The presence of SCA3tr-Q78 protein in flies can be verified on the one hand indirectly by the obvious pernicious effects triggered in the compound eye and on the other hand directly by immunostaining of eye imaginal discs as well as adult eye sections. Both show robust SCA3tr-Q78 expression coinciding with the onset of aggregation already in larval tissue. This explains why model flies already feature an REP at the time of hatching since the first morphological defects set in as early as in pupal stages. Despite the advantages of this robust degeneration phenotype and the mimicking of vertebrate disease processes, this early manifestation of polyQ toxicity consequences does not entirely reflect the late-onset situation of polyQ disease in humans and the overall pathogenesis is restricted to the photoreceptor subset of neurons. The high toxicity of SCA3tr-Q78 in this model may be explained with the truncation of the protein which is thought to be the process finally initiating aggregation. Eye-specific and pan-neural expression of the full length Ataxin-3 with an elongated stretch of 84 glutamines does not result in an overt eye phenotype at hatching yet exhibits late-stage and progressive neurodegeneration [209]. Possibly, these findings have their origin in the fact that the more toxic truncated variant of Ataxin-3 is produced in the first place whereas the full-length version initially has to be proteolytically processed before being rendered toxic. In addition, differences in the expression levels of the two transgenes might account for changes in toxicity. It is only when the cell's capacity to cope with the overload of toxic protein is exhausted that the toxic influences of the truncated proteins set in, which naturally happens faster with the originally shortened form. Expression of the admittedly truncated yet normal form of Ataxin-3 with 27 glutamine repeats does not exert any of the detrimental effects mentioned before, proving the crucial role of polyQ repeat elongation above a certain threshold.

Investigation of actual levels of truncated Ataxin-3 in Drosophila is rendered difficult by the highly aggregative nature of the protein, draining the pool of detectable monomeric protein and increasing the amount of aggregated protein not accessible to Western blot analysis. Given the large extend of photoreceptor loss in GMR_SCA3tr-Q78 flies, it is also reasonable to assume that substantial amounts of cells have already demised shortly after hatching. Thus the lack of protein production of these cells may account for low detection of the protein. Despite the lack of quantifiable biochemical detection of truncated Ataxin-3
in our disease model, there is sufficient evidence for SCA3tr-Q78 transgene expression combined with several options for analysis of genetic interactions. Despite frequent gene homologies between vertebrates and invertebrates, to date no ATXN3 orthologue has been described in Drosophila. Therefore no endogenous Ataxin-3 can interfere with transgene expression, protein levels or aggregation in a way that has been previously described [209], meaning there is full penetrance of toxic polyQ effects in SCA3tr-Q78-expressing flies.

A Drosophila model for frontotemporal dementia (FTD) and Parkinsonism linked to chromosome 17 comprising a mutant form of the mapt gene (coding for Tau[R406W]) [271] was used to determine the specificity of the obtained experimental results for polyQinduced toxicity. Targeting production of Tau[R406W] to postmitotic cells of the eye results in a severe REP presenting with disturbed external eye morphology and decreased eye size. Since function of normal Tau as well as mutant tau-induced pathogenesis are believed to be different from that of polyQ toxicity, modifiers exhibiting similar results in both disease models are unlikely to be specific for one of the proteins and are not analysed further as such.

### 6.2 Modifiers of Ataxin-3-induced REP in Drosophila

In the primary screen for modifiers of Ataxin-3-induced neurotoxicity, 529 RNAi lines were identified to change the polyQ-induced REP. Of this group, 21 RNAi strains also exhibited similar modulation of Tau[R406W]-induced degeneration and were therefore not considered specific for Ataxin-3, yet not excluded from further analysis. Finally, 508 RNAi lines representing 502 genes were identified as modulators of Ataxin-3-induced neurotoxicity in the Drosophila eye. Silencing of gene expression by 34 of these lines resulted in suppression of the REP, whereas 474 shRNAs rendered the REP more severe with the vast majority of lines being lethal in disease model progeny. These numbers are completed with 2 suppressing and 19 enhancing candidates featuring the same result in the Tau verification screen. By previous screening of the entire RNAi sublibrary devoted for the polyQ screen it was assured that the candidate genes are specific for the disease condition and are not of vital importance.

The high number of genes leading to lethal interactions appears surprising since expression of the toxic polyQ protein species during the screen is confined to differentiated
cells of the eye which should not interfere with the viability of the flies. This seeming contradiction may be explained by the fact that silencing of gene expression drains the cell of important regulatory mechanisms normally keeping the toxic effects of polyQ proteins at bay. Furthermore, the truncated Ataxin-3 used in the screen proved to be a highly toxic protein, not allowing for pan-neural expression and leading to a severe neurodegenerative phenotype. It stands to reason that massive cell demise in the course of the expression of elongated polyQ protein is even enhanced in combination with the lack of ameliorating gene action silenced by RNAi. Therefore, the extent of cell death might just overwhelm the capacity of the phagocytic clearance responsible for the uptake of apoptotic cell remainders [286, 287]. As a consequence, cellular debris and released polyQ aggregates would compromise the physiological functioning of adjacent tissues during development or even penetrate and infect other cells [275]. On top of that, GMR-positive cells and therefore polyQ protein expression have also been described to be present in non-retinal areas of the brain [252] whose demise may add to the detrimental effects of developing compound eye degeneration. Eventually, neighbouring neural tissue originally without polyQ protein expression is indirectly affected by polyQ toxicity and normal fly morphogenesis and hatching is prevented.

As a consequence of the large number of lethal candidates, only the RNAi lines producing vital offspring and having analyzable phenotypes were investigated further and grouped into categories reflecting the biological processes they are involved in. Nevertheless, the entire list of candidate lines was utilised for comparison of the results with the outcome of previously conducted modifier screens.

### 6.2.1 Comparison to related polyQ modifier screens

There are plenty screening approaches that have been implemented in order to discover and investigate modifiers of Ataxin-3 and other polyQ proteins in the physiological and disease state. The results obtained in the present work were compared to the outcome of three studies: an RNAi-based screen for modifiers of polyQ aggregation in C. elegans by Nollen et al. [267]; a genome-wide screen for modulators of Htt aggregation in Drosophila cells by Zhang et al. [268] and a genome-wide modifier screen for Ataxin-3induced neurotoxicity based on misexpression of endogenous Drosophila genes by Bilen and Bonini [256].

The categories of candidates obtained in the published screens resemble those specified in Table 9 representative for the complete modifier list. Zhang et al. identified for example chaperones, phosphatases/kinases, proteins involved in transcription and ubiquitin/proteasome pathways. Nollen et al. presented candidates grouping into the biological processes of protein synthesis, folding, transport, degradation and additionally RNA synthesis and processing. Bilen and Bonini for their part revealed genetic interactors acting as chaperones, in the ubiquitin pathway or having miscellaneous functions.

Comparing the three screens, the relatively high disparity in the number of obtained candidate genes is striking. Whereas the Bilen screen produced only 18 candidate genes, all other screens exhibit candidate numbers ten times and more as much, with the present work even yielding over five hundred modifiers of polyQ toxic action. This may speak in favour of a greater coverage of the genome by RNAi-based approaches compared to random misexpression of endogenous genes. Additionally, it is reasonable to assume that a loss-of-function of a gene product in a polyQ-burdened cell is more likely to occur and actually have an influence on toxicity than the artificial overexpression of a given gene. On the other hand, an unreasonable high number of candidate genes could also be an indicator for a high percentage of false-positive candidates produced by bystander effects not correlated to polyQ activity. Taking this into account, the chances of such false-positive modifier genes were minimised as far as possible by prior screening for RNAi effects in control flies and replication of crossbreeding for primary candidate hits. The at least in part comparable high numbers of the Nollen and Zhang screens and the present work should nevertheless not hide the fact that the first two were conducted in different models ( $C$. elegans and Drosophila cells, respectively) and were designed to investigate aggregation of polyQ proteins, not neurotoxicity as in the Bilen and the genome-wide RNAi screen. Consequently, results and possible similarities between all the screens should be taken cautiously. Although the Bilen and Bonini screen uses the same fly model and shares 22 \% of its candidate genes with the ones from this work, the higher number of overlapping modifiers in the other screens probably has its origin in the methodically similar RNAi approach (Figure 23).


Figure 23. Overlap between screens for genetic modifiers of polyQ-induced neurotoxicity or
aggregation.
Venn-like diagram showing genes mutually obtained as genetic modifiers in diverse polyQ protein disease models and screens. Depicted are only candidate genes shared by the different screens, not modifiers unique for one of the single screens. The present work is marked with red encircling. Modifier candidates genetically acting in the same direction (increasing or ameliorating toxicity/aggregation) are marked in green, candidates with opposing direction of action are grey.
Gene symbols are those for Drosophila as listed in Gene Database of NCBI [3].

### 6.2.2 Chaperones as polyQ misfolding and aggregation modifiers

What is evident yet not surprising in all the screens is the high portion of chaperone-related candidate genes. Chaperones and heat shock proteins have been implicated earlier in the amelioration of polyQ toxicity and aggregation [26, 30, 222, 288, 289]. Therefore, depletion of chaperones and their regulatory proteins mostly results in enhanced neurotoxicity and aggregation, whereas the opposite is the case upon increase of chaperone levels. Indeed, co-expression of human chaperone HSP70 substantially suppressed the REP of GMR_SCA3tr-Q78 flies (not shown). In whole, eight modifier genes of the chaperone class of proteins are shared in different combinations by the four previously described modifier screens. DnaJ-1 for example is a genetic suppressor of polyQ-induced neurotoxicity in the present work (enhancer if gene is silenced). An ameliorating influence on polyQ aggregation and neurotoxicity was shown by the Zhang and Bilen screens, also
confirmed by a fourth screen on polyQ by Kazemi-Esfarjani and Benzer [28]. A different chaperone-coding gene, $H s c 70-4$, has been published to mitigate SCA3tr-Q78 aggregation, and knockdown in the C. elegans screen facilitated aggregate formation. However, Zhang et al. reported the Hsc70-4 gene product to be an enhancer of Htt aggregation and also in the work at hand, silencing of Hsc70-4 produced an obvious suppression of the REP. The fact that in the present work two RNAi lines for Hsc70-4 produced comparable effects not only proves the principle of the screen, but additionally renders the obtained result credible. Modulating the activity of this gene might influence aggregation indirectly by interfering with protein functions apart from stress response, in the case of $H s c 70-4$ for example clathrin-dependent endocytosis [290, 291]. Despite that, overall chaperone functioning occupies a pivotal role in polyQ protein misfolding and aggregation by retaining or restoring native protein conformation. Thereby, chaperones interfere with the earliest steps of pathogenesis and putatively prevent accumulation and aggregation of toxic proteins in the first place.

### 6.2.3 Components of the UPS in polyQ pathogenesis

The next noteworthy functional group of modifiers is that of genes involved in ubiquitin- and proteasomal pathways. Ubiquitination of misfolded or dysfunctional proteins and their subsequent degradation by the proteasome is one of the key cellular processes to fight accumulation and aggregation of potentially toxic proteins. Naturally, genes involved in this pathway emerge as modifiers of neurotoxicity as well as aggregation. The RNAi-based screens on C. elegans and the present work have the UPS-related candidate $\operatorname{Pros} \beta 2$ in common, which genetically acts as a suppressor of aggregation/neurotoxicity in both screens. Another UPS example, l(2)05070, was identified as aggregation enhancer in the Zhang screen unlike being a suppressor in the Nollen work. To add to these findings, in the present screen four members of the UPS pathway have been identified as genetically enhancing candidates (Table 9) being responsible for either protein ubiquitination or deubiquitination. Additional to the UPS-related genes in Table 9, other UPS pathway genes exhibiting lethal effects when knocked down are also listed in Appendix Table 1, for example Uch-L3, encoding a deubiquitinating hydrolase described as a part of the regulatory complex of the 26 S proteasome [292]. One can conclude from these results that with regard to the UPS-related modifiers, a general statement about the impact of the
single components of the pathway on polyQ proteins is not possible. Instead it is necessary to take into account the specificity of the modifier protein (ubiquitinating or deubiquitinating) and its respective substrate protein and affected cellular process. Despite that, a lack of structural constituents of the proteasome, like Pros $\beta 2$, is in almost every case detrimental for the cell when facing an increased burden of misfolded protein or protein aggregation. An impact of ubiquitination on the physiological function of truncated Ataxin3 used in the present screen can be excluded since the protein lacks its enzymatically active domains.

In conclusion, by modulating the clearance of misfolded proteins, the members of the UPS pathway are of vital importance for cellular coping with elongated polyQ proteins and are potent modifiers of polyQ toxicity.

### 6.2.4 PolyQ-induced neurotoxicity modifiers involved in transcriptional regulation

As already mentioned in chapter 2.2.2, transcriptional dysregulation plays an important role in the course of polyQ pathogenesis either by loss-of-function of a mutated regulatory polyQ protein or interference of aggregates or the like with transcription itself. Like in the case of the UPS pathway, no generalised assumption can be made about an overall beneficial or harmful effect of transcriptional regulators on polyQ toxicity. Enhancement of the REP by silencing of chm (a histone acetyl transferase, HAT) is in line with the proposed beneficial effect of increasing HAT expression in polyQ disease [72]. Although also respresenting a HAT and being involved in cell cycle control via cdc2 [293], silencing of MRG15 led to suppression of the REP, demonstrating the possible opposing effects of genes with similar function. The same effect was observed for MRG15 knockdown in the Tau[R406W] model, hinting to a rather unspecific disadvantageous influence of MRG15 in cells affected by toxic proteins. One could speculate about a scenario in which acetylation of the $c d c 2$ promoter and thereby facilitation of transcription initiates a new mitotic cycle in $S$ phase. Activation of the cell cycle in neurons will drive the anyhow polyQstricken cells into apoptosis. This hypothesis demonstrates the detailed consideration of the specific processes the candidate genes are influencing with respect to polyQ toxicity.

### 6.2.5 Nuclear transport proteins are modifiers of polyQ toxicity

Export from the nuclear compartment via a nuclear export signal (NES) has been shown to be implicated in polyQ pathogenesis. The Drosophila orthologue for human exportin-1 (Xpo1), embargoed (emb), exhibited specificity for export of elongated polyQ proteins and disruption of this process increased polyQ toxicity by polyQ interference with transcription [294]. The deleterious effect of emb silencing was confirmed in the present work as well as in the screen of Bilen and Bonini [256]. Additionally, the genome-wide RNAi screen revealed another nuclear transporter, Exportin-6 (Exp6), as being involved in polyQ protein translocation since silencing of this gene resulted in lethality of polyQ flies. Studies have shown that the nuclear environment putatively fosters seeding of polyQ aggregates [295] and aggregation-prone polyQ fragments accumulate in the nucleus after escaping the cytoplasmic protein quality control [296]. Surprisingly, the RNAi screen in SCA3tr-Q78-expressing flies additionally revealed several importins, facilitating nuclear import, as being detrimental when knocked down. Among them are Trn, homologous to transportin-1 (TNPO1) and moleskin (msk), orthologue of importin-7 (IPO7), furthermore some import-related nuclear pores. It is not clear why a process opposed to nuclear export features the same findings after disruption. Computational analysis identified msk as a member of a gene cluster mainly involved in ribosomal and RNA biogenesis with the central gene Nop56 having recently been linked to SCA36 [297] (not shown). Possibly, the impact of nuclear import on polyQ toxicity is rather indirectly mediated by transcriptional and translational processes.

### 6.2.6 Further remarks on polyQ toxicity modifiers and the RNAi screen

Several other genes previously implicated in polyQ neurotoxicity and pathogenesis were identified in the present RNAi screen. Silencing of the Drosophila huntingtin orthologue had a lethal outcome in polyQ flies connecting SCA3 with the disease gene for Huntington's disease. In the case of another polyQ disorder, SCA2, and its disease gene ATXN2, no interaction could be proven despite findings reported previously [298]. Another noteworthy fact is the underrepresentation of autophagy-related genes in the screen results, a finding opposed to the described pivotal role of autophagy in mitigation of polyQrelated neurodegeneration (reviewed in [299]). Only one autophagy gene, Atg6, was shown
to potentially suppress SCA3tr-Q78 toxicity in its native state. A reason for this may be the prior screening for RNAi effects in GMR control flies. Members of the autophagy system and also several other genes (chaperones, structural cell constituents, transcription factors, proteasomal components etc.) are with high probability of vital importance for cellular survival themselves. In case that RNAi of these genes already exhibits changes in control flies, they were prevented from evolving as candidates for polyQ toxicity modulation due to the experimental design. However, this does not mean that they do not somehow interfere with elongated polyQ activity.

Representing a novel revelation, the biological process of lipid metabolism and, more precisely, of sphingolipid metabolism showed interesting influence on polyQ toxicity. Four members of this biological process are listed in Table 9 as obvious modifiers of polyQ-induced REP and several others exhibited lethal outcome following knockdown in SCA3tr-Q78 flies. Since sphingolipid pathways have been implicated in neurodegeneration [300] and impinge on diverse crucial cellular processes (apoptosis, differentiation, proliferation [301]), it would be worthwhile to further investigate the intertwining of these lipid-related mechanisms with respect to their impact on polyQ toxicity.

The relatively small overlap between the polyQ RNAi screen and the findings made in Tau[R406W]-expressing flies underlines the specificity of the discovered modifiers for SCA3-linked pathogenesis. Nevertheless, the majority of genes being modifiers in both screens exhibit the same mode of change in their respective REP. For some candidates, this might be explained by the general importance of these genes in cellular coping strategies against proteotoxic stress. Pros $\beta 2$ and Rpn9, both structural constituents of the proteasome, are examples for fundamental genes in order to fight misfolded/aggregated proteins. Due to the fact that silencing $\operatorname{Pros} \beta 2$ and Rpn9 had no effect in control flies, a general necessity for survival cannot be deduced from the lethal outcome of silencing of these two genes in the disease models.

### 6.3 Aggregation in SCA3tr-Q78-shRNA-coexpressing flies

It has been shown previously biochemically [276] and immunohistochemically [256] that several genes, especially chaperones, feature aggregate-mitigating properties on polyQ proteins. Despite these findings, the conduction of a large-scale analysis of aggregation with filter retardation approaches yielded contradictory results. Whereas
some, yet by far not the majority of REP suppressor candidates decreased the load of SDSinsoluble polyQ aggregates, this was also the case for a number of REP enhancers. Given the proposed connection between aggregate formation and polyQ neurotoxicity, these results are at least surprising and cannot be explained by slight changes in deployed protein amount during the experiment. It might well be that the mode of sample preparation depletes a certain amount of higher molecular aggregates; however, these are not considered to be the actual toxic species. One reasonable explanation for the differential outcome of REP suppressors may be the high number of investigated genes compared to previous studies, producing a more comprehensive image of aggregation modulation. Furthermore, the influences of the various modifiers on polyQ aggregation might be more complex and cannot be reduced to simple molecular mechanisms somehow being related to aggregation. The reason for the missing correlation between SDS-insoluble aggregates, microscopically visible inclusions and polyQ neurotoxicity lies presumably in the differential nature of the aggregate species. Inclusions are considered to be rather beneficial for the cell, protecting it from the detrimental effects of the unbound, yet SDSinsoluble polyQ oligomers. Here again, further investigations are necessary prior to giving a reliable statement about advantageous or disadvantageous effects of modifiers on the various aggregation states.

### 6.4 The role of TRMT2A in polyQ pathogenesis

Knockdown of the Drosophila orthologue of tRNA methyltransferase 2 homologue $A$ (TRMT2A), CG3808, in the SCA3tr-Q78 modifier screen exhibited some of the most potent effects with regard to the suppression of the REP and amelioration of polyQ aggregate load. Furthermore, CG3808 silencing in Drosophila rescued lethality of flies with pan-neural expression of SCA3tr-Q78, restored photoreceptor neuron loss and prolonged polyQcompromised longevity in an adult-onset disease model. Moreover, select findings could also be extended to fly models for other polyQ disorders like HD and SCA1. Given the fact that these are complete novel observations and tRNA methyltransferases have never before been implicated in polyQ pathogenesis, nor is there an obvious mechanistic connection, these results are rather surprising. Eventually, effects like mitigation of aggregate load could be even recapitulated in mammalian HEK cells stably transfected with shRNA against TRMT2A.

Studies on patient tissue presented an association between the expression of TRMT2A in humans and the recurrence in breast cancer in a subset of patients [302]. Apart from that, the only link to neurological disorders is a putative connection of a single nucleotide polymorphisms in the TRMT2A gene with schizophrenia [303]

The process of tRNA methylation is a crucial event in the high-fidelity translation of mRNA into polypeptides. tRNA methyltransferase enzymes are according to their specificity capable of linking a methyl group to a tRNA base, hereby changing the chemical properties of this residue. Dependent on the location of the mentioned nucleotide, tRNA methylation interferes with different mechanisms prior to and during translation of mRNA into proteins and exhibits a pivotal influence upon tRNA stability and maturation [304306]. It is reasonable to assume that silencing of methyltransferase expression and thus lacking tRNA modification might hamper one of these processes. As a consequence, aminoacylation of tRNA may decline in fidelity, leaving the tRNA associated with an incorrect amino acid. Upon introduction of this false component into the polypeptide chain, the chemical properties of the resulting protein are likely to be altered in conformation due to changed molecular interactions and also with respect to enzymatic activity caused by modified active sites.


Figure 24. Putative mechanistic explanation of polyQ toxicity amelioration by TRMT2A knockdown.
(A) Under normal conditions, the Drosophila TRMT2A orthologue, CG3808, transfers a methyl group onto uridine 54 of the tRNA. Thereby, high-fidelity amino acid loading is ensured. In the case of mutated polyQ disease genes, this results in an elongated glutamine repeat domain which is detrimental for the cell. (B) Silencing of CG3808 and consecutive lack of tRNA methylation may render tRNA-amino acid linkage inaccurate, resulting in introduction of a different amino acid into the polyQ stretch (here E, glutamate). This would decrease toxicity of the polyQ domain to a great extend.

Taking into account the generalised effect of TRMT2A knockdown in diverse polyQ models, the variances are more probable to be confined to the polyQ tract, the only common feature of the disease proteins. Unfortunately, until now we were not able to prove this hypothesis due to experimental restrictions, like impossible analysis of the polyQ stretch by mass spectrometry, or show experimental evidence for specificity of the putative mechanism for polyQ proteins. However, CG3808 silencing failed to show an effect on the REP of other neurodegenerative disease models (TDP-43, not shown) or exhibited a different REP modification in the Tau[R406W] flies (lethal, not shown). Therefore, a certain specificity for polyQ proteins was substantiated. Hopefully, future experiments profiting from the putative erratic introduction of amino acids in the polyQ domain following TRMT2A silencing will shed more light on the mechanistic basis of methyltransferase silencing and polyQ pathogenesis. The significant increase of polyQ inclusion bodies in TRMT2A knockdown cells leaves additional room for speculations as to how this presumably beneficial detoxification process is brought about following TRMT2A silencing. A slightly different explanatory approach is conceivable with respect to target specificity of TRMT2A. It transfers a methyl group onto uridine at position 54 of the tRNA, producing 5methyluridine (ribothymidine) in the T-loop structure (Figure 25A) [307]. Disruption of this process may lead to subsequent hampering of T-loop-related processes in the course of translation, for example binding to the ribosome or distinct aminoacylation. Since no general disturbance of protein production was observed, interference with translational processes according to this theory would be confined to specific amino acids and proteins like polyQ by unknown mechanisms.

Although investigations on the molecular basis of the TRMT2A influence on polyQ toxicity have not yielded enlightening results so far, the remarkable findings described in this work make this protein and its related processes a worthwhile target of further experimentation.

## 7 Summary and Concluding Remarks

Almost two decades have passed since the discovery of the elongated polyQ stretch as the molecularpathological basis of polyQ diseases. Despite this time of extensive scientific effort, it is still not clear which cellular pathways and mechanisms ultimately lead to polyQ toxicity or what the definite toxic species during pathogenesis is. Numerous approaches have focused on single molecular processes and yielded insight into their impact on polyQ proteins, nevertheless we are still lacking a comprehensive overview or network of polyQ pathogenesis and the cellular proteins involved therein. The respective knowledge is an imperative prerequisite in order to develop feasible treatment approaches to eventually cure these devastating disorders.

In an attempt to contribute to revelation of disease mechanisms and modulations thereof, we conducted a large-scale RNAi screen for modifiers of truncated SCA3 proteininduced neurotoxicity in Drosophila. As a result we were able to obtain a set of potential genetic interactors of polyQ proteins in the course of the disease. Together with already known modifier genes like chaperones, UPS pathway members and nuclear exportins, the experiments yielded novel genes involved in processes previously not described for polyQ disease. Genes responsible for sphingolipid metabolism seem to play a role in the emergence of neurodegeneration as well as nuclear importins and genes responsible for ribosome biogenesis. The results furthermore revealed the strong potency of the tRNA methyltransferase TRMT2A and its Drosophila equivalent CG3808 to modify polyQ toxicity by so far unknown mechanisms.

Concluding, the genes obtained as polyQ neurotoxicity modulators during the RNAi screen and in subsequent experiments provide a promising entity of genetic interactors of polyQ protein and a valuable pool for future research in order to shed light on polyQ pathogenesis.

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## Appendix

## I Additonal eye phenotypes



Appendix Figure 1. Additional eye phenotypes referred to in the thesis.
(A) REP induced by GMR-mediated Tau[R406W] expression and utilised in rescreening for polyQ specificity of the RNAi effects. (B) GMR-mediated overexpression of UAS-p35 in SCA3tr-Q78 flies results in amelioration of the polyQinduced REP. (C) A different SCA3tr-Q78 individual exhibits a worsened REP with UAS-p35. (D) Co-expression of SCA3tr-Q78 and shRNA against the white gene shows loss of white-mediated eye coloration, but no change in degenerative appearance of the compound eye.
All scale bars apply to $200 \mu \mathrm{~m}$ respectively.

## II RNAi lines modifying SCA3tr-Q78-induced REP

Appendix Table 1. RNAi lines modifying SCA3tr-Q78-induced REP.
Lines marked with (Tau) in REP modification column exhibited similar result in rescreening with Tau[R406W]induced REP. Lines listed in red colour show reduced vitality or lethality of progeny following ubiquitous shRNA expression with actin5C-GAL4.
S, suppression of REP; E, enhancement of REP; n.a., not available.

| Transformant ID | CG number | Gene name | REP <br> modification | Phenotype |
| :--- | :--- | :--- | :--- | :--- |


| 8780 | CG17048 | CG17048 | S |  |
| :---: | :---: | :---: | :---: | :---: |
| 7903 | CG9501 | ppk14 | S |  |
| 23121 | CG7123 | LanB1 | S | n.a. |
| 44362 | CG9131 | slmo | S | n.a. |
| 37221 | CG9153 | CG9153 | S |  |
| 15789 | CG8696 | LvpH | S |  |
| 26465 | CG4264 | Hsc70-4 | S |  |
| 50222 | CG4264 | Hsc70-4 | S |  |


| 11219 | CG3284 | Rpll15 | S |
| :---: | :---: | :---: | :---: |
| 3780 | CG5799 | dve-s | S |
| 24030 | CG9448 | trbd | S |
| 40006 | CG15618 | CG15618 | S |
| 41530 | CG14514 | Brd8 | S |
| 23637 | CG8863 | Droj2 | S |
| 43870 | CG7108 | DNApol- <br> alpha50 | S |
| 45596 | CG8937 | Hsc70-1 | S |
| 41696 | CG2720 | Hop | S (Tau) |


| 40044 | CG16890 | CG16890 | S |
| :---: | :---: | :---: | :---: |
| 8070 | CG13969 | bwa | S |
| 6282 | CG6755 | EloA | S |
| 31257 | CG10545 | Gbeta13F | S |
| 19066 | CG1658 | Doa | S |
| 33262 | CG5687 | CG5687 | S |
| 13005 | CG9695 | Dab | S |
| 16182 | CG1107 | $a u x$ | S |
| 37930 | CG14619 | CG14619 | S |
| 46473 | CG17919 | CG17919 | S |






| 8620 | CG4288 | CG4288 | E |
| :---: | :---: | :---: | :---: |
| 28019 | CG7436 | Nmt | lethal |
| 22574 | CG7843 | Ars2 | lethal |
| 51209 | CG10281 | TfIIFalpha | lethal |
| 14874 | CG2145 | CG2145 | lethal |
| 14890 | CG15739 | CG15739 | lethal |
| 22561 | CG7275 | CG7275 | lethal |
| 30179 | CG6921 | bond | lethal |
| 30214 | CG16785 | fz3 | lethal |
| 21393 | CG31687 | CG31687 | lethal |
| 1385 | CG9753 | AdoR | lethal |
| 9865 | CG7709 | Muc91C | lethal |
| 41130 | CG7807 | AP-2 | lethal |
| 46150 | CG7085 | 1(2)s5379 | lethal |
| 21985 | CG31318 | Rpb4 | lethal |
| 38471 | CG1129 | CG1129 | lethal |
| 45635 | CG6944 | Lam | lethal |
| 46072 | CG6589 | spag4 | lethal |
| 26959 | CG8431 | Aats-cys | lethal |
| 44942 | CG31321 | CG31321 | lethal |
| 40907 | CG5404 | CG5404 | lethal |
| 39402 | CG5310 | $n m d y n-D 6$ | lethal |
| 14210 | CG8189 | ATPsyn-b | lethal |
| 14194 | CG17081 | Cep135 | lethal |


| 39224 | CG18812 | CG18812 | lethal |
| :---: | :---: | :---: | :---: |
| 39256 | CG9742 | SmG | lethal |
| 20334 | CG16938 | Tif-IA | lethal |
| 18107 | CG7279 | Lip1 | lethal |
| 32680 | CG1640 | CG1640 | lethal |
| 44991 | CG11136 | Lrt | lethal |
| 16506 | CG5599 | CG5599 | lethal |
| 28745 | CG12524 | CG34356 | lethal |
| 34160 | CG6340 | CG6340 | lethal |
| 50706 | CG4241 | att-ORFA | lethal |
| 13613 | CG2917 | Orc4 | lethal |
| 35065 | CG6066 | CG6066 | lethal |
| 36050 | CG8849 | $m R p L 24$ | lethal |
| 21792 | CG4132 | pkaap | lethal |
| 40076 | CG17419 | CG41099 | lethal |
| 21793 | CG4152 | $1(2) 35 D f$ | lethal |
| 11852 | CG9245 | Pis | lethal |
| 15602 | CG3759 | CG3759 | lethal |
| 47537 | CG11010 | Ent3 | lethal |
| 49822 | CG9958 | snapin | lethal |


| 6143 | CG4928 | CG4928 | lethal |
| :---: | :---: | :---: | :---: |
| 40972 | CG17681 | CG17681 | lethal |
| 43998 | CG6335 | Aats-his | lethal (Tau) |
| 44104 | CG10126 | CG10126 | lethal |
| 39848 | CG14905 | CG14905 | lethal |
| 21308 | CG31291 | CG31291 | lethal |
| 33735 | CG31000 | heph | lethal |
| 33787 | CG31211 | CG31211 | lethal |
| 25195 | CG15143 | CG15143 | lethal |
| 14869 | CG2124 | CG2124 | lethal |
| 29332 | CG9927 | Art6 | lethal |
| 14861 | CG7598 | CG7598 | lethal |
| 34316 | CG5121 | MED28 | lethal |
| 38269 | CG3776 | CG3776 | lethal |
| 33186 | CG4482 | mol | lethal |
| 21139 | CG3035 | cm | lethal |
| 38963 | CG7935 | msk | lethal |
| 49153 | CG13779 | CG13779 | lethal |
| 24177 | CG9998 | U2af50 | lethal |
| 38491 | CG11360 | CG11360 | lethal |
| 39450 | CG31704 | CG31704 | lethal |
| 34479 | CG32253 | CG11583 | lethal |
| 21999 | CG5085 | Sirt2 | lethal |
| 22068 | CG5335 | CG5335 | lethal |
| 31789 | CG13779 | CG13779 | lethal |


| 33561 | CG2708 | Tom34 | lethal |
| :---: | :---: | :---: | :---: |
| 49879 | CG7014 | RpS5b | lethal |
| 3245 | CG10913 | Spn6 | lethal |
| 26432 | CG4202 | Sas10 | lethal |
| 6723 | CG10693 | slo | lethal |
| 30623 | CG5553 | DNApol- <br> alpha60 | lethal |
| 49525 | CG33505 | U3-55K | lethal |
| 51979 | CG10564 | Ac78C | lethal |
| 7802 | CG8933 | exd | lethal |
| 52165 | CG13849 | Nop56 | lethal |
| 7800 | CG4035 | elF-4E | lethal |
| 51363 | CG16884 | CG16884 | lethal |
| 4634 | CG30048 | CG30048 | lethal |
| 52094 | CG10582 | Sin | lethal |
| 46977 | CG1989 | Yippee | lethal |
| 44976 | CG7769 | pic | lethal |
| 27610 | CG4969 | Wnt6 | lethal |
| 42915 | CG7176 | Idh | lethal |
| 21374 | CG3158 | spn-E | lethal |
| 42716 | CG5911 | ETHR | lethal |
| 2487 | CG17075 | CG17075 | lethal |
| 8361 | CG11278 | Syx13 | lethal |
| 7787 | CG9696 | dom | lethal |


| 34145 | CG5869 | CG5869 | lethal |
| :---: | :---: | :---: | :---: |
| 38319 | CG10033 | for | lethal |
| 41406 | CG1882 | CG1882 | lethal |
| 44562 | CG11739 | CG11739 | lethal |
| 45402 | CG12298 | sub | lethal |
| 1335 | CG11958 | Cnx99A | lethal |
| 8907 | CG1139 | CG1139 | lethal |
| 24749 | CG3329 | Prosbeta2 | lethal (Tau) |
| 49245 | CG2241 | Rpt6R | lethal |
| 7308 | CG3305 | CG3305 | lethal |
| 52486 | CG2905 | Nipped-A | lethal |
| 9039 | CG6827 | Nrx-IV | lethal (Tau) |
| 2857 | CG7431 | CG7431 | lethal |
| 18440 | CG2918 | CG2918 | lethal |
| 31522 | CG11546 | kermit | lethal |
| 44325 | CG5651 | pix | lethal |
| 44589 | CG12275 | RpS10a | lethal (Tau) |
| 47116 | CG5969 | CG5969 | lethal |
| 17171 | CG13391 | Aats-ala | lethal |
| 12482 | CG2478 | bru | lethal (Tau) |
| 46499 | CG1030 | Scr | lethal |
| 38154 | CG3589 | CG3589 | lethal |
| 39091 | CG31289 | Dph5 | lethal |
| 22496 | CG6509 | CG6509 | lethal |
| 37250 | CG5751 | TrpA1 | lethal |
| 28341 | CG1903 | sno | lethal |


|  |  | APPENDIX |  | 120 |
| :---: | :---: | :---: | :---: | :---: |
| 27152 | CG10315 | elF2B-delta | lethal |  |
| 40477 | CG3843 | RpL10Aa | lethal |  |
| 13054 | CG7162 | MED1 | lethal |  |
| 52392 | CG4960 | CG4960 | lethal |  |
| 47126 | CG3849 | Lasp | lethal |  |
| 33135 | CG4521 | mthl1 | lethal |  |
| 33256 | CG3499 | CG3499 | lethal |  |
| 25246 | CG17293 | CG17293 | lethal |  |
| 15877 | CG12031 | MED14 | lethal (Tau) |  |
| 10639 | CG6146 | Top1 | lethal |  |
| 25547 | CG7757 | CG7757 | lethal |  |
| 17302 | CG12727 | CG32635 | lethal |  |
| 25535 | CG7742 | CG7742 | lethal |  |
| 42010 | CG4843 | Tm2 | lethal |  |
| 49800 | CG6835 | GS | lethal |  |
| 23689 | CG9344 | CG9344 | lethal |  |
| 36252 | CG12283 | kek1 | lethal |  |
| 31726 | CG12325 | CG12325 | lethal |  |
| 13503 | CG8222 | Pur | lethal |  |
| 30000 | CG4357 | Ncc69 | lethal |  |
| 41980 | CG4180 | $1(2) 35 B g$ | lethal |  |
| 36175 | CG9961 | CG9961 | lethal |  |
| 30462 | CG6534 | slou | lethal |  |
| 36121 | CG9619 | CG9619 | lethal |  |


| 30448 | CG5179 | Cdk9 | lethal |
| :--- | :---: | :---: | :---: |
| 30431 | CG7772 | CG7772 | lethal |
| 34618 | CG3431 | Uch-L3 | lethal |
| 41977 | CG4165 | CG4165 | lethal |
| 41965 | CG30000 | CG30000 | lethal |
| 35200 | CG7292 | RGrp6 | lethal |
| 34845 | CG12318 | CG1745 | CG4438 |


| 27498 | CG5735 | orb2 | lethal |
| :---: | :---: | :---: | :---: |
| 15261 | CG13926 | CG13926 | lethal |
| 18567 | CG8877 | Prp8 | lethal (Tau) |
| 25787 | CG31657 | PNUTS | lethal |
| 31456 | CG11201 | TTLL3B | lethal |
| 49547 | CG31201 | GluRIIE | lethal |
| 50510 | CG33931 | Rpp20 | lethal |
| 32443 | CG31639 | Uch-L3 | lethal |
| 34995 | CG5394 | Aats-glupro | lethal |
| 7752 | CG5353 | Aats-thr | lethal |
| 29589 | CG3071 | CG3071 | lethal |
| 23033 | CG8091 | Nc | lethal |
| 13044 | CG1271 | CG1271 | lethal |
| 41714 | CG7650 | CG7650 | lethal |
| 26001 | CG6852 | CG6852 | lethal |
| 11693 | CG6364 | CG6364 | lethal (Tau) |
| 26759 | CG10726 | barr | lethal |
| 5322 | CG5582 | $c \ln 3$ | lethal |
| 26007 | CG7039 | CG7039 | lethal |
| 7878 | CG7234 | GluRIIB | lethal |
| 10756 | CG10037 | vvl | lethal |
| 49844 | CG14206 | RpS10b | lethal |
| 46284 | CG15772 | CG15772 | lethal |


| 17002 | CG10811 | eIF4G | lethal |
| :---: | :---: | :---: | :---: |
| 43955 | CG1412 | RhoGAP19D | lethal |
| 3347 | CG13387 | emb | lethal |
| 44557 | CG6707 | CG6707 | lethal |
| 28065 | CG7788 | Ice | lethal |
| 28798 | CG9924 | $r d x$ | lethal |
| 45530 | CG4751 | CG4751 | lethal |
| 4047 | CG12891 | CPTI | lethal |
| 44263 | CG7480 | Pgant35A | lethal |
| 44535 | CG4780 | membrin | lethal |
| 46445 | CG33193 | sav | lethal |
| 44570 | CG9867 | CG9867 | lethal |
| 49372 | CG11877 | CG11877 | lethal |
| 12209 | CG2901 | CG2901 | lethal |
| 38481 | CG11299 | CG11299 | lethal |
| 45789 | CG8258 | CG8258 | lethal |
| 12746 | CG5163 | TfIIA-S | lethal |
| 12645 | CG1064 | Snr1 | lethal |
| 39207 | CG9802 | Cap | lethal |
| 22480 | CG6226 | FK506-bp1 | lethal |
| 18031 | CG10192 | elF4G2 | lethal |


| 49655 | CG10546 | Cralbp | lethal |
| :---: | :---: | :---: | :---: |
| 38399 | CG10716 | 4EHP | lethal |
| 23873 | CG13349 | CG13349 | lethal |
| 44484 | CG1616 | $d p a$ | lethal |
| 23851 | CG1316 | CG1316 | lethal |
| 44449 | CG1718 | CG1718 | lethal |
| 40336 | CG5913 | CG5913 | lethal |
| 4789 | CG10975 | Ptp69D | lethal |
| 10843 | CG9426 | CG9426 | lethal |
| 31619 | CG11989 | Ard1 | lethal (Tau) |
| 21010 | CG5994 | Nelf-E | lethal (Tau) |
| 842 | CG14396 | Ret | lethal |
| 31444 | CG11184 | Upf3 | lethal |
| 12768 | CG5499 | His2Av | lethal |
| 21258 | CG3058 | Dim1 | lethal |
| 23625 | CG8841 | CG8841 | lethal |
| 19208 | CG3011 | CG3011 | lethal |
| 32719 | CG1676 | cactin | lethal |
| 15627 | CG10984 | CG10984 | lethal |
| 43790 | CG5405 | KrT95D | lethal |
| 43549 | CG11899 | CG11899 | lethal |
| 50643 | CG31809 | CG31809 | lethal |
| 3326 | CG10657 | CG10657 | lethal |
| 34792 | CG4090 | Mur89F | lethal |


| 6315 | CG8384 | gro | lethal |
| :---: | :---: | :---: | :---: |
| 20876 | CG2503 | atms | lethal |
| 865 | CG10776 | wit | lethal |
| 35272 | CG7791 | CG7791 | lethal |
| 36028 | CG8786 | CG8786 | lethal |
| 26615 | CG4735 | shu | lethal |
| 12581 | CG8151 | Tfb1 | lethal |
| 12149 | CG7623 | sll | lethal |
| 49328 | CG11907 | Ent1 | lethal |
| 15453 | CG3644 | bic | lethal (Tau) |
| 50435 | CG32602 | Muc12Ea | lethal |
| 30884 | CG10374 | Lsd-1 | lethal |
| 32395 | CG16901 | sqd | lethal |
| 4180 | CG12929 | CG12929 | lethal |
| 16125 | CG10961 | Traf6 | lethal |
| 49848 | CG1740 | $N t f-2$ | lethal |
| 37663 | CG5640 | Utx | lethal |
| 7748 | CG7926 | Axn | lethal |
| 15185 | CG5186 | slim | lethal |
| 22548 | CG7257 | Rpt4R | lethal |
| 29253 | CG13628 | Rpb10 | lethal |
| 42779 | CG3881 | GlcAT-S | lethal |
| 16569 | CG12951 | CG12951 | lethal |
| 42776 | CG18419 | CG33298 | lethal |
| 27528 | CG5844 | CG5844 | lethal |


| 26075 | CG4599 | Tpr2 | lethal |
| :---: | :---: | :---: | :---: |
| 14268 | CG4086 | $\mathrm{Su}(\mathrm{P})$ | lethal |
| 1414 | CG5677 | Spase22-23 | lethal |
| 50221 | CG2076 | CG2076 | lethal |
| 23556 | CG17935 | Mst84Dd | lethal |
| 33516 | CG2272 | $s / p r$ | lethal |
| 27110 | CG32179 | Krn | lethal |
| 27002 | CG9100 | Rab30 | lethal (Tau) |
| 33423 | CG1911 | CAP-D2 | lethal |
| 30066 | CG7398 | CG8219 | lethal |
| 33507 | CG2253 | Upf2 | lethal |
| 8254 | CG7026 | CG7026 | lethal |
| 3016 | CG4001 | Pfk | lethal |
| 3166 | CG10778 | CG10778 | lethal |
| 3046 | CG11282 | caps | lethal |
| 33523 | CG2321 | CG2321 | lethal |
| 39937 | CG17083 | CG17083 | lethal |
| 39976 | CG1542 | CG1542 | lethal |
| 29295 | CG9836 | CG9836 | lethal |
| 48153 | CG15804 | Dhc62B | lethal |
| 6098 | CG18549 | CG18549 | lethal |
| 4801 | CG15744 | CG15744 | lethal |
| 43944 | CG1965 | CG1965 | lethal |
| 26277 | CG3733 | Chd1 | lethal |


| 42485 | CG3297 | mnd | lethal |
| :--- | :---: | :---: | :--- |
| 19616 | CG15816 | NA | lethal |
| 27607 | CG4364 | CG4364 | lethal |
| 35162 | CG7034 | sec15 | lethal |
| 15736 | CG6443 | CG6443 | lethal |
| 32085 | CG14034 | NA | lethal |
| 28982 | CG8887 | CG15666 | CG15666 |


| 34737 | CG3923 | Exp6 | lethal |
| :---: | :---: | :---: | :---: |
| 27486 | CG5692 | raps | lethal |
| 34331 | CG5596 | Mlc1 | lethal |
| 17463 | CG14286 | CG14286 | lethal |
| 21563 | CG5323 | CG5323 | lethal |
| 35343 | CG8108 | CG8108 | lethal (Tau) |
| 29070 | CG9177 | elF5 | lethal |
| 39529 | CG17743 | pho | lethal |
| 36584 | CG9973 | CG9973 | lethal |
| 11210 | CG9633 | RpA-70 | lethal |
| 28895 | CG8351 | Tcp-1eta | lethal |
| 26275 | CG3714 | CG3714 | lethal |
| 26227 | CG3542 | CG3542 | lethal |
| 49168 | CG14210 | CG14210 | lethal |
| 11205 | CG12005 | Mms19 | lethal |
| 21845 | CG4389 | CG4389 | lethal |
| 27943 | CG7293 | KIp68D | lethal |
| 16091 | CG10920 | CG10920 | lethal |
| 3909 | CG10165 | CG10165 | lethal |
| 21782 | CG4062 | Aats-val | lethal |
| 20144 | CG12085 | $p \cup f 68$ | lethal |
| 11227 | CG6349 | DNApol- <br> alpha180 | lethal |
| 12920 | CG7929 | ocn | lethal |
| 13566 | CG7665 | Fsh | lethal |
| 12662 | CG6545 | Ibe | lethal |


| 36308 | CG5528 | Toll-9 | lethal |
| :---: | :---: | :---: | :---: |
| 33650 | CG7686 | CG7686 | lethal |
| 28396 | CG5684 | Pop2 | lethal |
| 41740 | CG30390 | SGf29 | lethal |
| 41964 | CG3820 | Nup214 | lethal |
| 32025 | CG12812 | Fancl | lethal |
| 6236 | CG1378 | tll | lethal |
| 7563 | CG14077 | CG14077 | lethal |
| 51496 | CG13077 | CG13077 | lethal |
| 26309 | CG3931 | Rrp4 | lethal |
| 5150 | CG5950 | SrpRbeta | lethal |
|  |  |  |  |
| 18762 | CG12630 | tio | lethal |
| 51846 | CG1571 | CG1571 | lethal |
| 29072 | CG9198 | shtd | lethal |
| 40834 | CG13431 | Mgat1 | lethal |
| 27600 |  |  | lety)3 |


| 16744 | CG15749 | dmrt11E | lethal |
| :---: | :---: | :---: | :---: |
| 27515 | CG5788 | UbcD10 | lethal |
| 41819 | CG3358 | CG3358 | lethal |
| 28058 | CG7516 | $1(2) 34 \mathrm{Fd}$ | lethal |
| 35611 | CG7052 | TeplI | lethal |
| 41885 | CG32376 | CG32376 | lethal |
| 34070 | CG5374 | T-cp1 | lethal |
| 27457 | CG5546 | MED19 | lethal |
| 49345 | CG9155 | Myo61F | lethal |
| 48835 | CG7281 | CycC | lethal |
| 48793 | CG33051 | CG33051 | lethal |
| 48708 | CG10315 | elF2B-delta | lethal |
| 35222 | CG7376 | CG7376 | lethal |
| 27870 | CG7128 | Taf8 | lethal |
| 31320 | CG10687 | Aats-asn | lethal |
| 31311 | CG10645 | Iama | lethal |
| 31333 | CG10719 | brat | lethal |
| 35107 | CG6620 | ial | lethal |
| 6543 | CG7398 | Trn | lethal |
| 14444 | CG6343 | ND42 | lethal |
| 26585 | CG4649 | Sodh-2 | lethal |


| 41351 | CG31015 | PH4alphaPV | lethal |
| :---: | :---: | :---: | :---: |
| 40727 | CG9004 | CG9004 | lethal |
| 30673 | CG6772 | Slob | lethal |
| 44146 | CG12752 | Nxt1 | lethal |
| 8784 | CG10808 | synaptogyrin | lethal |
| 17701 | CG13778 | Mnn1 | lethal |
| 29462 | CG7564 | CG7564 | lethal |
| 40665 | CG9705 | CG9705 | lethal |
| 14107 | CG32374 | CG32374 | lethal |
| 30140 | CG6249 | CsI4 | lethal |
| 36085 | CG9049 | hiw | lethal |
| 36086 | CG9124 | elF-3p40 | lethal |
| 32749 | CG16837 | CG16837 | lethal |
| 36092 | CG9200 | Atac1 | lethal |
| 35165 | CG7070 | PуK | lethal |
| 46903 | CG14230 | CG14230 | lethal |
| 49030 | CG5440 | CG5440 | lethal |
| 32612 | CG15481 | Ski6 | lethal |
| 51088 | CG4602 | Srp54 | lethal |
| 34388 | CG7861 | tbce | lethal |
| 22108 | CG5383 | PSR | lethal |
| 51202 | CG9452 | CG9452 | lethal |
| 31206 | CG10230 | Rpn9 | lethal (Tau) |
| 15872 | CG3069 | Taf10b | lethal |


| 8573 | CG18578 | Ugt86Da | lethal |
| :---: | :---: | :---: | :---: |
| 22773 | CG11985 | CG11985 | lethal (Tau) |
| 31216 | CG10308 | CycJ | lethal (Tau) |
| 20567 | CG1789 | CG1789 | lethal |
| 15791 | CG8695 | LvpL | lethal |
| 41703 | CG2854 | CG2854 | lethal |
| 50126 | CG10098 | CG10098 | lethal |
| 36205 | CG9995 | $h t t$ | lethal |
| 39920 | CG30144 | CG33786 | lethal |
| 34847 | CG4448 | $w d a$ | lethal |
| 24054 | CG9527 | CG9527 | lethal |
| 50176 | CG3022 | $G A B A-B-R 3$ | lethal |
| 32521 | CG31809 | CG31809 | lethal |
| 52549 | CG9191 | Klp61F | lethal |
| 4659 | CG8657 | Dgkepsilon | lethal |
| 23037 | CG8107 | CalpB | lethal |
| 12618 | CG6258 | RfC38 | lethal |
| 6053 | CG14709 | CG14709 | lethal |
| 50021 | CG11276 | RpS4 | lethal |
| 40497 | CG4005 | yki | lethal |
| 28172 | CG7989 | wcd | lethal |
| 6832 | CG4907 | CG4907 | lethal |
| 25967 | CG4247 | mRpS10 | lethal |


| 24835 | CG1404 | ran | lethal |
| :--- | :---: | :---: | :---: |
| 36428 | CG5742 | CG5742 | lethal |
| 5985 | CG14690 | tomboy20 | lethal |
| 49325 | CG32708 | CG32708 | lethal |
| 31271 | CG10578 | DnaJ-1 | lethal |
| 46584 | CG9899 | CG9899 | lethal |
| 16313 | CG11804 | CGed-6 | lethal |
| 16806 | CG16804 | CGpt4 | lethal |
| 34498 | CG3229 | CG5367 | CG33123 |


| 31318 | CG10662 | sick | lethal |
| :---: | :---: | :---: | :---: |
| 51705 | CG11105 | CG42683 | lethal |
| 16331 | CG11861 | Cul-3 | lethal |
| 32482 | CG2578 | Ten-a | lethal |
| 28628 | CG7826 | mnb | lethal |
| 12616 | CG9207 | Gas41 | lethal |

## III Fly lines used for verification of RNAi

Appendix Table 2. Essential Drosophila genes exhibiting lethality/semilethality upon Actin5C-induced ubiquitous RNAi.

| Gene | Gene Symbol | Transformant ID |  |
| :---: | :---: | :---: | :---: |
| CG1030 | Scr | 46499 (enhancer) <br> 3033 (enhancer) | $\begin{aligned} & \text { semilethal }(2 / 15,13 \%) \\ & \text { lethal } \end{aligned}$ |
| CG1064 | Snr1 | 12645 (enhancer) | lethal |
| CG1378 | tll | 6236 (enhancer) | semilethal (13/82, 16 \%) |
| CG1433 | Atu | 17490 (enhancer) | lethal |
| CG1616 | dpa | 44484 (enhancer) | lethal |
| CG1903 | sno | 28341 (enhancer) | n.a. |
| CG2503 | atms | 20876 (enhancer) | lethal |
| CG2708 | Tom34 | 33561 (enhancer) | lethal |
| CG3035 | cm | 21139 (enhancer) | lethal |
| CG3158 | spn-E | 21374 (enhancer) | semilethal (8/57, 14 \%) |
| CG3297 | mnd | 42485 (enhancer) | semilethal (2/28, 7 \%) |
| CG3329 | Prosbeta2 | 24749 (enhancer) | lethal |
| CG3431 | Uch-L3 | 34618 (enhancer) | lethal |
| CG3644 | bic | 15453 (enhancer) | lethal |
| CG3820 | Nup214 | 41964 (enhancer) | n.a. |
| CG3923 | Exp6 | 34737 (enhancer) | n.a. |
| CG4001 | Pfk | 3016 (enhancer) | semilethal (7/42, 17 \%) |
| CG4035 | eIF-4E | 7800 (enhancer) | lethal |
| CG4062 | Aats-val | 21782 (enhancer) | lethal |
| CG4152 | $l(2) 35 D f$ | 21793 (enhancer) | n.a. |
| CG4180 | $1(2) 35 \mathrm{Bg}$ | 41980 (enhancer) | n.a. |
| CG4482 | mol | 33186 (enhancer) | lethal |
| CG4843 | Tm2 | 42010 (enhancer) | lethal |
| CG5163 | TfIIA-S | 12746 (enhancer) | lethal |
| CG5429 | Atg 6 | 22123 (enhancer) | lethal |
| CG5499 | His2Av | 12768 (enhancer) | lethal |
| CG5553 | DNApol-alpha60 | 30623 (enhancer) | lethal |


| CG5748 | Hsf | 37699 (enhancer) <br> 48692 (enhancer) | lethal semilethal (23/83, 28 \%) |
| :---: | :---: | :---: | :---: |
| CG5753 | stau | 27503 (enhancer) | lethal |
| CG6146 | Top1 | 10639 (enhancer) | lethal |
| CG6603 | Hsc70Cb | 27680 (enhancer) | n.a. |
| CG6827 | Nrx-IV | 9039 (enhancer) | lethal |
| CG6944 | Lam | 45635 (enhancer) | lethal |
| CG7085 | l(2)s5379 | 46150 (enhancer) | lethal |
| CG7128 | Taf8 | 27870 (enhancer) | lethal |
| CG7176 | Idh | 42915 (enhancer) | semilethal (12/74, 16 \%) |
| CG7436 | Nmt | 28019 (enhancer) | n.a. |
| CG7480 | Pgant35A | 44263 (enhancer) | semilethal (10/83, 12 \%) |
| CG7516 | l(2)34Fd | 28058 (enhancer) | lethal |
| CG7769 | pic | 44976 (enhancer) | n.a. |
| CG7807 | AP-2 | 41130 (enhancer) | lethal |
| CG8151 | Tfb1 | 12581 (enhancer) | lethal |
| CG8384 | gro | 6315 (enhancer) | lethal |
| CG8887 | ash1 | 28982 (enhancer) | semilethal (20/86, 23 \%) |
| CG8933 | exd | 7802 (enhancer) | lethal |
| CG9191 | Klp61F | 52549 (enhancer) | lethal |
| CG9998 | U2af50 | 24177 (enhancer) | n.a. |
| CG10033 | for | 38319 (enhancer) | n.a. |
| CG10037 | vvl | 10756 (enhancer) <br> 47182 (no effect) | lethal <br> lethal |
| CG10687 | Aats-asn | 31320 (enhancer) | lethal |
| CG10719 | brat | 31333 (enhancer) | lethal |
| CG10726 | barr | 26759 (enhancer) | lethal |
| CG10776 | wit | 42244 (no effect) <br> 856 (enhancer) | lethal <br> lethal |
| CG10975 | Ptp69D | 4789 (enhancer) <br> 942 (no effect) <br> 27090 (no effect) | lethal semilethal (2/15, 13 \%) semilethal (19/73, 26 \%) |
| CG11278 | Syx13 | 8361 (enhancer) | n.a. |
| CG11282 | caps | 3046 (enhancer) <br> 27097 (no effect) | $\begin{aligned} & \text { semilethal (8/39, } 20 \%) \\ & \text { n.a. } \end{aligned}$ |
| CG11546 | kermit | 31522 (enhancer) | lethal |
| CG11989 | vnc | 31619 (enhancer) | lethal |


| CG12238 | $e(y) 3$ | 38637 (enhancer) | lethal |
| :---: | :---: | :---: | :---: |
| CG12283 | kek1 | 36252 (enhancer) <br> 4761 (no effect) <br> 43521 (no effect) | ```n.a. semilethal (7/77, 9 %) n.a.``` |
| CG12298 | sub | 45402 (enhancer) | semilethal (11/49, 22 \%) |
| CG14206 | RpS10b | 49844 (enhancer) | lethal |
| CG17743 | pho | 39529 (enhancer) | lethal |
| CG31000 | heph | 33735 (enhancer) | lethal |

## IV List of screened RNAi lines obtained from the VDRC as human orthologue sublibrary

## Appendix Table 3.

| CG10001 | 1326 | CG10126 | 44104 | CG10250 | 51311 | CG10376 | 35473 | CG10546 | 31258 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| CG10002 | 37063 | CG10128 | 8868 | CG10251 | 9534 | CG10377 | 16040 | CG10546 | 49655 |
| CG10002 | 49961 | CG10130 | 8785 | CG10253 | 3321 | CG10379 | 16044 | CG10549 | 40789 |
| CG10006 | 44539 | CG10133 | 18014 | CG10254 | 15992 | CG10383 | 33911 | CG10555 | 16961 |
| CG10009 | 17954 | CG10137 | 38342 | CG10255 | 18600 | CG10384 | 16045 | CG10555 | 50115 |
| CG10011 | 45096 | CG10142 | 15246 | CG10257 | 8710 | CG10385 | 9239 | CG10564 | 51979 |
| CG10018 | 37591 | CG10143 | 51564 | CG10260 | 15993 | CG10390 | 37563 | CG10565 | 38393 |
| CG10021 | 3774 | CG10144 | 18019 | CG10261 | 2907 | CG10392 | 18611 | CG10566 | 27281 |
| CG10023 | 17957 | CG10145 | 15194 | CG10262 | 37672 | CG10393 | 11796 | CG1057 | 27284 |
| CG10029 | 16583 | CG10149 | 18022 | CG10272 | 16001 | CG10395 | 31244 | CG10571 | 49078 |
| CG10030 | 44480 | CG10155 | 18024 | CG10272 | 47199 | CG10399 | 18617 | CG10572 | 45370 |
| CG10032 | 51688 | CG10157 | 14004 | CG10275 | 37283 | CG10406 | 23363 | CG10573 | 31266 |
| CG10033 | 38319 | CG10158 | 47388 | CG10275 | 36246 | CG10406 | 50865 | CG10574 | 39053 |
| CG10034 | 30525 | CG10160 | 31192 | CG10277 | 7518 | CG10413 | 3882 | CG10575 | 31270 |
| CG10036 | 15425 | CG10162 | 23362 | CG10278 | 10418 | CG10414 | 47391 | CG10576 | 28761 |
| CG10037 | 10756 | CG10165 | 3909 | CG10280 | 5733 | CG10415 | 12591 | CG10578 | 31271 |
| CG10037 | 47182 | CG10166 | 46385 | CG10281 | 51209 | CG10417 | 27259 | CG10579 | 47859 |
| CG1004 | 51953 | CG10166 | 13731 | CG10286 | 16002 | CG10418 | 50245 | CG1058 | 8549 |
| CG10043 | 17966 | CG10168 | 1151 | CG10289 | 16006 | CG10419 | 47373 | CG10580 | 51977 |
| CG10047 | 33317 | CG1017 | 15610 | CG10293 | 13756 | CG10420 | 1753 | CG10581 | 18650 |
| CG10050 | 30020 | CG10170 | 1141 | CG10295 | 12553 | CG10423 | 12795 | CG10581 | 48315 |
| CG10052 | 44717 | CG10174 | 31195 | CG10298 | 28706 | CG10426 | 16048 | CG10582 | 52094 |
| CG10053 | 50811 | CG10175 | 1140 | CG1030 | 46499 | CG10435 | 38384 | CG10583 | 45092 |
| CG10053 | 17972 | CG10178 | 8064 | CG1030 | 3033 | CG10438 | 23296 | CG10584 | 28681 |
| CG10055 | 17973 | CG10181 | 9019 | CG10302 | 22837 | CG10443 | 36270 | CG10585 | 31273 |
| CG10060 | 28150 | CG10184 | 3311 | CG10305 | 16012 | CG10444 | 4722 | CG10588 | 18655 |
| CG10061 | 17975 | CG10185 | 18135 | CG10308 | 31216 | CG10446 | 27049 | CG1059 | 39711 |
| CG10062 | 4697 | CG10188 | 18029 | CG1031 | 31220 | CG10446 | 3066 | CG10590 | 5035 |
| CG10064 | 38322 | CG10189 | 51692 | CG10315 | 27152 | CG10447 | 45660 | CG10592 | 38171 |
| CG10066 | 23303 | CG1019 | 18593 | CG10315 | 48708 | CG10449 | 7183 | CG10593 | 3324 |
| CG10067 | 17979 | CG10191 | 38353 | CG10318 | 15451 | CG10459 | 41451 | CG10594 | 51081 |
| CG10068 | 15948 | CG10192 | 18031 | CG10320 | 8837 | CG10463 | 31248 | CG10597 | 6157 |
| CG10069 | 6591 | CG10198 | 31198 | CG10324 | 31226 | CG10466 | 23367 | CG10600 | 31276 |
| CG10072 | 29434 | CG10203 | 31202 | CG10325 | 51900 | CG10467 | 27263 | CG10601 | 50133 |
| CG10073 | 11133 | CG10206 | 31204 | CG10326 | 5894 | CG10470 | 3897 | CG10601 | 22841 |
| CG10075 | 50067 | CG1021 | 37336 | CG10326 | 47194 | CG10473 | 16052 | CG10602 | 31280 |
| CG10075 | 15399 | CG10210 | 38356 | CG10327 | 38377 | CG10474 | 41455 | CG10603 | 31285 |
| CG10076 | 44092 | CG10211 | 12352 | CG10333 | 18132 | CG10479 | 45098 | CG10604 | 15716 |
| CG10078 | 48823 | CG10212 | 10711 | CG10335 | 40612 | CG10480 | 38388 | CG10605 | 2931 |
| CG10078 | 17981 | CG10215 | 12622 | CG10336 | 16019 | CG10483 | 33276 | CG10610 | 31286 |
| CG10079 | 43268 | CG10220 | 45999 | CG10338 | 3215 | CG10484 | 49423 | CG10616 | 36605 |
| CG10080 | 46133 | CG10221 | 1163 | CG10340 | 16020 | CG10489 | 13625 | CG10619 | 45859 |
| CG10081 | 10066 | CG10222 | 18038 | CG10341 | 52092 | CG1049 | 18628 | CG10620 | 5236 |
| CG10082 | 38326 | CG10223 | 30625 | CG10343 | 38380 | CG10491 | 50358 | CG10621 | 31291 |
| CG10083 | 38330 | CG10225 | 38363 | CG10344 | 48026 | CG10492 | 12357 | CG10622 | 30889 |
| CG10084 | 38336 | CG10228 | 38365 | CG10346 | 31228 | CG10493 | 45364 | CG10623 | 31294 |
| CG10089 | 17991 | CG10229 | 38368 | CG10347 | 16025 | CG10495 | 18631 | CG10624 | 44928 |
| CG1009 | 35354 | CG10230 | 31206 | CG10348 | 39663 | CG10497 | 13322 | CG10626 | 22845 |
| CG10090 | 37346 | CG10231 | 18041 | CG10353 | 5550 | CG10505 | 6593 | CG10627 | 31298 |
| CG10096 | 50765 | CG10234 | 37124 | CG10354 | 27254 | CG10510 | 18635 | CG10628 | 47392 |
| CG10097 | 6090 | CG10236 | 18873 | CG10355 | 9308 | CG10517 | 44107 | CG1063 | 6484 |
| CG10098 | 50126 | CG10238 | 15990 | CG10361 | 16034 | CG10523 | 47636 | CG10635 | 35481 |
| CG10103 | 31174 | CG10240 | 33238 | CG10362 | 8317 | CG10524 | 33837 | CG10635 | 46220 |
| CG10104 | 13969 | CG10242 | 50169 | CG10363 | 13466 | CG10531 | 16071 | CG10637 | 35482 |
| CG10105 | 18002 | CG10242 | 4880 | CG10365 | 16036 | CG10532 | 16073 | CG10638 | 31306 |
| CG10106 | 7934 | CG10243 | 7398 | CG10369 | 3886 | CG10535 | 45366 | CG10639 | 30737 |
| CG10107 | 18004 | CG10243 | 49532 | CG10371 | 47623 | CG10536 | 16078 | CG1064 | 12645 |
| CG10110 | 18009 | CG10245 | 3313 | CG10372 | 31238 | CG10537 | 41101 | CG10640 | 30890 |
| CG10117 | 4871 | CG10246 | 29980 | CG10373 | 6375 | CG10539 | 18126 | CG10641 | 31307 |
| CG10118 | 3308 | CG10246 | 50262 | CG10374 | 48109 | CG10541 | 31253 | CG10642 | 45372 |
| CG10122 | 12688 | CG10247 | 3317 | CG10374 | 30884 | CG10542 | 15620 | CG10645 | 31311 |
| CG10123 | 10635 | CG10249 | 15009 | CG10375 | 16039 | CG10545 | 31257 | CG10646 | 38396 |


| CG1065 | 27298 | CG10798 | 2947 | CG10960 | 8359 | CG11094 | 11096 | CG11221 | 42947 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| CG10653 | 35483 | CG10803 | 27318 | CG10961 | 16125 | CG11095 | 37910 | CG11228 | 7823 |
| CG10655 | 2985 | CG10805 | 17000 | CG10962 | 30850 | CG11096 | 38444 | CG11233 | 45386 |
| CG10657 | 3326 | CG10806 | 33149 | CG10966 | 3024 | CG11098 | 7625 | CG11236 | 38460 |
| CG1066 | 46889 | CG10808 | 8784 | CG10967 | 16133 | CG11099 | 9272 | CG11237 | 38462 |
| CG10662 | 31318 | CG10809 | 38408 | CG10971 | 16138 | CG11102 | 13478 | CG11242 | 38463 |
| CG10663 | 27299 | CG10811 | 17002 | CG10973 | 41463 | CG11103 | 5562 | CG11246 | 11203 |
| CG10664 | 3923 | CG10814 | 27319 | CG10975 | 4789 | CG11105 | 51705 | CG11250 | 35501 |
| CG10667 | 46522 | CG1082 | 45102 | CG10975 | 942 | CG11107 | 44119 | CG11251 | 38467 |
| CG10670 | 47601 | CG10823 | 1783 | CG10975 | 27090 | CG11109 | 17528 | CG11253 | 31473 |
| CG10671 | 44435 | CG10824 | 16588 | CG10977 | 37238 | CG11110 | 3801 | CG11254 | 18198 |
| CG10672 | 18661 | CG10825 | 31360 | CG10979 | 16144 | CG11111 | 6226 | CG11255 | 17534 |
| CG10673 | 27301 | CG10827 | 30823 | CG1098 | 27346 | CG11115 | 49942 | CG11257 | 31475 |
| CG10674 | 11366 | CG10830 | 31362 | CG10981 | 16149 | CG1112 | 42942 | CG11258 | 23376 |
| CG10679 | 28445 | CG10833 | 7870 | CG10984 | 15627 | CG11121 | 8950 | CG11259 | 17537 |
| CG10681 | 27305 | CG10837 | 31364 | CG10986 | 31390 | CG11123 | 18142 | CG1126 | 18200 |
| CG10682 | 27306 | CG10838 | 28289 | CG10988 | 2983 | CG11124 | 13911 | CG11262 | 16912 |
| CG10685 | 17847 | CG1084 | 40613 | CG1099 | 16158 | CG11125 | 18143 | CG11263 | 31112 |
| CG10686 | 31319 | CG10840 | 31365 | CG10990 | 16162 | CG11128 | 45587 | CG11265 | 41097 |
| CG10687 | 31320 | CG10846 | 8057 | CG10992 | 45345 | CG11130 | 18145 | CG11266 | 12945 |
| CG10688 | 39715 | CG10847 | 15291 | CG10993 | 16165 | CG11133 | 18150 | CG11267 | 47087 |
| CG10689 | 31324 | CG10849 | 7480 | CG10996 | 16168 | CG11136 | 44991 | CG11268 | 5240 |
| CG10691 | 12358 | CG10850 | 18847 | CG10997 | 28303 | CG11136 | 4758 | CG11270 | 8322 |
| CG10692 | 37816 | CG10859 | 27322 | CG10999 | 16171 | CG11137 | 48241 | CG11274 | 6439 |
| CG10693 | 6723 | CG1086 | 13326 | CG1100 | 18676 | CG11137 | 8464 | CG11276 | 35718 |
| CG10694 | 13649 | CG10862 | 31373 | CG11001 | 45015 | CG11138 | 18154 | CG11276 | 50021 |
| CG10695 | 27307 | CG10863 | 48619 | CG11006 | 31394 | CG11139 | 17530 | CG11278 | 8361 |
| CG10697 | 3329 | CG10866 | 3234 | CG11007 | 40833 | CG11140 | 37726 | CG1128 | 6420 |
| CG10698 | 44309 | CG10868 | 45009 | CG11009 | 16173 | CG11141 | 18157 | CG11280 | 5242 |
| CG10699 | 45231 | CG10869 | 27326 | CG1101 | 12031 | CG11143 | 5616 | CG11281 | 5247 |
| CG10701 | 37917 | CG10872 | 30717 | CG11010 | 47537 | CG11144 | 1793 | CG11282 | 3046 |
| CG10702 | 3691 | CG10873 | 38235 | CG11015 | 30892 | CG11146 | 50598 | CG11284 | 31482 |
| CG10703 | 35486 | CG10877 | 45750 | CG1102 | 18970 | CG11148 | 18159 | CG1129 | 38471 |
| CG10706 | 28155 | CG10881 | 35495 | CG11024 | 46489 | CG11149 | 7882 | CG11290 | 37527 |
| CG1071 | 45743 | CG10882 | 37543 | CG11025 | 45497 | CG11151 | 18129 | CG11294 | 10497 |
| CG10711 | 16846 | CG10887 | 16082 | CG11027 | 12931 | CG11153 | 19022 | CG11295 | 31484 |
| CG10716 | 38399 | CG10889 | 27329 | CG11029 | 44228 | CG11154 | 37812 | CG11299 | 38481 |
| CG10718 | 31329 | CG1089 | 27332 | CG11030 | 43530 | CG11156 | 31432 | CG1130 | 41070 |
| CG10719 | 31333 | CG10890 | 50554 | CG11033 | 31402 | CG11162 | 8326 | CG11301 | 15344 |
| CG1072 | 46755 | CG10895 | 44981 | CG11034 | 37941 | CG11163 | 13311 | CG11305 | 18043 |
| CG1072 | 9830 | CG10897 | 38413 | CG11035 | 8478 | CG11164 | 3180 | CG11306 | 8448 |
| CG10721 | 33098 | CG10898 | 13643 | CG11041 | 45061 | CG11165 | 18166 | CG11308 | 31487 |
| CG10722 | 44128 | CG10899 | 16084 | CG11043 | 23370 | CG11166 | 31435 | CG11309 | 7513 |
| CG10724 | 22850 | CG1090 | 26783 | CG11044 | 16178 | CG11168 | 18170 | CG1131 | 18048 |
| CG10726 | 26759 | CG10903 | 27334 | CG11048 | 31407 | CG11170 | 41302 | CG11312 | 31488 |
| CG10728 | 50141 | CG10907 | 16085 | CG1105 | 28305 | CG11172 | 30566 | CG11315 | 38273 |
| CG10732 | 18664 | CG10908 | 44211 | CG11050 | 12371 | CG11173 | 18172 | CG11315 | 46906 |
| CG10739 | 31337 | CG10909 | 17522 | CG11052 | 23373 | CG11176 | 18175 | CG11318 | 3395 |
| CG10742 | 10140 | CG1091 | 16088 | CG11055 | 18686 | CG11177 | 45975 | CG11319 | 7621 |
| CG10743 | 16850 | CG10910 | 46896 | CG11058 | 31409 | CG11178 | 18178 | CG1132 | 30553 |
| CG10747 | 38403 | CG10913 | 3245 | CG11059 | 36348 | CG11180 | 31438 | CG11320 | 18054 |
| CG10749 | 27311 | CG10914 | 38419 | CG11059 | 37291 | CG11181 | 18179 | CG11321 | 18055 |
| CG10750 | 27313 | CG10915 | 31377 | CG1106 | 37867 | CG11182 | 15318 | CG11323 | 18057 |
| CG10751 | 22761 | CG10918 | 23190 | CG11061 | 38441 | CG11183 | 31442 | CG11324 | 18061 |
| CG10753 | 31343 | CG10920 | 16091 | CG11062 | 12174 | CG11184 | 31444 | CG11325 | 9546 |
| CG10754 | 31347 | CG10922 | 2989 | CG11063 | 38442 | CG11186 | 15919 | CG11326 | 7535 |
| CG10756 | 44466 | CG10923 | 52105 | CG11064 | 6878 | CG11188 | 18182 | CG11329 | 18065 |
| CG10757 | 45494 | CG10924 | 13929 | CG11069 | 51367 | CG1119 | 10942 | CG1133 | 51292 |
| CG1076 | 40807 | CG10927 | 16094 | CG1107 | 16182 | CG11190 | 14821 | CG11333 | 44110 |
| CG10760 | 13996 | CG1093 | 27335 | CG11070 | 31413 | CG11194 | 30561 | CG11334 | 38485 |
| CG10761 | 5474 | CG10931 | 52107 | CG11077 | 42671 | CG11196 | 36497 | CG11335 | 17259 |
| CG10772 | 22853 | CG10932 | 16099 | CG11079 | 23374 | CG11197 | 41368 | CG11337 | 16420 |
| CG10776 | 42244 | CG10938 | 16104 | CG1108 | 16184 | CG11198 | 8105 | CG11339 | 45390 |
| CG10776 | 865 | CG10939 | 16958 | CG11081 | 27238 | CG11199 | 51707 | CG1134 | 7481 |
| CG10777 | 46933 | CG10947 | 16117 | CG11081 | 4740 | CG1120 | 18184 | CG11342 | 38488 |
| CG10778 | 3166 | CG10948 | 31388 | CG11082 | 45349 | CG11200 | 4725 | CG11348 | 39421 |
| CG1078 | 27317 | CG10950 | 41462 | CG11084 | 11099 | CG11201 | 31456 | CG11348 | 33824 |
| CG10791 | 18140 | CG10951 | 16120 | CG11085 | 10493 | CG11202 | 37656 | CG1135 | 15613 |
| CG10792 | 15444 | CG10952 | 9127 | CG11086 | 18690 | CG11206 | 42943 | CG11356 | 33812 |
| CG10793 | 31351 | CG10954 | 45013 | CG1109 | 16186 | CG11208 | 4720 | CG11357 | 5027 |
| CG10795 | 8833 | CG10955 | 27341 | CG11092 | 16188 | CG11210 | 7363 | CG11360 | 38491 |
| CG10797 | 26299 | CG10958 | 26763 | CG11093 | 15478 | CG11217 | 28762 | CG11367 | 40900 |


| CG11367 | 40900 | CG11546 | 31522 | CG11755 | 17555 | CG11909 | 14735 | CG12082 | 17568 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| CG11374 | 15561 | CG11547 | 31525 | CG11757 | 16296 | CG11913 | 16366 | CG12083 | 7434 |
| CG11376 | 40673 | CG11551 | 48368 | CG11759 | 16298 | CG11922 | 13721 | CG12084 | 31646 |
| CG11386 | 38493 | CG11556 | 52439 | CG11760 | 11925 | CG11926 | 38600 | CG12085 | 20144 |
| CG11386 | 49657 | CG11561 | 9542 | CG11761 | 9963 | CG1193 | 31598 | CG12090 | 16390 |
| CG11386 | 49657 | CG11567 | 44232 | CG11763 | 45981 | CG11935 | 15472 | CG12091 | 13985 |
| CG11386 | 38493 | CG11568 | 30060 | CG11765 | 18708 | CG11936 | 37927 | CG12092 | 35514 |
| CG11387 | 5687 | CG11569 | 8706 | CG11770 | 16801 | CG11940 | 16369 | CG12093 | 41293 |
| CG11388 | 3604 | CG11573 | 52126 | CG11771 | 18946 | CG11943 | 38608 | CG12099 | 18734 |
| CG1139 | 8907 | CG11576 | 7577 | CG11778 | 7374 | CG11949 | 9787 | CG1210 | 18736 |
| CG11392 | 42959 | CG11577 | 14334 | CG11779 | 16309 | CG11951 | 16532 | CG12101 | 18739 |
| CG11393 | 33839 | CG11579 | 7767 | CG11780 | 42092 | CG11951 | 48791 | CG12106 | 16400 |
| CG11393 | 47725 | CG1158 | 9455 | CG11781 | 49584 | CG11956 | 3335 | CG12107 | 8731 |
| CG11395 | 16696 | CG11582 | 35379 | CG11783 | 10958 | CG11958 | 1335 | CG12108 | 5499 |
| CG11396 | 18073 | CG11586 | 31531 | CG11788 | 47245 | CG11963 | 20097 | CG12109 | 20151 |
| CG11397 | 10937 | CG11588 | 8348 | CG1179 | 14123 | CG11963 | 50300 | CG12110 | 38626 |
| CG11399 | 31489 | CG11590 | 19867 | CG1179 | 49832 | CG11964 | 35511 | CG12111 | 16513 |
| CG11401 | 16768 | CG11591 | 22770 | CG11793 | 31551 | CG11968 | 20130 | CG12113 | 18742 |
| CG11408 | 45394 | CG11592 | 2495 | CG11796 | 31563 | CG11975 | 26799 | CG12116 | 6498 |
| CG1141 | 18081 | CG11593 | 16227 | CG11799 | 45697 | CG11981 | 31608 | CG12117 | 17018 |
| CG11412 | 49370 | CG11594 | 28311 | CG1180 | 13842 | CG11982 | 38623 | CG12118 | 13340 |
| CG11416 | 40674 | CG11596 | 35505 | CG11801 | 30042 | CG11984 | 31615 | CG1212 | 41479 |
| CG11417 | 18087 | CG11597 | 38541 | CG11802 | 31570 | CG11985 | 22773 | CG12125 | 5492 |
| CG11418 | 31497 | CG11598 | 49831 | CG11804 | 16313 | CG11986 | 20136 | CG12127 | 3295 |
| CG11419 | 20027 | CG11600 | 42964 | CG11807 | 38564 | CG11987 | 10735 | CG12128 | 42979 |
| CG11423 | 38500 | CG11601 | 42737 | CG11811 | 30917 | CG11988 | 10662 | CG12129 | 17065 |
| CG11426 | 42600 | CG11601 | 50814 | CG11814 | 38567 | CG11989 | 31619 | CG1213 | 6487 |
| CG11427 | 38504 | CG11606 | 44093 | CG11819 | 31571 | CG11990 | 28318 | CG12130 | 7388 |
| CG11428 | 38508 | CG11607 | 15876 | CG11820 | 39724 | CG11992 | 49413 | CG12131 | 45744 |
| CG11430 | 12221 | CG11608 | 19932 | CG11821 | 26796 | CG11994 | 18719 | CG12132 | 31651 |
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| CG17597 | 49204 | CG1785 | 52467 | CG18174 | 19272 | CG1848 | 25343 | CG18769 | 9501 |
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| CG1946 | 33428 | CG2095 | 45032 | CG2248 | 49247 | CG2702 | 33557 | CG2931 | 20946 |
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| CG1970 | 38224 | CG2124 | 14869 | CG2316 | 12170 | CG2781 | 48139 | CG2964 | 42293 |
| CG1972 | 33450 | CG2125 | 51479 | CG2321 | 33523 | CG2789 | 2507 | CG2969 | 42751 |
| CG1973 | 19275 | CG2126 | 47598 | CG2330 | 46555 | CG2790 | 20903 | CG2970 | 48124 |
| CG1975 | 20771 | CG2128 | 20814 | CG2331 | 24354 | CG2791 | 42622 | CG2970 | 43497 |
| CG1977 | 25387 | CG2128 | 50213 | CG2358 | 9055 | CG2803 | 25160 | CG2971 | 14637 |
| CG1981 | 13657 | CG2136 | 33486 | CG2371 | 15483 | CG2807 | 25161 | CG2972 | 48771 |
| CG1982 | 47191 | CG2137 | 41235 | CG2380 | 14918 | CG2812 | 52485 | CG2974 | 32484 |
| CG1982 | 3761 | CG2140 | 43065 | CG2381 | 24989 | CG2816 | 29700 | CG2975 | 2601 |
| CG1983 | 24980 | CG2143 | 20819 | CG2397 | 4019 | CG2818 | 25441 | CG2976 | 20962 |
| CG1986 | 51669 | CG2144 | 3996 | CG2412 | 1494 | CG2826 | 46588 | CG2980 | 25470 |
| CG1989 | 46977 | CG2145 | 14874 | CG2453 | 20872 | CG2827 | 25444 | CG2984 | 33599 |
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| CG1998 | 37395 | CG2152 | 19121 | CG2488 | 10461 | CG2835 | 24959 | CG2988 | 9419 |
| CG2005 | 27208 | CG2155 | 3349 | CG2493 | 3929 | CG2839 | 17952 | CG2988 | 48649 |
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| CG2009 | 48037 | CG2161 | 20826 | CG2508 | 52280 | CG2845 | 20909 | CG2993 | 33605 |
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| CG2013 | 23229 | CG2163 | 33499 | CG2522 | 14877 | CG2849 | 43623 | CG2995 | 25473 |
| CG2014 | 50731 | CG2165 | 30203 | CG2525 | 15880 | CG2852 | 15069 | CG2996 | 14613 |
| CG2014 | 40165 | CG2168 | 50311 | CG2528 | 33532 | CG2854 | 41703 | CG2998 | 42419 |
| CG2017 | 20780 | CG2168 | 43627 | CG2534 | 7769 | CG2855 | 16820 | CG2998 | 35783 |
| CG2019 | 10004 | CG2173 | 36516 | CG2540 | 33536 | CG2856 | 10970 | CG2999 | 33609 |
| CG2022 | 9049 | CG2174 | 37530 | CG2543 | 25417 | CG2857 | 7417 | CG3000 | 25553 |
| CG2025 | 25392 | CG2177 | 51084 | CG2551 | 29635 | CG2859 | 15334 | CG30000 | 41965 |
| CG2028 | 13664 | CG2179 | 20829 | CG2574 | 40173 | CG2861 | 20918 | CG30005 | 25555 |
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| CG2062 | 4018 | CG2204 | 19124 | CG2647 | 5709 | CG2901 | 12209 | CG30022 | 30880 |
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| CG2065 | 19119 | CG2212 | 5469 | CG2662 | 33544 | CG2905 | 52486 | CG3004 | 46557 |
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| CG2075 | 13673 | CG2221 | 1054 | CG2671 | 51249 | CG2913 | 7028 | CG30048 | 4634 |
| CG2076 | 50221 | CG2221 | 27247 | CG2674 | 7167 | CG2914 | 51225 | CG30051 | 43066 |
| CG2076 | 5537 | CG2221 | 39505 | CG2675 | 10149 | CG2916 | 25456 | CG30051 | 49256 |
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| CG2083 | 20798 | CG2239 | 43044 | CG2681 | 33549 | CG2919 | 25457 | CG3006 | 38148 |
| CG2086 | 27084 | CG2241 | 49245 | CG2682 | 44783 | CG2921 | 33585 | CG30062 | 15433 |
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| CG2093 | 29971 | CG2246 | 24987 | CG2701 | 25433 | CG2930 | 7031 | CG3008 | 43731 |


| CG30084 | 36564 | CG30354 | 23575 | CG3093 | 33733 | CG31183 | 36323 | CG31447 | 25761 |
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| CG30097 | 8977 | CG30368 | 13913 | CG31000 | 33735 | CG31194 | 45433 | CG31452 | 3744 |
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| CG30103 | 4949 | CG30387 | 48313 | CG31009 | 27216 | CG31212 | 40214 | CG31477 | 42108 |
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| CG30105 | 19204 | CG30389 | 19002 | CG31009 | 3733 | CG31229 | 9660 | CG3149 | 47560 |
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| CG30109 | 50925 | CG30392 | 12862 | CG31012 | 38854 | CG31234 | 30389 | CG3151 | 33939 |
| CG3011 | 19208 | CG30394 | 3470 | CG31014 | 30975 | CG31237 | 23308 | CG31522 | 37329 |
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| CG30147 | 25580 | CG30420 | 7414 | CG31040 | 39926 | CG31278 | 38938 | CG31547 | 8551 |
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| CG31679 | 21392 | CG31832 | 47633 | CG3204 | 45230 | CG3220 | 23511 | CG32446 | 23058 |
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| CG3169 | 10818 | CG31855 | 34295 | CG32064 | 34051 | CG32226 | 34464 | CG32473 | 8281 |
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| CG3292 | 19989 | CG3353 | 41920 | CG3460 | 46165 | CG3634 | 16720 | CG3762 | 34389 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
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| CG3299 | 34585 | CG3356 | 34601 | CG3476 | 2734 | CG3644 | 15453 | CG3772 | 7238 |
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| CG3301 | 43225 | CG3359 | 37889 | CG3480 | 26205 | CG3648 | 10156 | CG3774 | 30238 |
| CG33012 | 48408 | CG3360 | 8287 | CG3483 | 41939 | CG3652 | 35574 | CG3776 | 38269 |
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| CG3917 | 34731 | CG4039 | 13661 | CG4163 | 51493 | CG4299 | 21827 | CG4485 | 12498 |
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| CG3924 | 30454 | CG4045 | 34784 | CG4170 | 26408 | CG4311 | 26504 | CG4501 | 34853 |
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| CG4759 | 21943 | CG4908 | 44845 | CG5029 | 29733 | CG5174 | 29752 | CG5317 | 46572 |
| CG4760 | 21536 | CG4909 | 26657 | CG5029 | 33803 | CG5178 | 9780 | CG5319 | 34986 |


| CG5320 | 22059 | CG5442 | 40590 | CG5560 | 14814 | CG5695 | 37535 | CG5840 | 22214 |
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| CG5327 | 25972 | CG5450 | 42114 | CG5569 | 21568 | CG5707 | 22195 | CG5850 | 1456 |
| CG5330 | 27392 | CG5451 | 42035 | CG5571 | 22160 | CG5708 | 23425 | CG5851 | 42051 |
| CG5333 | 22066 | CG5452 | 39137 | CG5577 | 22163 | CG5714 | 28398 | CG5855 | 37722 |
| CG5335 | 22068 | CG5454 | 22132 | CG5580 | 41845 | CG5715 | 14710 | CG5857 | 3408 |
| CG5336 | 10455 | CG5455 | 35011 | CG5582 | 5322 | CG5718 | 34239 | CG5859 | 45677 |
| CG5337 | 22069 | CG5458 | 27420 | CG5583 | 10932 | CG5720 | 27487 | CG5861 | 6360 |
| CG5338 | 22074 | CG5461 | 19679 | CG5585 | 22166 | CG5721 | 27488 | CG5861 | 47665 |
| CG5339 | 22075 | CG5462 | 27424 | CG5586 | 22169 | CG5723 | 51173 | CG5862 | 45155 |
| CG5341 | 22077 | CG5462 | 27424 | CG5589 | 44322 | CG5725 | 44157 | CG5863 | 14366 |
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| CG5343 | 10468 | CG5465 | 14916 | CG5594 | 10278 | CG5730 | 27493 | CG5870 | 42053 |
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| CG5352 | 40587 | CG5474 | 12101 | CG5602 | 51315 | CG5735 | 27498 | CG5880 | 1264 |
| CG5353 | 7752 | CG5475 | 52277 | CG5603 | 15340 | CG5737 | 41048 | CG5882 | 27532 |
| CG5354 | 22095 | CG5475 | 34238 | CG5604 | 27467 | CG5741 | 5158 | CG5884 | 19730 |
| CG5355 | 40588 | CG5479 | 22138 | CG5605 | 45027 | CG5742 | 36428 | CG5884 | 19731 |
| CG5358 | 15645 | CG5481 | 11823 | CG5608 | 45569 | CG5744 | 23428 | CG5886 | 22216 |
| CG5359 | 21565 | CG5482 | 4991 | CG5610 | 1189 | CG5745 | 35034 | CG5887 | 47142 |
| CG5362 | 27399 | CG5483 | 22139 | CG5610 | 48159 | CG5748 | 37699 | CG5887 | 33338 |
| CG5363 | 41838 | CG5484 | 2679 | CG5613 | 24694 | CG5748 | 48692 | CG5889 | 27535 |
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| CG5366 | 12067 | CG5486 | 26027 | CG5626 | 27469 | CG5753 | 27503 | CG5892 | 6807 |
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| CG5374 | 34070 | CG5493 | 26082 | CG5634 | 8016 | CG5783 | 23429 | CG5902 | 6274 |
| CG5375 | 22099 | CG5495 | 27436 | CG5637 | 22693 | CG5784 | 49386 | CG5903 | 31098 |
| CG5377 | 34392 | CG5497 | 41800 | CG5638 | 1748 | CG5786 | 39001 | CG5904 | 35048 |
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| CG5379 | 14461 | CG5499 | 12768 | CG5640 | 37663 | CG5788 | 27515 | CG5906 | 5227 |
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| CG5384 | 27405 | CG5508 | 1316 | CG5642 | 46592 | CG5792 | 34143 | CG5911 | 42716 |
| CG5387 | 34990 | CG5510 | 28393 | CG5645 | 9289 | CG5793 | 35044 | CG5912 | 4819 |
| CG5389 | 22112 | CG5514 | 27445 | CG5650 | 35025 | CG5793 | 47883 | CG5912 | 36286 |
| CG5392 | 52427 | CG5515 | 27447 | CG5651 | 44325 | CG5794 | 27517 | CG5913 | 40336 |
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| CG5406 | 27406 | CG5524 | 13650 | CG5659 | 35029 | CG5803 | 3091 | CG5926 | 24996 |
| CG5407 | 27411 | CG5525 | 22155 | CG5660 | 29445 | CG5803 | 42229 | CG5927 | 17853 |
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| CG5429 | 22123 | CG5548 | 24714 | CG5680 | 34139 | CG5823 | 8301 | CG5949 | 41028 |
| CG5430 | 44712 | CG5549 | 8222 | CG5684 | 28396 | CG5826 | 27521 | CG5950 | 5150 |
| CG5432 | 27417 | CG5550 | 31000 | CG5685 | 42660 | CG5827 | 23590 | CG5952 | 17849 |
| CG5433 | 22125 | CG5553 | 30623 | CG5686 | 7777 | CG5828 | 27522 | CG5954 | 13994 |
| CG5434 | 21641 | CG5554 | 20101 | CG5687 | 33262 | CG5830 | 40611 | CG5955 | 15836 |
| CG5435 | 35005 | CG5555 | 35013 | CG5688 | 27482 | CG5832 | 16755 | CG5958 | 20982 |
| CG5439 | 27418 | CG5558 | 39560 | CG5690 | 28651 | CG5837 | 22207 | CG5960 | 20983 |
| CG5440 | 49030 | CG5558 | 46814 | CG5692 | 27486 | CG5838 | 22210 | CG5962 | 20989 |


| CG5965 | 20994 | CG6094 | 27564 | CG6213 | 25986 | CG6369 | 27598 | CG6513 | 34173 |
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| CG5966 | 13164 | CG6096 | 37691 | CG6218 | 35069 | CG6372 | 52508 | CG6514 | 27649 |
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| CG5970 | 20998 | CG6098 | 27566 | CG6222 | 10854 | CG6376 | 15887 | CG6516 | 27654 |
| CG5973 | 24998 | CG6106 | 30120 | CG6223 | 15419 | CG6378 | 16677 | CG6517 | 39622 |
| CG5974 | 2889 | CG6110 | 22221 | CG6224 | 22476 | CG6379 | 29611 | CG6518 | 2894 |
| CG5977 | 33110 | CG6113 | 31021 | CG6225 | 8276 | CG6380 | 29950 | CG6519 | 13860 |
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| CG5986 | 25479 | CG6120 | 3422 | CG6230 | 8897 | CG6386 | 48980 | CG6523 | 34174 |
| CG5987 | 21005 | CG6121 | 22233 | CG6232 | 31020 | CG6390 | 4385 | CG6524 | 33286 |
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| CG6008 | 5717 | CG6143 | 22245 | CG6264 | 5963 | CG6410 | 24701 | CG6550 | 4947 |
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| CG6083 | 27549 | CG6203 | 8933 | CG6349 | 11227 | CG6502 | 27645 | CG6652 | 26706 |
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| CG6092 | 44165 | CG6208 | 40348 | CG6363 | 43802 | CG6509 | 22496 | CG6659 | 8038 |
| CG6094 | 48721 | CG6210 | 5215 | CG6364 | 11693 | CG6512 | 8515 | CG6660 | 6835 |


| CG6662 | 26713 | CG6782 | 50714 | CG6914 | 35139 | CG7041 | 26097 | CG7168 | 27894 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| CG6664 | 12780 | CG6784 | 39312 | CG6915 | 35141 | CG7042 | 23452 | CG7172 | 27898 |
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| CG6673 | 41806 | CG6792 | 35118 | CG6928 | 5231 | CG7050 | 4306 | CG7183 | 40429 |
| CG6674 | 26716 | CG6794 | 30578 | CG6930 | 35147 | CG7050 | 36328 | CG7184 | 34373 |
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| CG6678 | 26719 | CG6798 | 1199 | CG6937 | 22315 | CG7052 | 35611 | CG7187 | 28610 |
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| CG6684 | 52603 | CG6811 | 34250 | CG6939 | 22317 | CG7054 | 40416 | CG7190 | 31070 |
| CG6686 | 21573 | CG6812 | 8534 | CG6944 | 45635 | CG7055 | 37684 | CG7192 | 52526 |
| CG6690 | 14439 | CG6814 | 22442 | CG6946 | 27752 | CG7056 | 15719 | CG7193 | 40434 |
| CG6691 | 3951 | CG6815 | 39675 | CG6948 | 22318 | CG7057 | 27820 | CG7194 | 24723 |
| CG6692 | 13959 | CG6816 | 5601 | CG6949 | 40405 | CG7059 | 21651 | CG7195 | 39303 |
| CG6693 | 27717 | CG6817 | 10102 | CG6950 | 22321 | CG7060 | 42883 | CG7195 | 28160 |
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| CG6704 | 46856 | CG6840 | 23290 | CG6971 | 48986 | CG7075 | 3666 | CG7215 | 27916 |
| CG6706 | 1784 | CG6841 | 34253 | CG6972 | 27772 | CG7076 | 28740 | CG7217 | 49806 |
| CG6707 | 44557 | CG6842 | 35126 | CG6975 | 6314 | CG7077 | 33835 | CG7217 | 35196 |
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| CG6726 | 34247 | CG6863 | 2656 | CG6999 | 41829 | CG7103 | 46875 | CG7234 | 7878 |
| CG6733 | 50172 | CG6866 | 22453 | CG7000 | 42496 | CG7106 | 45634 | CG7235 | 22539 |
| CG6737 | 46197 | CG6867 | 37416 | CG7002 | 37005 | CG7107 | 27853 | CG7238 | 34382 |
| CG6738 | 48918 | CG6868 | 1216 | CG7004 | 27785 | CG7108 | 43870 | CG7241 | 5851 |
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| CG6742 | 27738 | CG6871 | 6283 | CG7007 | 33342 | CG7112 | 35174 | CG7246 | 34256 |
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| CG6746 | 46513 | CG6878 | 9224 | CG7010 | 40410 | CG7121 | 17903 | CG7254 | 27928 |
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| CG6757 | 22412 | CG6894 | 9154 | CG7023 | 27799 | CG7137 | 27884 | CG7265 | 27932 |
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| CG6767 | 35112 | CG6903 | 3285 | CG7034 | 35162 | CG7152 | 27893 | CG7275 | 22561 |
| CG6768 | 13645 | CG6904 | 35136 | CG7035 | 22331 | CG7154 | 37669 | CG7277 | 30691 |
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| CG6772 | 30673 | CG6906 | 8357 | CG7037 | 22335 | CG7158 | 35179 | CG7280 | 18550 |
| CG6778 | 44603 | CG6907 | 22460 | CG7038 | 47159 | CG7161 | 35186 | CG7281 | 27937 |
| CG6779 | 37742 | CG6910 | 22464 | CG7038 | 40413 | CG7162 | 13054 | CG7281 | 48835 |
| CG6781 | 34227 | CG6913 | 46690 | CG7039 | 26007 | CG7163 | 38247 | CG7282 | 50406 |


| CG7282 | 27941 | CG7424 | 44630 | CG7568 | 45783 | CG7727 | 42673 | CG7851 | 33157 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| CG7283 | 52411 | CG7425 | 26011 | CG7571 | 37295 | CG7728 | 21048 | CG7852 | 7178 |
| CG7283 | 23458 | CG7427 | 28006 | CG7573 | 8337 | CG7729 | 37010 | CG7855 | 22590 |
| CG7285 | 13560 | CG7429 | 28008 | CG7577 | 36659 | CG7734 | 3226 | CG7860 | 34395 |
| CG7288 | 47663 | CG7430 | 28011 | CG7578 | 33634 | CG7735 | 43508 | CG7861 | 34388 |
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| CG7291 | 30725 | CG7432 | 31091 | CG7581 | 21037 | CG7737 | 49437 | CG7865 | 15782 |
| CG7292 | 35200 | CG7433 | 28014 | CG7582 | 1353 | CG7739 | 51521 | CG7867 | 28069 |
| CG7293 | 27943 | CG7436 | 28019 | CG7590 | 25506 | CG7740 | 51956 | CG7870 | 2802 |
| CG7301 | 39384 | CG7437 | 28023 | CG7595 | 9265 | CG7741 | 33655 | CG7872 | 9134 |
| CG7307 | 18553 | CG7438 | 12558 | CG7597 | 25510 | CG7742 | 25535 | CG7873 | 26019 |
| CG7317 | 28694 | CG7439 | 49473 | CG7598 | 14861 | CG7744 | 33247 | CG7875 | 1365 |
| CG7319 | 27949 | CG7441 | 28027 | CG7600 | 47473 | CG7747 | 44854 | CG7878 | 35288 |
| CG7322 | 9258 | CG7446 | 5329 | CG7601 | 4456 | CG7749 | 5098 | CG7879 | 15260 |
| CG7323 | 15631 | CG7449 | 40898 | CG7602 | 37594 | CG7749 | 3749 | CG7882 | 8103 |
| CG7324 | 31064 | CG7449 | 27065 | CG7605 | 43730 | CG7749 | 27113 | CG7883 | 40321 |
| CG7328 | 27951 | CG7449 | 9471 | CG7609 | 21041 | CG7757 | 25547 | CG7885 | 14000 |
| CG7329 | 48340 | CG7452 | 36595 | CG7610 | 16539 | CG7758 | 12823 | CG7886 | 22610 |
| CG7331 | 27955 | CG7457 | 46671 | CG7611 | 25511 | CG7760 | 37611 | CG7887 | 1374 |
| CG7332 | 27959 | CG7457 | 26740 | CG7614 | 12575 | CG7761 | 8692 | CG7887 | 43329 |
| CG7334 | 13375 | CG7459 | 5805 | CG7615 | 47312 | CG7762 | 25549 | CG7888 | 37264 |
| CG7335 | 27962 | CG7460 | 13115 | CG7616 | 41408 | CG7764 | 12596 | CG7891 | 26085 |
| CG7337 | 35204 | CG7461 | 28028 | CG7620 | 30391 | CG7765 | 44337 | CG7892 | 3002 |
| CG7338 | 27963 | CG7462 | 40638 | CG7621 | 48032 | CG7766 | 52573 | CG7893 | 6241 |
| CG7339 | 13629 | CG7464 | 10558 | CG7622 | 39914 | CG7768 | 35266 | CG7894 | 3060 |
| CG7340 | 3174 | CG7466 | 42462 | CG7623 | 12149 | CG7769 | 44976 | CG7894 | 869 |
| CG7343 | 35206 | CG7467 | 7810 | CG7625 | 30382 | CG7770 | 34203 | CG7894 | 42251 |
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| CG7345 | 10813 | CG7471 | 46930 | CG7628 | 49973 | CG7773 | 35267 | CG7896 | 36340 |
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| CG7351 | 27966 | CG7473 | 39151 | CG7632 | 45675 | CG7777 | 8124 | CG7899 | 3579 |
| CG7352 | 40636 | CG7480 | 44263 | CG7633 | 6178 | CG7779 | 29046 | CG7900 | 43157 |
| CG7354 | 14305 | CG7484 | 13358 | CG7635 | 9160 | CG7785 | 36650 | CG7904 | 37279 |
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| CG7360 | 40773 | CG7487 | 10614 | CG7637 | 23669 | CG7791 | 35272 | CG7910 | 51546 |
| CG7362 | 7556 | CG7490 | 28618 | CG7638 | 8235 | CG7793 | 42848 | CG7911 | 23075 |
| CG7364 | 7706 | CG7494 | 48961 | CG7639 | 33641 | CG7804 | 49886 | CG7912 | 1377 |
| CG7365 | 14318 | CG7494 | 28042 | CG7640 | 23671 | CG7804 | 28067 | CG7913 | 22614 |
| CG7367 | 43822 | CG7497 | 9374 | CG7642 | 25172 | CG7806 | 2804 | CG7914 | 22619 |
| CG7368 | 27978 | CG7499 | 9179 | CG7646 | 35742 | CG7807 | 41130 | CG7915 | 22620 |
| CG7371 | 27984 | CG7504 | 28050 | CG7650 | 41714 | CG7808 | 35278 | CG7917 | 22623 |
| CG7371 | 48711 | CG7507 | 28053 | CG7654 | 47988 | CG7809 | 22564 | CG7919 | 5117 |
| CG7375 | 35220 | CG7508 | 48674 | CG7655 | 12429 | CG7810 | 22565 | CG7921 | 1381 |
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| CG7378 | 35226 | CG7509 | 51585 | CG7659 | 12639 | CG7813 | 22566 | CG7925 | 52533 |
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| CG7379 | 27988 | CG7511 | 52654 | CG7662 | 46962 | CG7815 | 22567 | CG7927 | 22627 |
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| CG7387 | 27993 | CG7513 | 35250 | CG7664 | 26885 | CG7818 | 48559 | CG7931 | 21578 |
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| CG7392 | 35232 | CG7515 | 35251 | CG7669 | 29200 | CG7821 | 40641 | CG7935 | 38963 |
| CG7394 | 9210 | CG7516 | 28058 | CG7670 | 44595 | CG7823 | 46154 | CG7940 | 22633 |
| CG7395 | 9379 | CG7518 | 26745 | CG7671 | 33645 | CG7825 | 44723 | CG7943 | 28632 |
| CG7397 | 13429 | CG7519 | 21653 | CG7678 | 11649 | CG7826 | 28628 | CG7945 | 35296 |
| CG7398 | 30066 | CG7520 | 15927 | CG7686 | 33650 | CG7828 | 7728 | CG7946 | 35298 |
| CG7398 | 4769 | CG7530 | 5872 | CG7694 | 25520 | CG7830 | 4253 | CG7948 | 14021 |
| CG7398 | 6543 | CG7532 | 12405 | CG7698 | 39557 | CG7831 | 22570 | CG7949 | 21657 |
| CG7399 | 35240 | CG7536 | 11576 | CG7700 | 12152 | CG7832 | 49339 | CG7950 | 22635 |
| CG7400 | 48719 | CG7538 | 10967 | CG7704 | 45957 | CG7833 | 44030 | CG7951 | 46696 |
| CG7400 | 9406 | CG7542 | 47336 | CG7706 | 25524 | CG7834 | 36661 | CG7954 | 52538 |
| CG7402 | 37302 | CG7546 | 35253 | CG7708 | 30301 | CG7837 | 22573 | CG7955 | 40838 |
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| CG7413 | 10696 | CG7556 | 28621 | CG7716 | 25526 | CG7842 | 46334 | CG7960 | 13679 |
| CG7414 | 18804 | CG7560 | 28063 | CG7717 | 25528 | CG7842 | 14279 | CG7961 | 35306 |
| CG7415 | 35242 | CG7562 | 30441 | CG7718 | 25532 | CG7843 | 22574 | CG7962 | 5121 |
| CG7417 | 37555 | CG7564 | 29462 | CG7719 | 21046 | CG7845 | 22578 | CG7964 | 50645 |
| CG7420 | 46316 | CG7565 | 36291 | CG7722 | 25534 | CG7847 | 9921 | CG7966 | 22639 |
| CG7421 | 27995 | CG7565 | 8396 | CG7724 | 43927 | CG7849 | 22588 | CG7970 | 8857 |
| CG7423 | 28003 | CG7565 | 3707 | CG7726 | 45427 | CG7850 | 3018 | CG7971 | 34262 |


| CG7972 | 28072 | CG8111 | 29391 | CG8254 | 13716 | CG8372 | 8043 | CG8494 | 42609 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| CG7974 | 28074 | CG8112 | 37345 | CG8257 | 35865 | CG8376 | 37791 | CG8495 | 28849 |
| CG7975 | 40645 | CG8114 | 35349 | CG8257 | 50371 | CG8378 | 40705 | CG8497 | 24111 |
| CG7978 | 51974 | CG8116 | 45735 | CG8258 | 45789 | CG8379 | 35911 | CG8498 | 35388 |
| CG7979 | 5126 | CG8117 | 23254 | CG8261 | 28844 | CG8380 | 12082 | CG8500 | 28795 |
| CG7980 | 28169 | CG8117 | 47174 | CG8266 | 35867 | CG8383 | 35915 | CG8506 | 24114 |
| CG7986 | 22646 | CG8127 | 44851 | CG8267 | 35870 | CG8384 | 6315 | CG8507 | 42640 |
| CG7988 | 46277 | CG8128 | 47740 | CG8268 | 23678 | CG8385 | 23082 | CG8509 | 28915 |
| CG7988 | 22651 | CG8129 | 46959 | CG8269 | 23728 | CG8386 | 35919 | CG8520 | 36546 |
| CG7989 | 28172 | CG8129 | 24201 | CG8270 | 23755 | CG8390 | 46229 | CG8522 | 37640 |
| CG7990 | 46157 | CG8132 | 17254 | CG8271 | 4607 | CG8392 | 35923 | CG8523 | 51165 |
| CG7993 | 35314 | CG8134 | 24204 | CG8272 | 24262 | CG8394 | 45917 | CG8525 | 28916 |
| CG7995 | 22652 | CG8135 | 42635 | CG8273 | 28887 | CG8395 | 24305 | CG8527 | 39580 |
| CG7997 | 16840 | CG8138 | 24208 | CG8274 | 24265 | CG8396 | 47383 | CG8529 | 44360 |
| CG7998 | 22654 | CG8142 | 10881 | CG8276 | 28888 | CG8400 | 35419 | CG8531 | 24122 |
| CG7999 | 15878 | CG8144 | 24214 | CG8277 | 24267 | CG8401 | 45237 | CG8532 | 35949 |
| CG8001 | 35317 | CG8146 | 48210 | CG8280 | 49890 | CG8402 | 24308 | CG8534 | 49893 |
| CG8003 | 22659 | CG8146 | 23126 | CG8280 | 24270 | CG8403 | 42478 | CG8536 | 4867 |
| CG8005 | 22664 | CG8147 | 2892 | CG8282 | 24275 | CG8404 | 45482 | CG8538 | 35952 |
| CG8007 | 11462 | CG8149 | 24215 | CG8284 | 35872 | CG8405 | 11127 | CG8542 | 24125 |
| CG8008 | 4158 | CG8151 | 12581 | CG8285 | 4365 | CG8407 | 23504 | CG8545 | 35954 |
| CG8009 | 41105 | CG8152 | 13978 | CG8286 | 14154 | CG8408 | 12432 | CG8546 | 5110 |
| CG8009 | 48955 | CG8153 | 15695 | CG8287 | 28092 | CG8409 | 31995 | CG8548 | 28920 |
| CG8013 | 42423 | CG8155 | 24218 | CG8288 | 24278 | CG8411 | 28897 | CG8549 | 28924 |
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| CG8019 | 41022 | CG8166 | 8137 | CG8290 | 12739 | CG8415 | 50956 | CG8553 | 35959 |
| CG8020 | 44076 | CG8167 | 44761 | CG8293 | 2972 | CG8415 | 35421 | CG8556 | 28926 |
| CG8021 | 23675 | CG8169 | 39193 | CG8297 | 8972 | CG8416 | 12734 | CG8556 | 50349 |
| CG8023 | 34210 | CG8171 | 23131 | CG8297 | 46760 | CG8417 | 49508 | CG8557 | 28927 |
| CG8025 | 26039 | CG8173 | 35845 | CG8298 | 43541 | CG8418 | 35929 | CG8561 | 44361 |
| CG8026 | 4163 | CG8174 | 26933 | CG8300 | 24291 | CG8419 | 24097 | CG8566 | 23464 |
| CG8029 | 48016 | CG8177 | 39492 | CG8302 | 51943 | CG8421 | 52589 | CG8566 | 47171 |
| CG8031 | 35326 | CG8184 | 26935 | CG8303 | 4917 | CG8425 | 44049 | CG8567 | 39592 |
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| CG8038 | 28179 | CG8189 | 14210 | CG8311 | 8985 | CG8428 | 3229 | CG8571 | 35967 |
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| CG8043 | 28181 | CG8194 | 13018 | CG8315 | 28892 | CG8432 | 28866 | CG8578 | 35969 |
| CG8048 | 46563 | CG8197 | 42125 | CG8318 | 35877 | CG8433 | 49808 | CG8580 | 24130 |
| CG8049 | 22675 | CG8199 | 24231 | CG8320 | 8797 | CG8433 | 4902 | CG8581 | 16923 |
| CG8053 | 26022 | CG8200 | 42130 | CG8321 | 8765 | CG8434 | 43898 | CG8581 | 24475 |
| CG8058 | 13314 | CG8202 | 35410 | CG8322 | 30280 | CG8434 | 42570 | CG8581 | 29909 |
| CG8060 | 46991 | CG8203 | 35855 | CG8323 | 4861 | CG8434 | 4319 | CG8582 | 35970 |
| CG8060 | 22684 | CG8207 | 24236 | CG8325 | 35881 | CG8435 | 28900 | CG8583 | 33282 |
| CG8064 | 28182 | CG8208 | 9261 | CG8326 | 23760 | CG8440 | 6216 | CG8584 | 26988 |
| CG8064 | 49076 | CG8209 | 35858 | CG8327 | 35883 | CG8442 | 44439 | CG8587 | 26989 |
| CG8067 | 47871 | CG8211 | 24239 | CG8330 | 23763 | CG8443 | 42136 | CG8589 | 24180 |
| CG8067 | 22687 | CG8211 | 24237 | CG8331 | 35377 | CG8444 | 5830 | CG8590 | 35975 |
| CG8068 | 30709 | CG8212 | 44731 | CG8332 | 35415 | CG8445 | 47743 | CG8591 | 30713 |
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| CG8073 | 23020 | CG8219 | 24244 | CG8335 | 15506 | CG8449 | 24102 | CG8595 | 6541 |
| CG8075 | 7376 | CG8222 | 13503 | CG8336 | 23729 | CG8453 | 4615 | CG8595 | 39176 |
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| CG8086 | 23028 | CG8224 | 3825 | CG8340 | 35890 | CG8461 | 24103 | CG8601 | 28932 |
| CG8090 | 30341 | CG8224 | 853 | CG8343 | 7735 | CG8464 | 24104 | CG8603 | 47148 |
| CG8091 | 23033 | CG8226 | 8747 | CG8344 | 15692 | CG8465 | 24107 | CG8603 | 26992 |
| CG8092 | 28196 | CG8230 | 37160 | CG8349 | 48682 | CG8468 | 6452 | CG8604 | 9264 |
| CG8093 | 19561 | CG8231 | 23751 | CG8351 | 28895 | CG8470 | 40712 | CG8605 | 29435 |
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| CG8104 | 29788 | CG8243 | 26952 | CG8361 | 16753 | CG8485 | 35940 | CG8616 | 38249 |
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| CG8107 | 46241 | CG8244 | 30642 | CG8363 | 35904 | CG8487 | 42140 | CG8625 | 6208 |
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| CG8108 | 35343 | CG8251 | 24257 | CG8368 | 45259 | CG8492 | 14929 | CG8628 | 35392 |
| CG8110 | 35345 | CG8253 | 35411 | CG8370 | 42509 | CG8493 | 24109 | CG8629 | 39155 |


| CG8630 | 33340 | CG8768 | 36022 | CG8893 | 23645 | CG9012 | 23666 | CG9139 | 46329 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| CG8630 | 50290 | CG8772 | 7192 | CG8895 | 7866 | CG9013 | 4964 | CG9140 | 43184 |
| CG8631 | 2998 | CG8773 | 10203 | CG8895 | 33919 | CG9014 | 36084 | CG9143 | 46330 |
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| CG8636 | 28937 | CG8776 | 40803 | CG8896 | 44386 | CG9022 | 45173 | CG9147 | 29050 |
| CG8637 | 35988 | CG8778 | 23621 | CG8900 | 23083 | CG9023 | 51936 | CG9148 | 45224 |
| CG8639 | 29968 | CG8779 | 979 | CG8902 | 23650 | CG9025 | 44428 | CG9150 | 16877 |
| CG8641 | 35993 | CG8779 | 30073 | CG8905 | 42162 | CG9027 | 37794 | CG9151 | 47881 |
| CG8641 | 52260 | CG8779 | 37282 | CG8907 | 28987 | CG9031 | 42189 | CG9151 | 9827 |
| CG8642 | 35997 | CG8781 | 36025 | CG8909 | 29900 | CG9032 | 23685 | CG9153 | 37221 |
| CG8645 | 35431 | CG8782 | 28950 | CG8912 | 28989 | CG9032 | 50958 | CG9154 | 29054 |
| CG8648 | 15698 | CG8783 | 40721 | CG8914 | 26915 | CG9033 | 44287 | CG9154 | 49534 |
| CG8649 | 47514 | CG8784 | 15989 | CG8915 | 28857 | CG9035 | 8759 | CG9155 | 49345 |
| CG8649 | 6276 | CG8785 | 4650 | CG8916 | 9138 | CG9038 | 29012 | CG9156 | 29057 |
| CG8651 | 37715 | CG8786 | 36028 | CG8918 | 28994 | CG9041 | 28163 | CG9159 | 7893 |
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| CG8655 | 40715 | CG8795 | 1768 | CG8922 | 36060 | CG9045 | 37711 | CG9163 | 45927 |
| CG8656 | 24184 | CG8798 | 36035 | CG8923 | 36063 | CG9046 | 13230 | CG9163 | 30075 |
| CG8657 | 4659 | CG8799 | 39539 | CG8928 | 23480 | CG9047 | 23153 | CG9166 | 28109 |
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| CG8669 | 2935 | CG8806 | 40723 | CG8937 | 45596 | CG9062 | 3810 | CG9176 | 40964 |
| CG8675 | 26997 | CG8808 | 37966 | CG8938 | 50140 | CG9063 | 40738 | CG9177 | 29070 |
| CG8676 | 37694 | CG8809 | 39076 | CG8938 | 23084 | CG9064 | 2647 | CG9181 | 37437 |
| CG8677 | 23608 | CG8811 | 29774 | CG8942 | 9976 | CG9065 | 33879 | CG9184 | 10283 |
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| CG8681 | 1479 | CG8816 | 36045 | CG8948 | 42165 | CG9071 | 4062 | CG9198 | 29072 |
| CG8690 | 15798 | CG8817 | 13081 | CG8949 | 48307 | CG9075 | 42201 | CG9200 | 36092 |
| CG8693 | 7947 | CG8819 | 49637 | CG8950 | 36069 | CG9081 | 38218 | CG9201 | 29073 |
| CG8694 | 28292 | CG8821 | 37660 | CG8954 | 23659 | CG9084 | 45609 | CG9203 | 29075 |
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| CG8696 | 15789 | CG8825 | 46268 | CG8958 | 42169 | CG9088 | 42203 | CG9206 | 3785 |
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| CG8706 | 8397 | CG8827 | 41219 | CG8962 | 29003 | CG9090 | 44297 | CG9209 | 44638 |
| CG8706 | 39215 | CG8830 | 28960 | CG8963 | 42110 | CG9092 | 51445 | CG9210 | 11547 |
| CG8706 | 3710 | CG8831 | 42153 | CG8967 | 30834 | CG9095 | 23159 | CG9211 | 29898 |
| CG8707 | 36003 | CG8833 | 36408 | CG8967 | 42565 | CG9096 | 29023 | CG9211 | 42577 |
| CG8708 | 45194 | CG8839 | 4620 | CG8968 | 48894 | CG9098 | 27001 | CG9212 | 28116 |
| CG8709 | 36007 | CG8841 | 48253 | CG8968 | 26923 | CG9099 | 28106 | CG9214 | 42209 |
| CG8711 | 44829 | CG8841 | 23625 | CG8969 | 30483 | CG9099 | 49895 | CG9218 | 28119 |
| CG8714 | 9951 | CG8843 | 28873 | CG8972 | 45845 | CG9100 | 27002 | CG9219 | 35750 |
| CG8717 | 8739 | CG8844 | 35437 | CG8974 | 5572 | CG9102 | 8943 | CG9220 | 29085 |
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| CG8726 | 40719 | CG8858 | 23634 | CG8980 | 42175 | CG9113 | 3275 | CG9238 | 24149 |
| CG8727 | 11765 | CG8860 | 8768 | CG8981 | 28098 | CG9115 | 29032 | CG9240 | 14833 |
| CG8728 | 48677 | CG8862 | 38085 | CG8983 | 51675 | CG9116 | 50537 | CG9242 | 24152 |
| CG8728 | 23617 | CG8863 | 23637 | CG8987 | 3133 | CG9116 | 38139 | CG9243 | 37422 |
| CG8729 | 15533 | CG8865 | 23639 | CG8988 | 4601 | CG9117 | 52545 | CG9244 | 12455 |
| CG8730 | 23772 | CG8873 | 45618 | CG8989 | 12771 | CG9118 | 49813 | CG9245 | 11852 |
| CG8732 | 3222 | CG8874 | 36053 | CG8993 | 41126 | CG9118 | 14931 | CG9246 | 24136 |
| CG8733 | 51921 | CG8877 | 18567 | CG8995 | 23665 | CG9119 | 46326 | CG9247 | 52612 |
| CG8734 | 7949 | CG8881 | 28975 | CG8996 | 44378 | CG9120 | 49896 | CG9248 | 41226 |
| CG8735 | 4025 | CG8882 | 28976 | CG8998 | 28102 | CG9124 | 36086 | CG9249 | 47643 |
| CG8739 | 47750 | CG8884 | 35445 | CG9000 | 37179 | CG9126 | 47073 | CG9250 | 48548 |
| CG8743 | 45989 | CG8886 | 46702 | CG9001 | 4931 | CG9126 | 47074 | CG9250 | 36095 |
| CG8757 | 13110 | CG8887 | 28982 | CG9002 | 49812 | CG9127 | 47972 | CG9257 | 6406 |
| CG8759 | 36017 | CG8888 | 30336 | CG9003 | 23481 | CG9128 | 37216 | CG9258 | 46542 |
| CG8760 | 28945 | CG8890 | 24148 | CG9004 | 40727 | CG9131 | 44362 | CG9265 | 46577 |
| CG8764 | 35829 | CG8891 | 36055 | CG9005 | 36079 | CG9134 | 48756 | CG9267 | 2879 |
| CG8766 | 4658 | CG8892 | 28985 | CG9009 | 12016 | CG9135 | 36091 | CG9270 | 29961 |
| CG8767 | 36533 | CG8893 | 50351 | CG9010 | 40728 | CG9138 | 1047 | CG9271 | 15547 |


| CG9272 | 41018 | CG9414 | 30479 | CG9540 | 14807 | CG9699 | 7742 | CG9879 | 29311 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| CG9283 | 15223 | CG9415 | 15347 | CG9542 | 45620 | CG9701 | 3358 | CG9881 | 28141 |
| CG9286 | 23735 | CG9416 | 10064 | CG9548 | 35453 | CG9702 | 6859 | CG9882 | 29312 |
| CG9288 | 46191 | CG9418 | 37665 | CG9550 | 43996 | CG9703 | 6137 | CG9886 | 48079 |
| CG9290 | 14349 | CG9422 | 30171 | CG9554 | 43911 | CG9705 | 40665 | CG9886 | 29320 |
| CG9291 | 15302 | CG9423 | 36103 | CG9556 | 48044 | CG9706 | 49347 | CG9890 | 23062 |
| CG9294 | 29092 | CG9426 | 10843 | CG9564 | 45717 | CG9709 | 29119 | CG9895 | 41035 |
| CG9296 | 29096 | CG9427 | 15375 | CG9565 | 37803 | CG9712 | 23944 | CG9899 | 46584 |
| CG9300 | 24139 | CG9428 | 3986 | CG9569 | 1820 | CG9717 | 42669 | CG9900 | 24171 |
| CG9302 | 15544 | CG9429 | 51272 | CG9569 | 1820 | CG9722 | 6679 | CG9901 | 29944 |
| CG9304 | 11142 | CG9430 | 7339 | CG9571 | 10480 | CG9723 | 37412 | CG9903 | 42690 |
| CG9305 | 30523 | CG9433 | 41021 | CG9573 | 49820 | CG9725 | 23945 | CG9904 | 45478 |
| CG9306 | 23088 | CG9438 | 37148 | CG9576 | 47261 | CG9726 | 41347 | CG9906 | 5597 |
| CG9307 | 23163 | CG9441 | 33923 | CG9577 | 24064 | CG9727 | 30575 | CG9907 | 6131 |
| CG9308 | 6606 | CG9443 | 5843 | CG9578 | 45082 | CG9728 | 42895 | CG9908 | 7001 |
| CG9310 | 12692 | CG9444 | 43275 | CG9580 | 50546 | CG9730 | 36139 | CG9910 | 24175 |
| CG9311 | 14173 | CG9448 | 24030 | CG9581 | 48220 | CG9732 | 27035 | CG9910 | 24175 |
| CG9313 | 29099 | CG9450 | 24031 | CG9581 | 39208 | CG9734 | 23483 | CG9911 | 46585 |
| CG9314 | 44647 | CG9451 | 14344 | CG9582 | 2845 | CG9735 | 23951 | CG9913 | 36459 |
| CG9320 | 24141 | CG9452 | 51202 | CG9586 | 28250 | CG9738 | 26928 | CG9914 | 29322 |
| CG9322 | 29100 | CG9453 | 47262 | CG9588 | 47763 | CG9739 | 44390 | CG9916 | 41015 |
| CG9323 | 44984 | CG9455 | 13263 | CG9590 | 29482 | CG9741 | 51061 | CG9920 | 29326 |
| CG9325 | 29101 | CG9456 | 37955 | CG9591 | 44696 | CG9742 | 39256 | CG9921 | 14921 |
| CG9326 | 24157 | CG9458 | 48700 | CG9593 | 24165 | CG9747 | 1394 | CG9922 | 35465 |
| CG9328 | 28125 | CG9459 | 48905 | CG9594 | 13636 | CG9748 | 6299 | CG9924 | 28798 |
| CG9331 | 44653 | CG9459 | 5948 | CG9595 | 24068 | CG9749 | 36142 | CG9925 | 29328 |
| CG9333 | 52551 | CG9460 | 24036 | CG9596 | 27025 | CG9750 | 19021 | CG9927 | 29332 |
| CG9334 | 49899 | CG9461 | 24039 | CG9597 | 36117 | CG9752 | 50282 | CG9930 | 47793 |
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| CG9342 | 15775 | CG9463 | 15587 | CG9601 | 24070 | CG9753 | 1385 | CG9938 | 29337 |
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| CG9344 | 23689 | CG9466 | 46288 | CG9603 | 37496 | CG9761 | 23171 | CG9941 | 29902 |
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| CG9346 | 27013 | CG9467 | 45806 | CG9610 | 48121 | CG9764 | 28674 | CG9943 | 48887 |
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| CG9350 | 30619 | CG9469 | 15469 | CG9613 | 5801 | CG9772 | 15636 | CG9945 | 23742 |
| CG9351 | 24143 | CG9472 | 8424 | CG9615 | 24072 | CG9774 | 3793 | CG9946 | 7799 |
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| CG9359 | 24144 | CG9484 | 44676 | CG9623 | 5600 | CG9783 | 29276 | CG9952 | 29903 |
| CG9360 | 13189 | CG9485 | 45809 | CG9629 | 44700 | CG9784 | 30098 | CG9953 | 9024 |
| CG9361 | 8564 | CG9488 | 29720 | CG9630 | 31081 | CG9786 | 11775 | CG9954 | 12712 |
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| CG9363 | 37012 | CG9491 | 27015 | CG9636 | 28133 | CG9796 | 36452 | CG9958 | 28145 |
| CG9364 | 30730 | CG9493 | 40743 | CG9637 | 9073 | CG9802 | 39207 | CG9961 | 36175 |
| CG9373 | 44658 | CG9494 | 44877 | CG9638 | 24076 | CG9804 | 46579 | CG9968 | 36185 |
| CG9376 | 9457 | CG9495 | 3795 | CG9643 | 24081 | CG9805 | 28140 | CG9968 | 29693 |
| CG9377 | 42837 | CG9496 | 2824 | CG9646 | 14982 | CG9811 | 30103 | CG9973 | 36187 |
| CG9378 | 28130 | CG9499 | 7900 | CG9648 | 15351 | CG9818 | 29285 | CG9973 | 36584 |
| CG9379 | 22824 | CG9501 | 7903 | CG9650 | 23170 | CG9819 | 30105 | CG9976 | 38002 |
| CG9381 | 44662 | CG9508 | 24052 | CG9655 | 37309 | CG9828 | 29289 | CG9977 | 49573 |
| CG9383 | 23737 | CG9510 | 44683 | CG9657 | 43922 | CG9834 | 29290 | CG9977 | 36193 |
| CG9384 | 14169 | CG9512 | 14809 | CG9660 | 24083 | CG9836 | 29295 | CG9981 | 11566 |
| CG9386 | 47755 | CG9514 | 37403 | CG9662 | 7278 | CG9839 | 36455 | CG9983 | 29523 |
| CG9388 | 24017 | CG9517 | 24162 | CG9662 | 7278 | CG9847 | 12863 | CG9984 | 42217 |
| CG9389 | 44663 | CG9518 | 8328 | CG9666 | 45658 | CG9849 | 12850 | CG9985 | 6229 |
| CG9391 | 23723 | CG9519 | 16501 | CG9667 | 36127 | CG9852 | 48717 | CG9986 | 46113 |
| CG9393 | 44400 | CG9519 | 47195 | CG9668 | 46919 | CG9854 | 42283 | CG9987 | 36198 |
| CG9394 | 13879 | CG9520 | 2826 | CG9670 | 24086 | CG9855 | 33309 | CG9987 | 36494 |
| CG9398 | 29110 | CG9521 | 16497 | CG9674 | 24089 | CG9862 | 29302 | CG9994 | 36201 |
| CG9399 | 13788 | CG9521 | 47136 | CG9677 | 27032 | CG9865 | 40701 | CG9994 | 43486 |
| CG9400 | 43296 | CG9522 | 19861 | CG9678 | 23342 | CG9867 | 44570 | CG9995 | 29531 |
| CG9401 | 28132 | CG9526 | 51451 | CG9680 | 36131 | CG9868 | 45506 | CG9995 | 36205 |
| CG9406 | 48893 | CG9527 | 24054 | CG9682 | 16549 | CG9870 | 44215 | CG9996 | 36207 |
| CG9406 | 29765 | CG9528 | 44687 | CG9683 | 22830 | CG9873 | 29760 | CG9998 | 24177 |
| CG9410 | 44669 | CG9533 | 5569 | CG9688 | 43887 | CG9876 | 10481 | CG9999 | 30568 |
| CG9412 | 29113 | CG9536 | 7907 | CG9695 | 13005 | CG9878 | 23705 |  |  |
| CG9413 | 45180 | CG9537 | 29374 | CG9696 | 7787 | CG9878 | 50446 |  |  |

